

# Agilent G1978B Multimode Source for 6500 Series Q-TOF LC/MS

# Set-Up Guide



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# WARNING

A WARNING notice denotes a hazard. It calls attention to an operating procedure, practice, or the like that, if not correctly performed or adhered to, could result in personal injury or death. Do not proceed beyond a WARNING notice until the indicated conditions are fully understood and met.

# In This Guide

This guide explains how to install, maintain and troubleshoot your multimode ion source.

#### **1** Installation

This chapter tells you how to install the multimode source.

#### 2 Set-Up

This chapter describes basic operation and maintenance for the multimode source.

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This chapter contains instructions to install the multimode source on a 6510 Series Q-TOF LC/MS system, and also to remove and replace the source.



# Step 1. Prepare to install

The Multimode Enablement Kit, G1978-60451, is shipped with the multimode source. This kit needs to be installed before the multimode source is used.

Note that the multimode source and its accessories are to be installed by an Agilent Customer Engineer.

- 1 Check that the Multimode Enablement Kit contains the following parts:
  - Multimode Bd HV Cable, p/n G1960-60858
  - Multimode HV PCA, p/n G1960-61015
  - Multimode Bd Power/Data Cable, p/n G1960-60873







**2** Install the APCI Enablement Kit, G1947-60451, which is shipped with the multimode source.

The APCI Enablement kit contains the following parts:

- Fast APCI HV Supply, p/n G1946-80058
- Valve BD-APCI Supply Cable, p/nG1960-60802
- Valve BD-APCI Needle Interlock Cable, p/n G1960-60856







Figure 2 From left to right: G1946-80058, G1960-60802 and G1960-60856

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# Step 2. Install the HV control PCA and cables

**1** Turn off the system power and remove the system power cord.

The power cord should be kept intact if the vacuum control switch box is used. The switch box is intended to keep the vacuum on while a service engineer works on the electronics. The switch box is for service engineer use only.

- **2** Remove the CDS cover, top, side, front, and the Aux Module cover.
- **3** Disconnect the ribbon cable that connects the valve PCA to the Vcap/Vchamber power supply. Then disconnect the Vcap and Vchamber cable from the power supply.







- **4** Place the multimode HV power supply PCA in the slot between the valve PCA and the Vcap/Vchamber power supply. Secure the board by pressing it down into its slot and then attach it with two screws.
- **5** Connect the short gray cable from the valve PCA to the multimode HV power supply.



Figure 4 Connecting the valve PCA to the multimode HV power supply.

Step 2. Install the HV control PCA and cables

- **6** Install the APCI HV power supply. The APCI HV power supply is located at the end of the AUX Module.
- 7 Connect ribbon cable between the valve PCA and Vcap/Vchamber power supply.



Figure 5 Connecting the valve PCA to the Vcap/Vchamber power supply.

8 Connect the Vcap and Vchamber cables to the Vcap/Vchamber power supply.



**Figure 6** Connecting the Vcap and Vchamber cables to the power supply.

**9** Connect the long ribbon cable, p/n G1960-60802, from the APCI HV power supply to the valve PCA.

Step 2. Install the HV control PCA and cables



Figure 7 Connecting the APCI HV power supply to the valve PCA.

**10** Insert one end of the APCI Needle Interlock cable, G1960-60856, through the slot at the front of the system and then plug it to the APCI HV connector. Attach the other end to the chassis with the o-ring and the nut (see Figure 8).





Figure 8 Connecting the APCI HV to the chassis.

**11** Insert the cable, G1960-60858, to the top slot and attach it to the chassis. Plug the other two ends into the multimode HV PCA.





Figure 9 Connecting the HV PCA to the chassis.

**12** Close the AUX Module cover and reconnect all cables.

13 Install the multimode source onto the system and connect all connectors.

**Step 2. Install the HV control PCA and cables** 



Figure 10 Installing the multimode source (left) and connecting all connectors.

- 14 Put back the side, top, front and CDS cover.
- **15** Plug the system power cord back on and turn the front switch on.

The pump down process will start.

- **16** Start the MassHunter Workstation program and verify that the software recognizes the source.
- **17** Set the **Context** view to **Tune**, and in **Manual Tune**, verify that the system can generate the proper tune peaks.

1

# To remove the multimode source

Do the following steps to remove the multimode source.

- **1** Turn off the multimode source temperatures and flows:
  - **a** Change the **Context** view to **Acquisition**.
  - **b** Click the **MS Q-TOF** tab.
  - c Turn off all voltages and temperatures in the Source tab.
  - **d** Wait approximately 20 minutes for the source to cool down.

