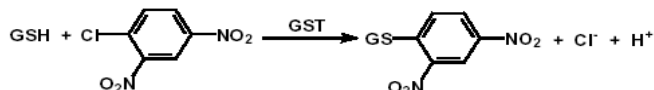


GST Colorimetric Activity Assay Kit

(Catalog #K263-100; 100 assays; Store kit at -20°C)

I. Introduction:

Glutathione S-transferase (GST) is a family of enzymes that plays an important role in detoxification of xenobiotics. GST catalyzes attachment of the thiol of glutathione to electrophiles. Glutathione is used to scavenge potentially toxic compounds including those produced as a result of oxidative stress and is part of the defense mechanism neutralizing the mutagenic, carcinogenic and toxic effects of such compounds. The *GST Colorimetric Activity Assay Kit* is based upon the GST-catalyzed reaction between GSH and the GST substrate, CDNB (1-chloro-2,4-dinitrobenzene, which has the broadest range of isozyme detectability (e.g., alpha-, mu-, pi-, and other GST isoforms, except theta). Under certain conditions, the interaction between glutathione and CDNB is totally dependent on the presence of active GST.



The GST-catalyzed formation of GS-DNB produces a dinitrophenyl thioether which can be detected by spectrophotometry at 340 nm. One unit of GST activity is defined as the amount of enzyme producing 1 μmol of GS-DNB conjugate/min under the conditions of the assay. The kit can detect GST activity in crude cell lysate or purified protein fractions, and can quantitate GST-tagged fusion proteins. Detect limit: Active GST < 1mU.

II. Kit Contents:

Component	K263-100	Cap Code	Part Number
GST Assay Buffer	25 ml	WM	K263-100-1
GST Substrate (CDNB)	0.1 ml	Red	K263-100-2
Glutathione (GSH, lyophilized)	2 x 17 mg	Yellow	K263-100-3
GST Positive Control	10 μl	Green	K263-100-4

III. Reagent Preparation and Storage Conditions:

GST Assay Buffer: store at 4 °C

GSH: Add 275 μl of GST Assay Buffer to each vial just before use. One vial is sufficient for 50 assays. The Remaining solution can be kept at -20°C for 1 week.

CDNB: This vial contains a DMSO solution of 1-chloro-2, 4-dinitrobenzene (CDNB) and should be stored at -20°C.

GST Positive Control: Store at -20 °C

IV. Sample Preparation Guideline:

A. Cell Sample Preparation:

1. Collect cells by centrifugation. For adherent cells, use a rubber policeman to scrape and collect the cells.
2. Homogenize or sonicate the cells in GST Assay Buffer (typically 3 -4 volumes).
3. Centrifuge at 10,000 x g for 15 min at 4°C.
4. Collect supernatant and use for the assay. The remaining sample should be stored at -80°C, and is stable for at least 1 month.

B. Tissue Sample Preparation:

1. Prior to dissection, perfuse tissue with PBS containing heparin (0.15 mg/ml) to remove red blood cells and clots.
2. Homogenize tissue in GST Assay Buffer (100 mg/0.5 ml).
3. Centrifuge at 10,000 x g for 15 minutes at 4°C.
4. Collect supernatant and use for the assay. The remaining sample should be stored at -80°C, and is stable for at least 1 month.

C. Plasma and Erythrocyte Sample Preparation:

1. Centrifuge anticoagulant treated blood at 1000 x g for 10 min at 4°C.

2. Transfer the top plasma layer (without disturbing the white buffy layer) to a new tube and store on ice for assay or store at -80°C for future use. The plasma should be stable for 1 month.
3. Remove the white buffy layer and discard (leukocytes).
4. Lyse the erythrocytes (red blood cells) in 4 times its volume of ice-cold GST Assay Buffer.
5. Centrifuge at 10,000 x g for 15 min at 4°C.
6. Transfer supernatant (erythrocyte lysate) to a new tube, and use it for the GST assay. The remaining samples should be stored at -80°C for future use and is stable for at least one month.

