

# RayBio<sup>®</sup> Human/Mouse/Rat Gastrin-1 Enzyme Immunoassay Kit

Catalog #: EIA-GAS, EIAM-GAS, EIAR-GAS

User Manual

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Caution:  
Extraordinarily useful information enclosed



ISO 13485 Certified

3607 Parkway Lane, Suite 100  
Norcross, GA 30092

Tel: 1-888-494-8555 (Toll Free) or 770-729-2992, Fax: 770-206-2393

Web: [www.RayBiotech.com](http://www.RayBiotech.com), Email: [info@raybiotech.com](mailto:info@raybiotech.com)

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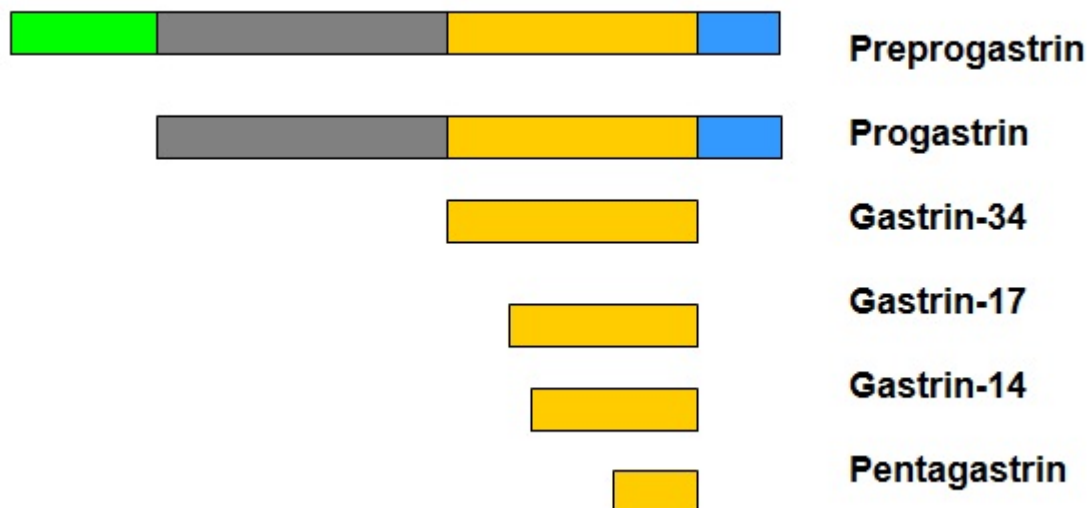
Please read the entire manual carefully before starting your experiment

# I. Introduction

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Gastrin is a hormone that stimulates secretion of gastric acid by the parietal cells of the stomach and aids in gastric motility. It is released by G cells in the stomach, duodenum, and the pancreas.

Gastrin is a linear peptide that is synthesized as a preprohormone and post-translationally cleaved to form a family of peptides with identical carboxytermini. The predominant circulating form is gastrin-34 (a.k.a. "big gastrin"), but full biologic activity is present in the smaller peptides, i.e., gastrin-17 (a.k.a. "little gastrin") and gastrin-14 (a.k.a. "minigastrin"). In addition, pentagastrin is an artificial 5-amino-acid peptide identical to the carboxyterminus of gastrin. It is worth noting that the five C-terminal amino acids of gastrin and cholecystikinin are identical, which explains their overlapping biological effects.



Gastrin is released in response to certain stimuli including stomach distension, vagal stimulation, hypercalcemia and the presence of partially digested proteins. On the other hand, gastrin release can be inhibited by the presence of gastric acid or inhibitory hormones such as secretin, GIP, VIP, glucagon and calcitonin. As a peptide hormone in GI system, gastrin stimulates gastric acid secretion from parietal cells of the stomach. Additional functions by gastrin include the following, Stimulating parietal cell maturation and fundal growth; Causing chief cells to secrete pepsinogen; Increasing antral muscle mobility and promoting stomach contractions; Strengthening antral contractions against the pylorus, and constricting the pyloric sphincter to slow gastric emptying; inducing pancreatic secretions and gallbladder emptying.