**WARNING** Do not touch the multimode source or the capillary cap. They may be very hot. Let the parts cool before you handle them.

# WARNING Never touch the source surfaces, especially when you analyze toxic substances or when you use toxic solvents. The source has several sharp pieces which can pierce your skin including the APCI corona needle, vaporizer sensor and counter current electrode.

# WARNING

Do not insert fingers or tools through the openings on the multimode chamber. When in use, the capillary and capillary cap are at high voltages up to 4 kV.

- 2 Wait approximately 20 minutes or until the source is cool.
- **3** Open the CDS door at the front of the MS to access the cables.
- 4 Disconnect the ESI high voltage charging electrode cable.
- **5** Disconnect the APCI Needle Interlock, and multimode HV cable.
- **6** Unscrew the nebulizer gas line from the nebulizer.
- 7 Unscrew the LC sample tubing from the nebulizer.
- **8** Open the latch on the source and open the source.
- **9** Remove the multimode source from the spray chamber mount.
- **10** Place the source shipping cover on the source.

To convert from multimode to ESI or APCI

# To convert from multimode to ESI or APCI

# WARNING

Never touch the source surfaces, especially when you analyze toxic substances or when you use toxic solvents. The source has several sharp pieces which can pierce your skin including the APCI corona needle, vaporizer sensor and counter current electrode.

- **1** Unscrew and remove the multimode spray shield with the field shaping electrodes.
- **2** Install the new source and the standard spray shield, making sure that the hole in the spray shield is in the 12 o'clock position.
- **3** For an APCI ion source, connect the vaporizer heater cable and the APCI high voltage cable.
- **4** For all sources, reconnect the nebulizer gas line tubing and the LC/MS sample tubing.

To convert from ESI or APCI to the multimode source

# To convert from ESI or APCI to the multimode source

# CAUTION

If you are installing this source on this instrument for the first time, follow the steps in "Installation" on page 7.

- **1** Turn off the multimode source temperatures and flows:
  - a Change the Context view to Acquisition.
  - **b** Click the **MS Q-TOF** tab.
  - c Turn off all voltages and temperatures in the Source tab.
  - **d** Wait approximately 20 minutes for the source to cool down.
- **2** Wait for the source to cool (until temperatures are at least below 100°C).
- **3** Disconnect the nebulizer gas tubing from the currently installed ion source.
- 4 Disconnect the LC/MS sample inlet tubing.
- **5** If the APCI source is installed, remove the APCI vaporizer heater cable and APCI high voltage cable.
- **6** Remove the currently installed ion source.
- 7 Unscrew and remove the spray shield. See Figure 11.

# WARNING

Do not touch the multimode source or the capillary cap. They may be very hot. Let the parts cool before you handle them.

# WARNING

Do not insert fingers or tools through the openings on the multimode chamber. When in use, the capillary and capillary cap are at high voltages up to 4 kV.

To convert from ESI or APCI to the multimode source



Figure 11 Standard spray shield and capillary cap for ESI or APCI

8 Remove the capillary cap. If needed, moisten a clean cloth with isopropyl alcohol and wipe the capillary cap. See Figure 12.



Capillary cap

**Figure 12** Spray shield removed.

**9** Place the capillary cap back on the capillary.

**10** Install the new spray shield with field shaping electrodes. See Figure 13.

To convert from ESI or APCI to the multimode source



Figure 13 Multimode spray shield

**11** Screw the multimode spray shield into the holder for the spray shield. See Figure 14.



Figure 14Multimode spray shield installed

NOTE

The field shaping electrodes should be in the nine o'clock and the six o'clock position. Loosen the end plate screws on each side to adjust the field shaping electrodes position.

12 Remove the shipping cover from the multimode source spray chamber.

To convert from ESI or APCI to the multimode source



Figure 15 Multimode Spray Chamber

**13** Install the spray chamber on the spray chamber mount.



Figure 16 Multimode source with I-Button

**14** Install the nebulizer on the multimode source spray chamber.

To convert from ESI or APCI to the multimode source



Figure 17 No nebulizer on top of the multimode source

**15** Connect the 1/8-inch nebulizer gas tubing from the LC/MS mainframe to the nebulizer gas fitting. See Figure 18.



Figure 18 Nebulizer with gas tubing connected

To convert from ESI or APCI to the multimode source

16 Connect the LC/MS sample tubing to the LC/MS diverter valve inlet filter.

WARNING The LC/MS Liquid Chromatograph diverter valve is an integral part of the G1978B safety system. The LC mobile phase flow must always be connected to the diverter valve inlet filter. Never bypass the diverter valve and connect directly to the nebulizer. If the diverter valve is used in a manner not specified by Agilent Technologies, the protections provided by the diverter valve may be impaired.