D. Preparation of Bacterially Expressed GST-Fusion Protein Sample:

1. Collect bacteria by centrifugation. Freeze/thaw the pellet two times, then sonicate in GST Assay Buffer.
2. Centrifuge at 10,000 x g for 15 min at 4°C.
3. Transfer supernatant to a new tube, and use it for the GST assay. The remaining samples should be stored at -80°C for future use and is stable for at least one month.

V. GST Assay Protocol:

1. **Sample, Negative Control and Positive Control Preparation:** Prepare samples in a total 50 μl volume with GST Assay Buffer, including a negative control with 50 μl of GST Assay buffer only. For GST Positive Control, dilute 100 time by adding 2 μl of Positive Control into 198 μl GST Assay Buffer, add 2-10 μl of diluted GST Positive Control into desired well (s) and adjust the final volume to 50 μl with GST Assay Buffer. **Note:** We recommend preparing several dilutions of your sample and running duplicate wells for each measurement.
2. **Glutathione Addition:** Add 5 μl of Glutathione to each well containing the sample or control above.
3. **Substrate Mix:** Mix enough reagents for the number of assays to be performed. For each well, prepare a total 50 μl Substrate Mix containing:
 - GST Assay Buffer 49 μl
 - GST Substrate (CDNB) Solution 1 μl
 Mix well and transfer 50 μl of the Mix into each sample (including the standard) well.
4. **Measurement:** Carefully shake the plate to start the reaction. Read the absorbance once every minute at 340 nm using a plate reader to obtain at least 5 time points. For low GST activity samples, the reaction can be continued for longer time periods

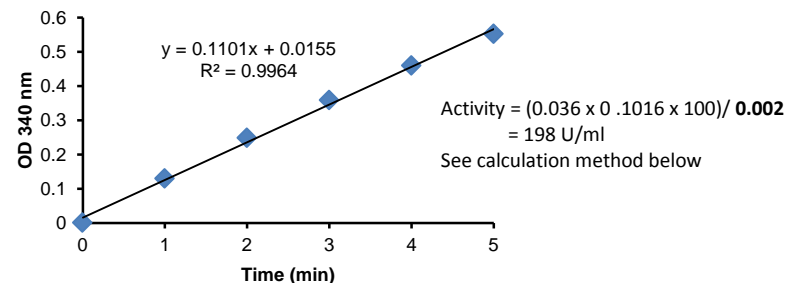


Figure : GST Kinetic Assay Performed According to This Protocol

5. Calculation of GST Assay Results:

- a) Determine the change in absorbance (ΔA_{340}) per minute by:
 - i. Plotting the absorbance values as a function of time to obtain the slope (rate) of the linear portion of the curve.
 - ii. Select two points on the linear portion of the curve and determine the change in absorbance during that time, using the following equation:

$$\Delta A_{340}/\text{min} = \frac{A_{340}(\text{Time 2}) - A_{340}(\text{Time 1})}{\text{Time 2 (min)} - \text{Time 1 (min)}}$$

- b) Determine the rate of $\Delta A_{340}/\text{min}$ for the background wells and subtract the rate from that of the sample wells.
- c) Use the following formula to calculate the GST activity (U/ml of sample). The reaction rate at 340 nm can be determined using the GS-DNB extinction coefficient at 340 nm $0.0096 \mu\text{M}^{-1}\text{cm}^{-1}$. The value has been adjusted for the path length of the solution in the well 0.2893 cm).

$$\text{GST Activity} = \frac{\Delta A_{340}\text{min}^{-1} \times \text{Reaction Volume (ml)}}{0.0096 \mu\text{mol}^{-1}\text{cm}^{-1} \times 1000 \text{ ml} \times 0.2893 \text{ cm} \times V} \times D$$

$$= \Delta A_{340}\text{min}^{-1} / x 0.036x D/V (\mu\text{mol}/\text{min}/\text{ml})$$

Where:

$0.0096 \mu\text{mol}^{-1}\text{cm}^{-1}$ is the extinction coefficient of the glutathione-DNB adduct.