The role of gastrin in certain diseases has also been reported, including Zollinger-Ellison Syndrome, autoimmune gastritis and mucopolidosis. In Zollinger-Ellison syndrome, gastrin is produced at excessive levels, often by a gastrinoma of the duodenum or the pancreas. In autoimmune gastritis, the immune system attacks the parietal cells leading to hypochlorhydria. This results in an elevated gastrin level in an attempt to compensate for increased pH in the stomach. Eventually, all the parietal cells are lost and achlorhydria results leading to a loss of negative feedback on gastrin secretion. Plasma gastrin concentration is elevated in virtually all individuals with mucopolidosis type IV secondary to a constitutive achlorhydria.

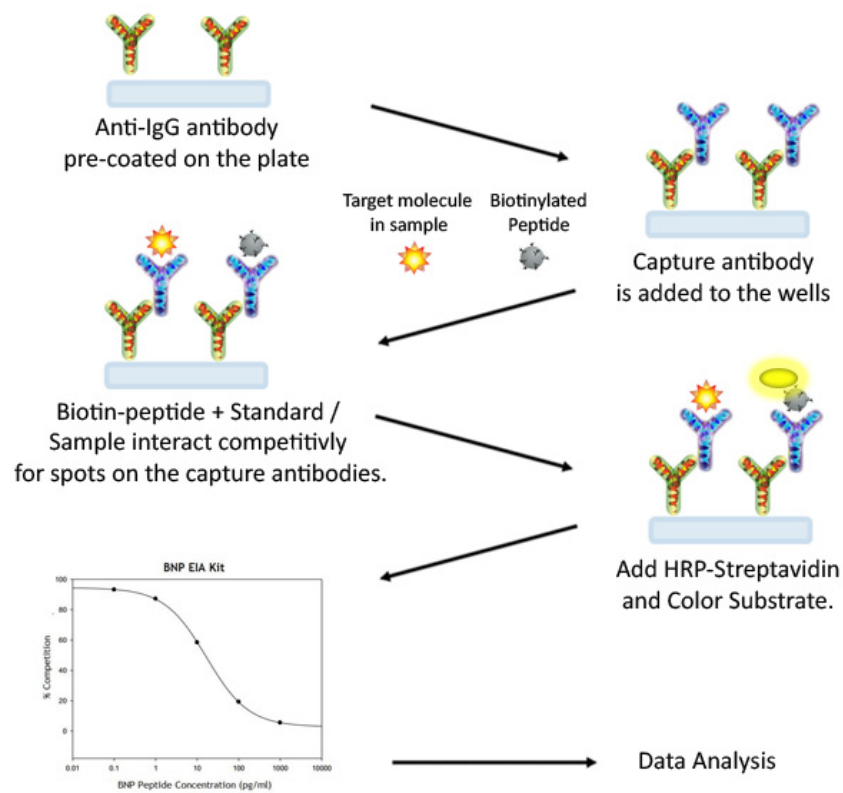
## **II. General Description**

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The RayBio<sup>®</sup> Gastrin-1 Enzyme Immunoassay (EIA) Kit is an in vitro quantitative assay for detecting Gastrin-1 peptide based on the competitive enzyme immunoassay principle.

In this assay, a biotinylated Gastrin-1 peptide is spiked into the samples and standards. The samples and standards are then added to the plate, where the biotinylated Gastrin-1 peptide competes with endogenous (unlabeled) Gastrin-1 for binding to the anti-Gastrin-1 antibody. After a wash step, any bound biotinylated Gastrin-1 then interacts with horseradish peroxidase (HRP)-streptavidin, which catalyzes a color development reaction. The intensity of the colorimetric signal is directly proportional to the amount of captured biotinylated Gastrin-1 peptide and inversely proportional to the amount of endogenous Gastrin-1 in the standard or samples. A standard curve of known concentration of Gastrin-1 peptide can be established and the concentration of Gastrin-1 peptide in the samples can be calculated accordingly.