**17** If you are installing the multimode source for the first time, follow the steps in "Step 2. Install the HV control PCA and cables" on page 9.



This chapter describes the tasks that you need to operate and maintain the multimode source.





#### 2 Set-Up

To set up a method to use the multimode source

# To set up a method to use the multimode source

# WARNING

The LC/MS diverter valve is an integral part of the G1978B safety system. The LC mobile phase flow must always be connected to the diverter valve inlet filter. Never bypass the diverter valve and connect directly to the nebulizer. If the diverter valve is used in a manner not specified by Agilent Technologies, the protections provided by the diverter valve may be impaired and the system may catch fire.

- **1** In the MassHunter software, change the **Context** to **Acquisition**.
- **2** In the MS Q-TOF tab, set **Ion source** to **Multimode** (see Figure 19 on page 23).
- **3** In the **Sources** tab, choose an ionization mode from the **Ion Modes (Seg)** list. You may set the ionization mode to one of the following:
  - ESI
  - APCI
  - Mixed

The Ion Mode selection Mixed will specify a method for simultaneous ESI and APCI operation.

Note that the Ionization Modes selection is only visible if **Ion source** is set to **Multimode**.

- **4** In the **Source** tab, set the desired source conditions. See "Guidelines" in the *Agilent G1978A/B Multimode Source Maintenance Guide* for suggested source conditions for the multimode source for the different ionization modes.
- **5** Make any other changes that are necessary for your method.
- **6** Save the method.

#### Set-Up 2

To set up a method to use the multimode source

Multimode (Se	eg)					MS TOF (Expt)
Gas Temp	325	°С		324	°C	Fragmentor 175 V
Vaporizer	200	°C		198	°C	Skimmer 65 V
Drying Gas	5	1/min		5.0	1/min	
Nebulizer	30	psig		30	psig	UCITREVpp 750 V
Multimode (E)	(pt)					Ionization Modes (seg)
VCap	2000	۷	Capillary	0.141	uА	Mixed 👻
Corona+	4	uА	Corona	110	۷	
			Chamber	3.61	uА	Charging Voltage 2000 V

Figure 19 Multimode acquisition settings

To open the multimode source

# To open the multimode source

Open the multimode source to access the end cap and the capillary cap for cleaning and inspection.

# WARNING

Do not touch the multimode source or the capillary cap. They may be very hot. Let the parts cool before you handle them.

# WARNING

Never touch the source surfaces, especially when you analyze toxic substances or when you use toxic solvents. The source has several sharp pieces which can pierce your skin including the APCI corona needle, vaporizer sensor and counter current electrode.

# **WARNING** Do not insert fingers or tools through the openings on the multimode chamber. When in use, the capillary and capillary cap are at high voltages up to 4 kV.

- **1** Turn off the multimode source temperatures and flows:
  - a Change the **Context** view to **Acquisition**.
  - **b** Click the **MS Q-TOF** tab.
  - **c** Put the instrument in Standby mode.
  - **d** Wait approximately 20 minutes for the source to cool down.
- **2** Open the spray chamber cover by pulling the latch.

The high voltage automatically turns off when the chamber door is opened so that no high voltages are present within the chamber.

- **3** Check that the vaporizer temperature sensor is straight and extends 15 mm from back of chamber.
- **4** Check that the separator is aligned vertically.
- **5** Check that the APCI corona needle is in and extends approximately 3 mm from the corona guide.
- **6** Check that the source is clean.

2

# To check tuning with the multimode source

Autotune is currently only available for the G3251B Dual Electrospray source. However, mass calibrations and manual optimization of mass resolution can be done using the G1978B source. To calibrate mass accuracy, do these steps.

- 1 Run an Autotune with the G3251B Dual Electrospray source installed.
- **2** Remove the G3251B Dual Electrospray source and install the G1978B multimode source.
- **3** Uninstall the Electrospray Calibrant Bottle B from the instrument. Cap the calibrant bottle with one of the supplied bottle caps (p/n 9300-2575).
- **4** Rinse one of the extra calibrant bottles (p/n 9300-2576) that was supplied as part of the Q-TOF Shipping Kit (p/n G2581-60170) with high purity acetonitrile. Pour the contents of the MMI-L Low Concentration Tuning Mix (p/n G1969-85020) into the rinsed calibrant bottle. Install the calibrant bottle on the Q-TOF mainframe in the bottle B location.
- 5 Set the Context view to Tune in the MassHunter Workstation program.
  - **a** Load the most recently used autotune file. Change the source type Multimode.
  - **b** Click the **Mass TOF Calibration** tab and do a mass calibration.
  - **c** Adjust the lens voltages and other tune parameters as required to optimize the mass resolution of the instrument. If changes are made to the Mid Mirror, a mass calibration will have to be done again.
  - **d** Verify that you have sufficient abundance for the tune peaks, that the tune peak at 2122 has greater than 10,000 resolution, and that all mass assignments are with 2 ppm after a mass calibration has been done.
- **6** Save the tune file and close the tune context.