V = Sample Volume added to well (ml)

D = Sample Dilution Factor

0.2893 cm is light path of the 0.1 ml Reaction Volume in a Greiner Bio One 655101 96 well plate (cm). Other plates must be calibrated for accurate results.

Unit Definition: One unit is the amount of enzyme that conjugates 1.0 μmol of 1-Chloro-2,4-Dinitrobenzene with reduced glutathione per min. at pH 6.4 at 25°C.

RELATED PRODUCTS:

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- Caspase Assay Kits & Reagents
- Mitochondrial Apoptosis Kits & Reagents
- Nuclear Apoptosis Kits & Reagents

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- Mitochondria/Cytosol Fractionation Kit
- Nuclear/Cytosol Fractionation Kit
- Membrane Protein Extraction Kit
- Cytosol/Particulate Rapid Separation Kit
- Mammalian Cell Extraction Kit
- FractionPREP Fractionation System

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- Senescence Detection Kit
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- LDH-Cytotoxicity Assay Kit
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Monoclonal and Polyclonal Antibodies and Cytokines

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GENERAL TROUBLESHOOTING GUIDE:

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> • Use of ice-cold assay buffer • Omission of a step in the protocol • Plate read at incorrect wavelength • Use of a different 96-well plate 	<ul style="list-style-type: none"> • Assay buffer must be at room temperature • Refer and follow the data sheet precisely • Check the wavelength in the data sheet and the filter settings of the instrument • Fluorescence: Black plates (clear bottoms) ; Luminescence: White plates ; Colorimeters: Clear plates
Samples with erratic readings	<ul style="list-style-type: none"> • Use of an incompatible sample type • Samples prepared in a different buffer • Cell/ tissue samples were not completely homogenized • Samples used after multiple free-thaw cycles • Presence of interfering substance in the sample • Use of old or inappropriately stored samples 	<ul style="list-style-type: none"> • Refer data sheet for details about incompatible samples • Use the assay buffer provided in the kit or refer data sheet for instructions • Use Dounce homogenizer (increase the number of strokes); observe for lysis under microscope • Aliquot and freeze samples if needed to use multiple times • Troubleshoot if needed • Use fresh samples or store at correct temperatures until use
Lower/ Higher readings in Samples and Standards	<ul style="list-style-type: none"> • Improperly thawed components • Use of expired kit or improperly stored reagents • Allowing the reagents to sit for extended times on ice • Incorrect incubation times or temperatures • Incorrect volumes used 	<ul style="list-style-type: none"> • Thaw all components completely and mix gently before use • Always check the expiry date and store the components appropriately • Always thaw and prepare fresh reaction mix before use • Refer datasheet & verify correct incubation times and temperatures • Use calibrated pipettes and aliquot correctly
Readings do not follow a linear pattern for Standard curve	<ul style="list-style-type: none"> • Use of partially thawed components • Pipetting errors in the standard • Pipetting errors in the reaction mix • Air bubbles formed in well • Standard stock is at an incorrect concentration • Calculation errors • Substituting reagents from older kits/ lots 	<ul style="list-style-type: none"> • Thaw and resuspend all components before preparing the reaction mix • Avoid pipetting small volumes • Prepare a master reaction mix whenever possible • Pipette gently against the wall of the tubes • Always refer the dilutions in the data sheet • Recheck calculations after referring the data sheet • Use fresh components from the same kit
Unanticipated results	<ul style="list-style-type: none"> • Measured at incorrect wavelength • Samples contain interfering substances • Use of incompatible sample type • Sample readings above/below the linear range 	<ul style="list-style-type: none"> • Check the equipment and the filter setting • Troubleshoot if it interferes with the kit • Refer data sheet to check if sample is compatible with the kit or optimization is needed • Concentrate/ Dilute sample so as to be in the linear range
<p>Note: The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.</p>		