### III. How It Works



## IV. Storage

The entire kit may be stored at -20°C to -80°C for up to 6 months from the date of shipment. For extended storage, it is recommended to store at -80°C. **Avoid repeated freeze-thaw cycles.** For prepared reagent storage, see table below.

## V. Reagents

Component	Size / Description	Storage / Stability After Preparation
Gastrin-1 Microplate (Item A)	96 wells (12 strips x 8 wells) coated with secondary antibody.	1 month at 4°C*
Wash Buffer Concentrate (20X) (Item B)	25 ml of 20X concentrated solution.	1 month at 4°C
Standard Gastrin-1 Peptide (Item C)	2 vials of Gastrin-1 Peptide. 1 vial is enough to run each standard in duplicate.	The first standard: 2-3 days at 4°C Additional dilutions: Do not store
Anti-Gastrin-1 Polyclonal Antibody (Item N)	2 vials of anti-Gastrin-1.	1 month at 4°C
Assay Diluent A (Item D)	30 ml, contains 0.09% sodium azide as preservative. Diluent for standards and serum or plasma.	N/A
Assay Diluent B (Item E)	15 ml of 5X concentrated buffer. Diluent for standards, cell culture media or other sample types, and HRP-Streptavidin.	1 month at 4°C
Biotinylated Gastrin-1 Peptide (Item F)	2 vials of Biotinylated Gastrin-1 Peptide, 1 vial is enough to assay the whole plate.	2-3 days at 4°C
HRP-Streptavidin Concentrate (Item G)	600 µl 200X concentrated HRP-conjugated streptavidin.	Do not store and reuse
Positive Control (Item M)	1 vial of Positive Control.	2-3 days at 4°C
TMB One-Step Substrate Reagent (Item H)	12 ml of 3,3',5,5'-tetramethylbenzidine (TMB) in buffer solution.	N/A
Stop Solution (Item I)	8 ml of 0.2 M sulfuric acid.	N/A

\*Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

## VI. Additional Materials Required

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1. Microplate reader capable of measuring absorbance at 450 nm
2. Precision pipettes to deliver 2 µl to 1 ml volumes
3. Adjustable 1-25 ml pipettes for reagent preparation
4. 100 ml and 1 liter graduated cylinders
5. Absorbent paper
6. Distilled or deionized water
7. SigmaPlot software (or other software which can perform four-parameter logistic regression models)
8. Tubes to prepare standard or sample dilutions
9. Orbital shaker
10. Aluminum foil
11. Plastic wrap

## VII. Reagent Preparation

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Keep kit reagents on ice during reagent preparation steps.

Note: **Assay Diluent A** should be used for dilution of samples, Item F and Item C when testing **plasma or serum samples**. **1X Assay Diluent B** should be used for dilution of samples, Item F and Item C when testing **cell culture media or other sample types**.