# 2 Set-Up

To check tuning with the multimode source



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# **Installation Verification**

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In this chapter, you create and run methods to check out the system.



# Step 1. Auto tune

This step applies to MassHunter Workstation Software - Acquisition for TOF/Q-TOF revision B.01.03 or higher.

- Run autotune with the G1969-85000 ESI-L Low Concentration Tuning Mix. There are no tune specific methods.
  - Tune the 6220 in 2GHz extended dynamic range for both positive and negative.
  - Tune the 6210 in Standard (3200 m/z) mode 1GHz.

# Step 2. Set up method names and parameters

- 1 Create six methods from Default.m for the multimode ESI + APCI LC Demo Sample (p/n G1978-85000), using these method names:
  - MMCHECKTOF\_EI\_POS.m
  - MMCHECKTOF\_EI\_NEG.m
  - MMCHECKTOF\_CI\_POS.m
  - MMCHECKTOF\_CI\_NEG.m
  - MMCHECKTOF\_MX\_EI POS\_CI POS.m
  - MMCHECKTOF\_MX\_EI NEG\_CI NEG.m
- **2** Use these parameters for each method:

#### Table 1

Parameter/Tab	Value
Column	Cartridge Hardware, Rapid Resolution, (p/n 820555-901)
	SB-C18 Rapid Res 3.5um,2.1x30mm, (p/n 873700-902)
Sample Tab	Name: MM Demo Sample
	Position 1
	Run Type: Standard
	Acquisition only
	Path D:\PE Sciex Data\Projects\Data
ALS Tab	Standard Injection 1µL
	Bin Pump Tab:
	Flow .4mL/min
	Stop time: 3 min
	Solvent A 100.0 % (65%MeOH:35%H <sub>2</sub> O + 0.2%acetic acid)
	Run time same as pump
Data files (data files	Data File: MM_ESI_POS.wiff
for B.01.03 or greater	Data File: MM_ESI_NEG.wiff
use the suffix .d)	Data File: MM_APCI_POS.wiff
	Data File: MM_APCI_NEG.wiff
	Data File: MM_ESI_APCI_POS.wiff
	Data File: MM_ESI_APCI_NEG.wiff

# Step 3. Create MMCHECKTOF\_EI\_POS.m

The graphics in this topic differ slightly for MassHunter B.01.03 or higher. For B.01.03, access these tabs from the Acquisition view.

• Set the parameters for MMCHECKTOF\_EI\_POS.m:

	- MMI (Seq.)-	uisition	Hef. Ma	asses L	hromatogram   I	- MS TOF (S	ican]	Parameters	MMI
Ion Polarity (Seg.)	Gas Temp	350	с	300	c	Fragmentor	225	v	Source (Seg.)
Polarity Switch per scan	Vaporizer	200	С	200	с	Skimmer	60	- 1/	
Select Scan to Display	Drying Gas	5.0	1/min	3.4	1/min	- Craning	100	Y	C APCI
Time and Scan Segments	Nebulizer	60	psig	20	psig	OCT RF V	250	v	C ESI/APU
Add 0.00	MMI (Scan) VCap	1000	v	Capillary	μA	Chamber	0.00	μΑ	Charging Voltage
Del 0.00 1 Add	Corona +	4.0	μA	Corona	77 V				2000 V
Mod Del	-L								

Figure 20 Acquisition parameters

MMI  MMI Ion Polarity (Seg.)	Chromatogram Details	Segments	Chromatograms	Reset
Positive C Negative Polarity Switch per scan	Type EIC  Label Crystal Violet	All 0.00	dd -> 2 1-Hexanesulfonic acid 3 Carbozole 4 9-Phenanthrol	
Select Scan to Uisplay Time and Scan Segments Time (minutes) Scans	Extracted 372-372.3	Scans N	lodify	
Add 0.00	Y-axis range 1000000 counts	Scan 2 D Scan 2 D Scan 3	relete	
Mod	Add Chromatogram with index			



Step 3. Create MMCHECKTOF\_EI\_POS.m

Ionization Mode		MM-ES	
Polarity		Negative	
1100 Binary Pump 1			
Control			
Column flow		0.400 mL/min	
Stop Time		No Limit	
Post Time		Off	
Solvents			
Solvent A		100.0 % (65%MeOH:35%H	I <sub>2</sub> O + 0.2%acetic acid))
Solvent B		0.0 %	
Pressure Limits			
Minimum Pressure		0 bar	
Maximum Pressure		400 bar	
Spray Chamber			
[MSZones]			
Gas Temp	350 $^{\circ}$ C		Maximum 350 $^{\circ}$ C
Vaporizer	200 $^{\circ}$ C		Maximum 250 $^{\circ}$ C
Drying Gas	5.0 L/m	in	Maximum 13.0 L/min
Neb Pres	60 psig		Maximum 60 psig
VCap (Positive)	1000 V		
VCap (Negative)	1000 V		
VCharge (Positive)	2000 V		
VCharge (Negative)	2000 V		
Corona (Positive)	0.0 μΑ		
Corona (Negative)	0.0 μΑ		

# Step 4. Create MMCHECKTOF\_EI\_NEG.m

• Set the parameters for MMCHECKTOF\_EI\_NEG.m:

	Data Acq	uisition	Ref. Ma	asses   C	Chromatogran	n   T	une Calibr	ation   can)	Parameters	Diagnostics
on Polarity (Seg.) Positive ( Negative	Gas Temp	350	с	300	c		Fragmentor	225	v	Source (Seg.)
Polarity Switch per scan	Vaporizer Drying Gas	200	C I/min	200	C Vmin		Skimmer	60	v	C APCI
Time and Scan Segments	Nebulizer	60	psig	20	psig		OCT RF V	250	v	C ESI/APCI
Add 0.00	MMI (Scan) VCap	1000	v	Capillary	0.000	_μΑ	Chamber	0.00	μΑ	Charging Voltage
Del Add	Corona -	4.0	μA	Corona	77	v				12000 V



MMI	Chromatogram Details	Segments	ration Parameters	Chromatograms	Reset
Negative     Scan	Type EIC  Label 1-Hexanesulfonic acid	0.00	Add->	2 1-Hexanesultonic acid 3 Carbazole 4 9-Phenanthrol	
n Segments	Extracted 165-165.3	Scens	Modify		
i) Scens	Offset 15 % Y-axis range 1000000 counts	Scan 1 Scan 2 Scan 3	Delete		
Add	Add Chromatogram with index	Scan 4			

Figure 23 Chromatogram

Step 4. Create MMCHECKTOF\_EI\_NEG.m

Ionization Mode		MM-ES	
Polarity		Negative	
1100 Binary Pump 1			
Control			
Column flow		0.400 mL/min	
Stop Time		No Limit	
Post Time		Off	
Solvents			
Solvent A		100.0 % (65%MeOH:35%H	I <sub>2</sub> O + 0.2%acetic acid))
Solvent B		0.0 %	
Pressure Limits			
Minimum Pressure		0 bar	
Maximum Pressure		400 bar	
Spray Chamber			
[MSZones]			
Gas Temp	350 °C		Maximum 350 °C
Vaporizer	200 °C		Maximum 250 °C
Drying Gas	5.0 L/mi	'n	Maximum 13.0 L/min
Neb Pres	60 psig		Maximum 60 psig
VCap (Positive)	1000 V		
VCap (Negative)	1000 V		
VCharge (Positive)	2000 V		
VCharge (Negative)	2000 V		
Corona (Positive)	0.0 μΑ		
Corona (Negative)	0.0 μΑ		

# Step 5. Create MMCHECKTOF\_CI\_POS.m

• Set the parameters for MMCHECKTOF\_CI\_POS.m:

Ion Source	Data Acq	uisition	Ref. Ma	nsses   (	Chromatogram	) T	une ) Calibr	ation )	Parameters	Diagnostics
Ion Polarity (Seg.) Positive C Negative	Gas Temp	350	c	300	с		Fragmentor	225	v	Source (Seg.)
Polarity Switch per scan	Vaporizer Drying Gas	200	C Vmin	3.4	C I/min		Skimmer	60		C ESI C APCI C ESI/APCI
Time and Scan Segments Time (minutes) Scans Add 0.00	MMI (Scan)	1000	psig	Capillary	psig	μA	Chamber	0.78	μA	Charging Voltage
Del 0.00 Add	Corona +	6.0	μA	Corona	4923	v				2000 V