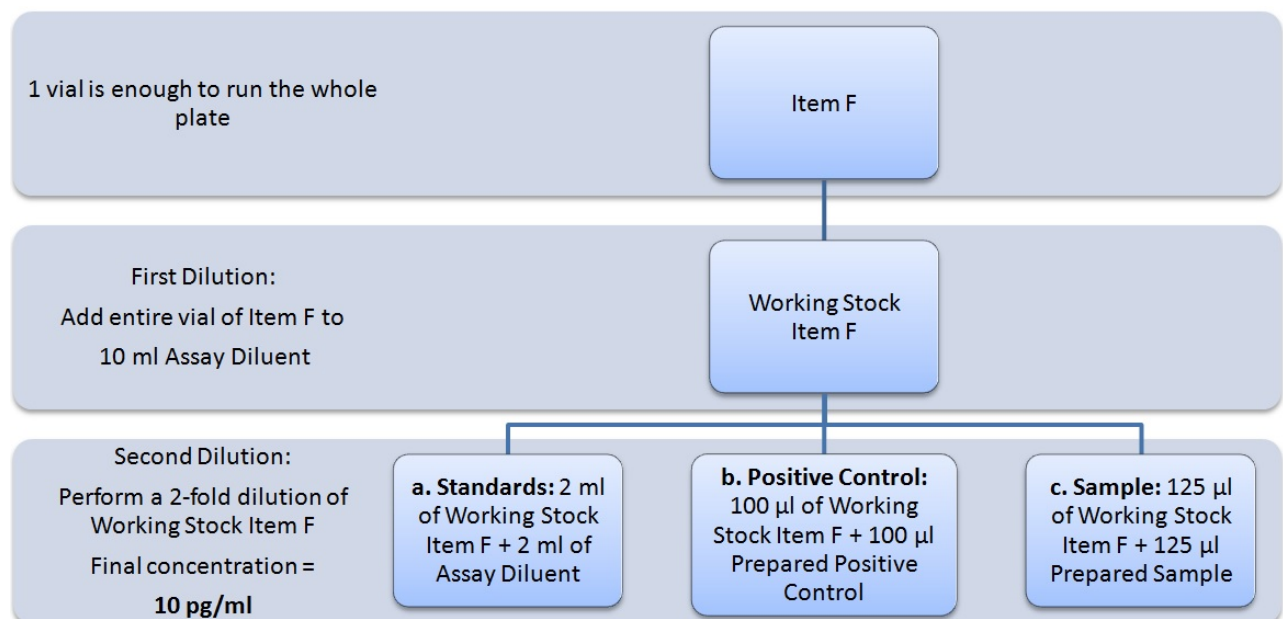
### A. Preparation of Plate and Anti-Gastrin-1 Antibody

1. Equilibrate plate to room temperature before opening the sealed pouch.
2. Label removable 8-well strips as appropriate for your experiment.
3. 5X Assay Diluent B (Item E) should be diluted 5-fold with deionized or distilled water.
4. Briefly centrifuge the anti-Gastrin-1 antibody vial (Item N) Then add 50 µl of 1X Assay Diluent B to the vial to prepare the antibody concentrate. Pipette up and down to mix gently.
5. The antibody concentrate should then be diluted 100-fold with 1X Assay Diluent B. This is your anti-Gastrin-1 antibody working solution, which will be used in step 2 of Assay Procedure (Section VIII).

*Note: The following steps may be done during the antibody incubation procedure (step 2 of Assay Procedure)*

## B. Preparation of Biotinylated Gastrin-1 (Item F)

5. Briefly centrifuge the vial of Biotinylated Gastrin-1 (Item F) before use.
6. See the image below for proper preparation of Item F. Transfer the entire contents of the Item F vial into a tube containing 10 ml of the appropriate Assay Diluent. This is your Working Stock of Item F. Pipette up and down to mix gently. *The final concentration of biotinylated Gastrin-1 will be **20 pg/ml**.*
  - a. Second Dilution of Item F for Standards: Add 2 ml of Working Stock Item F to 2 ml of the appropriate Assay Diluent. The final concentration of biotinylated Gastrin-1 will be **10 pg/ml**.
  - b. Second Dilution of Item F for Positive Control: Add 100  $\mu$ l of Working Stock Item F to 100  $\mu$ l of the prepared Positive Control (Item M). (See section D for Positive Control preparation) The final concentration of biotinylated Gastrin-1 will be **10 pg/ml**.
  - c. Second Dilution of Item F for samples: Add 125  $\mu$ l of Working Stock Item F to 125  $\mu$ l of prepared sample (see section E for sample preparation). This is a 2-fold dilution of your sample. The final concentration of biotinylated Gastrin-1 will be **10 pg/ml**.