MMI MMI	Chromatogram Details	Segments	rauon   Parameo	Chromatograms	Reset
Positive     C Negative	Type EIC 💌	All 0.00		2 1-Hexanesulfonic acid	
Polarity Switch per scan	Label Carbozole		Add->	4 9-Phenanthrol	
Select Scan to Display	Extracted [168-168.3	Scans	Modify		
Add 0.00	Offset 15 % Y-axis range 1000000 counts	Scan 1 Scan 2 Scan 3	Delete		
Del Add	Add Chrometogram with index	Scan 4			
Mod					



Step 5. Create MMCHECKTOF\_CI\_POS.m

Ionization Mode		MM-APCI
Polarity		Positive
1100 Binary Pump 1		
Control		
Column flow		0.400 mL/min
Stop Time		No Limit
Post Time		Off
Solvents		
Solvent A		100.0 % (65%MeOH:35%H <sub>2</sub> O + 0.2%acetic acid))
Solvent B		0.0 %
Pressure Limits		
Minimum Pressure		0 bar
Maximum Pressure		400 bar
Spray Chamber		
[MSZones]		
Gas Temp	350 °C	Maximum 350 °C
Vaporizer	200 °C	Maximum 250 °C
Drying Gas	5.0 L/min	Maximum 13.0 L/min
Neb Pres	20 psig	Maximum 60 psig
VCap (Positive)	1000 V	
VCap (Negative)	1000 V	
VCharge (Positive)	2000 V	
VCharge (Negative)	2000 V	
Corona (Positive)	6.0 μA	
Corona (Negative)	6.0 μA	

# Step 6. Create MMCHECKTOF\_CI\_NEG.m

• Set the parameters for MMCHECKTOF\_CI\_NEG.m:

Ion Source	Data Acq	uisition	Ref. Ma	asses   C	Chromatogram	Tune Calibr	ation	Parameters	Diagnostics
Ion Polarity (Seg.) C Positive G Negative	Gas Temp	350	с	300	С	Fragmentor	225	v	Source (Seg.)
Polarity Switch per scan Select Scan to Display	Vaporizer Drying Gas	200	C I/min	200	C 1/min	Skimmer	60	v	C ESI
Time and Scan Segments	Nebulizer	20	psig	20	psig	OCT RFV	250	v	C ESI/APCI
Add 0.00	MMI (Scan) VCap	1000	v	Capillary	μ 800.0	A Chamber	1.41	μA	Charging Voltage
Mod Del	Lorona -	J6.0	μA	Lorona	2781 V				



MMI MMI	Chromatogram Details Type EIC	Segments	ion   Parameters	Chromatograms	Reset
Polarity Switch per scan	Label 9-Phenanthrol	0.00	Add->	3 Carbazole 4 9 Phenonthrol	
Select Scan to Display Time and Scan Segments Time (minutes)	Extracted 193-193.3	Scans	Modify		
Add 0.00	Offset 15 % Y-axis range 1000000 counts	Scan 1 Scan 2 Scan 3	Delete		
Del Add	Add Chromatooram with index	Scan 4			
MOD				J	]

Figure 27 Chromatogram

Step 6. Create MMCHECKTOF\_CI\_NEG.m

Ionization Mode		MM-APCI
Polarity		Negative
1100 Binary Pump 1		
Control		
Column flow		0.400 mL/min
Stop Time		No Limit
Post Time		Off
Solvents		
Solvent A		100.0 % (65%MeOH:35%H <sub>2</sub> O + 0.2%acetic acid))
Solvent B		0.0 %
Pressure Limits		
Minimum Pressure		0 bar
Maximum Pressure		400 bar
Spray Chamber		
[MSZones]		
Gas Temp	350 °C	Maximum 350 °C
Vaporizer	200 °C	Maximum 250 °C
Drying Gas	5.0 L/min	Maximum 13.0 L/min
Neb Pres	20 psig	Maximum 60 psig
VCap (Positive)	1000 V	
VCap (Negative)	1000 V	
VCharge (Positive)	2000 V	
VCharge (Negative)	2000 V	
Corona (Positive)	6.0 μA	
Corona (Negative)	6.0 μA	

Step 7. Create MMCHECKTOF\_MX\_EI POS\_CI POS.m

# Step 7. Create MMCHECKTOF\_MX\_EI POS\_CI POS.m

• Set the parameters for MMCHECKTOF\_MX\_EI POS\_CI POS.m.