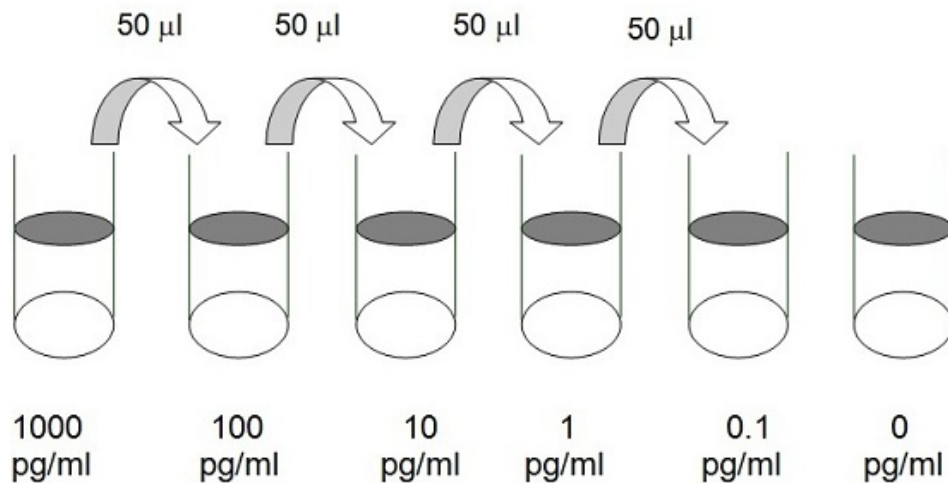


### C. Preparation of Standards

7. Label 6 microtubes with the following concentrations: 1000 pg/ml, 100 pg/ml, 10 pg/ml, 1 pg/ml, 0.1 pg/ml and 0 pg/ml. Pipette 450  $\mu$ l of biotinylated Gastrin-1 Item F working solution (prepared in step 6a) into each tube, except the 1,000 pg/ml (leave this one empty).

*It is very important to make sure the concentration of biotinylated Gastrin-1 is 10 pg/ml in all standards.*

8. Briefly centrifuge the vial of Gastrin-1 Standard (Item C). Pipette 8  $\mu$ l of Item C and 792  $\mu$ l of 10 pg/ml biotinylated Gastrin-1 working solution (prepared in step 6a) into the tube labeled 1000 pg/ml. Mix thoroughly. This solution serves as the first standard (1000 pg/ml Gastrin-1 standard, 10 pg/ml biotinylated Gastrin-1).
9. To make the 100 pg/ml standard, pipette 50  $\mu$ l of the 1000 pg/ml Gastrin-1 standard into the tube labeled 100 pg/ml. Mix thoroughly.
10. Repeat this step with each successive concentration, preparing a dilution series as shown in the illustration below. Each time, use 450  $\mu$ l of biotinylated Gastrin-1 and 50  $\mu$ l of the prior concentration until the 0.1 pg/ml is reached. Mix each tube thoroughly before the next transfer.



## D. Positive Control Preparation

11. Briefly centrifuge the Positive Control vial (Item M).
12. Refer to step 6b. This is a 2-fold dilution of the Positive Control. The final concentration of biotinylated Gastrin-1 should still be 10 pg/ml.

The Positive Control is a cell culture media sample that serves as a system control to verify that the kit components are working. The resulting OD will not be used in any calculations; if no positive competition is observed please contact RayBiotech Technical Support. The Positive Control may be diluted further if desired, but be sure the final concentration of biotinylated Gastrin-1 is 10 pg/ml.

## E. Sample Preparation

13. If you wish to perform a 2-fold dilution of your sample, proceed to step 6c. If you wish to perform a higher dilution of your sample, dilute your sample with the appropriate Assay Diluent before performing step 6c.  
EXAMPLE (to make a 4-fold dilution of sample):
  - a. Dilute sample 2-fold (62.5 µl of sample + 62.5 µl of the appropriate Assay Diluent.).
  - b. Perform step 6c (125 µl of working solution Item F + 125 µl of sample prepared above).

The total volume is 250 µl, enough for duplicate wells on the microplate.

It is very important to make sure the final concentration of the biotinylated Gastrin-1 is **10 pg/ml**.

Note: Optimal sample dilution factors should be determined empirically, however you may reference below for recommended dilution factors for serum: Human=2X  
Mouse=2X Rat=2X.

If you have any questions regarding the recommended dilutions you may contact technical support at 888-494-8555 or [techsupport@raybiotech.com](mailto:techsupport@raybiotech.com).

## F. Preparation of Wash Buffer and HRP

14. If Item B (20X Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved.
15. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
16. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use.
17. Dilute the HRP-Streptavidin concentrate 200-fold with 1X Assay Diluent B.

*Note: do **not** use Assay Diluent A for HRP-Streptavidin preparation in step 17*

## VIII. Assay Procedure

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1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100  $\mu$ l of Anti-Gastrin-1 Antibody (Item N) (See Reagent Preparation step 3) to each well. Incubate for 1.5 hours at room temperature with gentle shaking (1-2 cycle/sec). You may also incubate overnight at 4°C.
3. Discard the solution and wash wells 4 times with 1X Wash Solution Buffer (200-300  $\mu$ l each). Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100  $\mu$ l of each standard (see Reagent Preparation Section C), Positive Control (see Reagent Preparation Section D) and sample (see Reagent Preparation Section E) in appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1-2 cycles/sec) overnight or at 4°C.
5. Discard the solution and wash 4 times as directed in Step 3.
6. Add 100  $\mu$ l of prepared HRP-Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle

shaking. It is recommended that incubation time should not be shorter or longer than 45 minutes.

7. Discard the solution and wash 4 times as directed in Step 3.
8. Add 100  $\mu$ l of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1-2 cycles/sec).
9. Add 50  $\mu$ l of Stop Solution (Item I) to each well. Read at 450 nm immediately.

## **IX. Assay Procedure Summary**

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1. Prepare all reagents, samples and standards as instructed.
2. Add 100  $\mu$ l anti-Gastrin-1 to each well. Incubate 1.5 hours at room temperature or overnight at 4°C.
3. Add 100  $\mu$ l standard or sample to each well. Incubate 2.5 hours at room temperature or overnight at 4°C.
4. Add 100  $\mu$ l prepared Streptavidin solution. Incubate 45 minutes at room temperature.
5. Add 100  $\mu$ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
6. Add 50  $\mu$ l Stop Solution to each well. Read at 450 nm immediately.

## X. Calculation of Results

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Calculate the mean absorbance for each set of duplicate stands, controls, and samples and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit curve through the standard points.

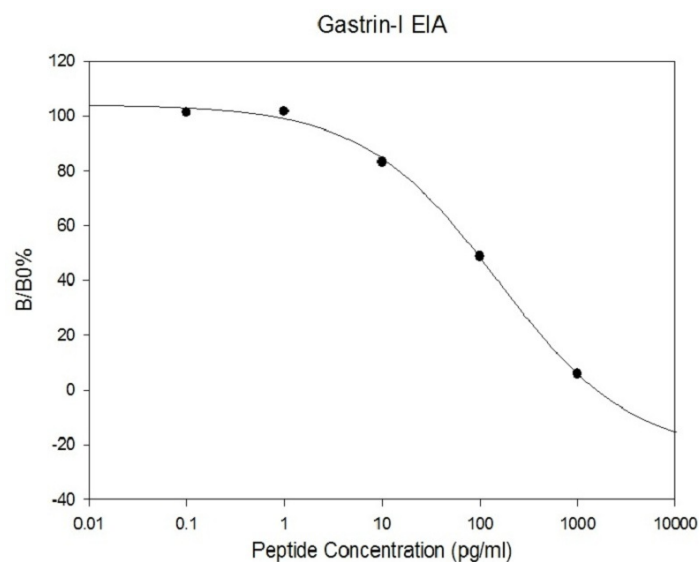
Percentage absorbance =  $(B - \text{blank OD}) / (B_0 - \text{blank OD})$  where

B = OD of sample or standard and

$B_0$  = OD of zero standard (total binding)

### A. Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



### B. Sensitivity

The minimum detectable concentrations of Gastrin-1 is 9.92 pg/ml.