MMI MMI	Data Acq	isition	Ref. Ma	asses   (	Chromatogram   T	- MS TOP IS	ation	Parameters	Diagnostics
lon Polarity (Seg.)	Gas Temp	350	с	300	с	Fragmentor	225	v	Source (Seg.)
Polarity Switch per scan	Vaporizer Drying Gas	200	C I/min	3.4	C - I/min	Skimmer	60	v	C APCI
Time and Scan Segments Time (minutes) Scans	Nebulizer	60	psig	20	psig	OCT RF V	250	V	C ESI/APCI
Add 0.00	- MMI (Scan) VCap	1000	v	Capillary	μΑ	Chamber	0.00	_μΑ	Charging Voltage
Mod Del	Corona +	1.0	μĄ	Corona	3291 V				



MMI MMI Ion Polarity (Seg.) © Positive © Negative	Chromatogram Details Type EIC _	Segments		Chromatograms	Reset
Polarity Switch per scan	Label Crystal Violet	0.00	Add ->	3 Carbazole 4 9-Phenanthrol	
Select Scan to Display Time and Scan Segments Time (minutes) Scans	Extracted Masses 372-372.4	Scens .	Modify		
Add 0.00	Y-axis range 1000000 counts	Scan 1 Scan 2 Scan 3	Delete		
Mod Dei	Add Chromatogram with index	Scan 4			
	-				

Figure 29 Chromatogram

Step 7. Create MMCHECKTOF\_MX\_EI POS\_CI POS.m

Ionization Mode		MM-ES+APCI	
Polarity		Positive	
1100 Binary Pump 1			
Control			
Column flow		0.400 mL/min	
Stop Time		No Limit	
Post Time		Off	
Solvents			
Solvent A		100.0 % (65%MeOH:35%H	20 + 0.2% acetic acid))
Solvent B		0.0 %	
Pressure Limits			
Minimum Pressure		0 bar	
Maximum Pressure		400 bar	
Spray Chamber			
[MSZones]			
Gas Temp	350 °C		Maximum 350 °C
Vaporizer	200 °C		Maximum 250 °C
Drying Gas	5.0 L/m	nin	Maximum 13.0 L/min
Neb Pres	60 psig		Maximum 60 psig
VCap (Positive)	1000 V		
VCap (Negative)	1000 V		
VCharge (Positive)	2000 V		
VCharge (Negative)	2000 V		
Corona (Positive)	1.0 µA		
Corona (Negative)	1.0 µA		

Step 8. Create MMCHECKTOF\_MX\_EI NEG\_CI NEG.m

# Step 8. Create MMCHECKTOF\_MX\_EI NEG\_CI NEG.m

• Set the parameters for MMCHECKTOF\_MX\_EI NEG\_CI NEG.m:

MMI MMI	Data Acqu	uisition	Ref. Ma	asses   C	hromatogram   T	une Calibr	ation   Parame	eters Diagnostics
Ion Polarity (Seg.)	Gas Temp	350	С	300	С	Fragmentor	225 V	Source (Seg.)
Polarity Switch per scan	Vaporizer Drying Gas	200	C I/min	199 3.4	C Vmín	Skimmer	60 V	C ESI C APCI
Time and Scan Segments	Nebulizer	60	psig	20	psig	OCT RF V	250 V	
Add 0.00	MMI (Scan) VCap	1000	v	Capillary	0.000 μA	Chamber	μA	Charging Voltage
Del 0.00 Add	Corona -	1.0	μΑ	Corona	2551 V			J2000 V



MMI MMI on Polarity (Seg.) ^ Positive & Negative	Chromatogram Details Type EIC	Segments	Chromatograms 1 Crystal Violet 21+Hexenesulfonic acid	Reset
Polarity Switch per scan Select Scan to Display	Label 1-Hexanesulfonic acid	A	dd-> 3 Carbazole 4 9-Phenanthrol	
Time and Scan Segments Time (minutes) Scans	Masses 165-165.3	ScansM	odity	
Add 0.00 1 Add	V-axis range 1000000 counts	Scan 2 D Scan 2 D Scan 3	elete	
Mod	Add Chromatogram with index			

Figure 31 Chromatogram

Ionization Mode	MM-ES+A	APCI
Polarity	Negative	
1100 Binary Pump 1		
Control		
Column flow	0.400 mL/	′min
Stop Time	No Limit	
Post Time	Off	
Solvents		
Solvent A	100.0 % (6	55%MeOH:35%H <sub>2</sub> O + 0.2% acetic acid))
Solvent B	0.0 %	
Pressure Limits		
Minimum Pressure	0 bar	
Maximum Pressure	400 bar	
Spray Chamber		
[MSZones]		
Gas Temp	350 °C	Maximum 350 °C
Vaporizer	200 °C	Maximum 250 °C
Drying Gas	5.0 L/min	Maximum 13.0 L/min
Neb Pres	60 psig	Maximum 60 psig
VCap (Positive)	1000 V	
VCap (Negative)	1000 V	
VCharge (Positive)	2000 V	
VCharge (Negative)	2000 V	
Corona (Positive)	1.0 μA	
Corona (Negative)	1.0 µA	

Step 9. Run each of the methods created

# Step 9. Run each of the methods created

 Run each of the methods that you just created. The real time plot below shows the six runs.