### C. Detection Range

0.1-1,000 pg/ml

### D. Reproducibility

Intra-Assay: CV<10%

Inter-Assay: CV<15%

## E. Assay Diagram

Recommended Plate Layout:

Blank	Blank	SA1	SA1	SA9	SA9	SA17	SA17	SA25	SA25	SA33	SA33
Total Binding	Total Binding	SA2	SA2	SA10	SA10	SA18	SA18	SA26	SA26	SA34	SA34
Standard1	Standard1	SA3	SA3	SA11	SA11	SA19	SA19	SA27	SA27	SA35	SA35
Standard2	Standard2	SA4	SA4	SA12	SA12	SA20	SA20	SA28	SA28	SA36	SA36
Standard3	Standard3	SA5	SA5	SA13	SA13	SA21	SA21	SA29	SA29	SA37	SA37
Standard4	Standard4	SA6	SA6	SA14	SA14	SA22	SA22	SA30	SA30	SA38	SA38
Standard5	Standard5	SA7	SA7	SA15	SA15	SA23	SA23	SA31	SA31	SA39	SA39
Pos Control	Pos Control	SA8	SA8	SA16	SA16	SA24	SA24	SA32	SA32	SA40	SA40

Key:

Blank = Buffer Only

Total Binding = Biotin-Gastrin-1 only

Standard 1 = 1000 pg/ml

Standard 2 = 100 pg/ml

Standard 3 = 10 pg/ml

Standard 4 = 1 pg/ml

Standard 5 = 0.1 pg/ml

Pos Control = Biotin with Item M

## XI. Specificity

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Cross Reactivity: This EIA kit shows no cross-reactivity with any of the cytokines tested: Ghrelin, Nesfatin, Angiotensin II, NPY and APC.

## XIV. Publications Citing This Product

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1. Singh PK., Hota D., Dutta P., et al. Pantoprazole improves glycemic control in type 2 diabetes: a randomized, double-blind, placebo-controlled trial. J Clin Endocrinol Metab. 2012 Nov;97(11):E2105-8. doi: 10.1210/jc.2012-1720

**Species: Human**

**Sample Type: Plasma**

### XIII. Troubleshooting Guide

Problem	Cause	Solution
Poor standard curve	<ul style="list-style-type: none"> <li>• Inaccurate pipetting</li> <li>• Improper standard dilution</li> </ul>	<ul style="list-style-type: none"> <li>• Check pipettes</li> <li>• Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing</li> </ul>
Low signal	<ul style="list-style-type: none"> <li>• Improper preparation of standard and/or biotinylated antibody</li> <li>• Too brief incubation times</li> <li>• Inadequate reagent volumes or improper dilution</li> </ul>	<ul style="list-style-type: none"> <li>• Briefly spin down vials before opening. Dissolve the powder thoroughly.</li> <li>• Ensure sufficient incubation time; assay procedure step 2 may be done overnight</li> <li>• Check pipettes and ensure correct preparation</li> </ul>
Large CV	<ul style="list-style-type: none"> <li>• Inaccurate pipetting</li> <li>• Air bubbles in wells</li> </ul>	<ul style="list-style-type: none"> <li>• Check pipettes</li> <li>• Remove bubbles in wells</li> </ul>
High background	<ul style="list-style-type: none"> <li>• Plate is insufficiently washed</li> <li>• Contaminated wash buffer</li> </ul>	<ul style="list-style-type: none"> <li>• Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed.</li> <li>• Make fresh wash buffer</li> </ul>
Low sensitivity	<ul style="list-style-type: none"> <li>• Improper storage of the ELISA kit</li> <li>• Stop solution</li> </ul>	<ul style="list-style-type: none"> <li>• Follow storage recommendations in sections IV and V. Keep substrate solution protected from light.</li> <li>• Add stop solution to each well before reading plate</li> </ul>

# RayBio<sup>®</sup> ELISA Kits

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