**2** View the data from Analyst for MM\_ESI\_pos.wif. Exstract Ion 372- 372.4. Record peak height Example: 91,000.



Step 9. Run each of the methods created



**3** View the data in the data analysis program for MM\_ESI\_Neg. Extract Ion 165-165.4. Record the peak height Example 97,000.

Step 9. Run each of the methods created



**4** View the data in the data analys program for MM\_APCI\_POS. Extract Ion 168-168.4. Record the peak height. Example 140,000.

Step 9. Run each of the methods created



**5** View the data in the data analysis program for MM\_APCI\_NEG. Extract Ion 193-193.4. Record the peak height. Example 640,000.

Step 9. Run each of the methods created



**6** View the data in the data analysis program for MM\_ESI\_APCI\_POS. Extract Ion 372-372.4. Record the peak height. Example: 57,000.

Step 9. Run each of the methods created



7 View the data in the data analysis program for MM\_ESI\_APCI\_POS. Extract Ion 168-168.4. Record the peak height. Example: 34,000.

Step 9. Run each of the methods created



8 View the data in the data analysis program for MM\_ ESI\_APCI\_NEG. Extract Ion 165-165.4. Record the peak height. Example: 110,000.

Step 9. Run each of the methods created



**9** View the data in the data analysis program for MM\_ ESI\_APCI\_NEG. Extract Ion 193-193.4. Record the peak height. Example: 400,000.

Step 10. Calculate the response of Multimode Demo

# Step 10. Calculate the response of Multimode Demo

1 Manually fill in the values in the Multimode Ion Source report.

The values in the example report below have been manually entered from the data collected in the runs from the previous steps. This is an example of how to enter the values from the instrument being installed and verified. The blank report is on the next page for installed instruments data.

м	ultimode	Ion Source	Report			
MSD type: TOF		Instrument	name:		Operator	name:
Acquisition date:	23-Feb-2	2006				
Datafiles: MM_ESI_pos.wif MM_ASI_POS.wif MM_APCI_POS.wif MM_ESI_APCI_POS. MM_ESI_APCI_POS. MM_ESI_APCI_NEG.	wif wif	ESI Com	bound Resul			
Compound	   m/z	Polarity	ESI mode	Mixed mode	Mixed:ESI ratio	Result
Crystal violet	372.2	Positive	 91k	57k	63%	Pass

		APCI CON	npound Resu	ults		
Compound	m/z	Polarity	APCI mode	Mixed mode	Mixed:APCI ratio	Result
Carbazole	168.1	Positive	140k	34k	24%	Pass
9-Phenanthrol	193.1	Negative	 640k	400k	63%	Pass

Passing criteria: Mixed mode response 20% or greater of single-mode response.

2 Run all methods and get the peak heights. Calculate the amount of signal.

# Step 11. Fill out Multimode Report for calculation of peak heights

• Use the graphic below to fill out the multimode report for calculation of peak heights.

M	ultimode	Ion Source	Report			
SD type: TOF		Instrument	name:		Operator r	name:
cquisition date:	23-Feb-2	2006				
atafiles: MM_ESI_pos.wif MM_ESI_Neg.wif MM_APCI_POS.wif MM_APCI_POS.wif MM_ESI_APCI_POS. MM_ESI_APCI_NEG.	wif wif					
		ESI Comp	ound Resul	ts		
Compound	m/z	Polarity	ESI   mode	Mixed mode	Mixed:ESI ratio	Result
Crystal violet	372.2	Positive				
1-Hexanesulf- onic acid	165.1	Negative				
		APCI COM	pound Resu	lts		
Compound	m/z	Polarity	APCI   mode	Mixed mode	Mixed:APCI ratio	Result
Carbazole	168.1	Positive				

Passing criteria: Mixed mode response 20% or greater of single-mode response.

3

Step 11. Fill out Multimode Report for calculation of peak heights

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# www.agilent.com

# In This Book

This book contains installation, operation, maintenance and troubleshooting instruction for the Multimode Source for 6500 Series Q-TOF LC/MS.

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