

Thermo Scientific KingFisher® mL User Manual

Rev. 1.3



Thermo Scientific

KingFisher[®] mL

User Manual

Rev. 1.3

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1 Safety Symbols and Markings

These symbols are intended to draw your attention to particularly important information and alert you to the presence of hazards as indicated.

SAFETY SYMBOLS USED IN THE KINGFISHER mL



Power ON



Power OFF



WEEE symbol This product is required to comply with the European Union's Waste Electrical & Electronic Equipment (WEEE) Directive 2002/96/EC.

WARNING MARKINGS USED IN THE DOCUMENTATION



Caution: risk of electric shock.



Caution: biohazard risk.



Caution: risk of personal injury to the operator or a safety hazard to the surrounding area.



Caution: risk of damage to the instrument, other equipment or loss of performance or function in a specific application.

2 About the User Manual

This user manual has been written for the actual end user (for example, laboratory technician) and provides information on the Thermo Scientific KingFisher mL magnetic particle processor, including the installation and operating instructions.

Read the manual in its entirety prior to operating the instrument.

This user manual has been designed to give you the information you need to:

- Install the KingFisher mL
- Use the KingFisher mL in routine jobs – the processing step
- Perform basic maintenance procedures.

This user manual also describes all the features and specifications of the KingFisher mL hardware and on-board software.

Chapter 15 Appendix A. Application Protocols for KingFisher mL explains the processing principles and procedures.

In Chapter 9 Troubleshooting Guide you will find explanations of all error messages and a problem-solving guide. The user should be familiar with the contents of Chapter 7 on maintenance.

For warranty and ordering information, refer to Chapters 10 Warranty Certificate and 11 Ordering Information.

In an effort to produce useful and appropriate documentation, we appreciate your comments on this document to your local Thermo Fisher Scientific representative.

3 Introduction to the KingFisher mL

3.1 Intended use

The KingFisher mL magnetic particle processor (*Fig. 3.1*) is intended for professional research use by trained personnel. The instrument is intended for automated transfer and processing of magnetic particles in a 1 ml tube scale. Use for self-testing is excluded. It is recommended that Good laboratory Practices (GLP) are followed to guarantee reliable analyses.

3.2 Principle of operation

The KingFisher mL magnetic particle processor (*Fig. 3.1*) is designed for the automated transfer and processing of magnetic particles in a 1 ml tube well scale.

The principle of the KingFisher mL system (patents pending) is based on the use of magnetic rods covered with disposable, specially designed tip combs and tube strips. The instrument functions without any dispensing or aspiration parts or devices.

Samples and reagents including magnetic particles are dispensed into the tubes according to the corresponding kit instructions. The steps of the protocol that is selected by the user via the keypad and display have already been preloaded in the on-board software.



Fig. 3.1 KingFisher mL magnetic particle processor

3.3 Advantages of processing magnetic particles with the KingFisher mL

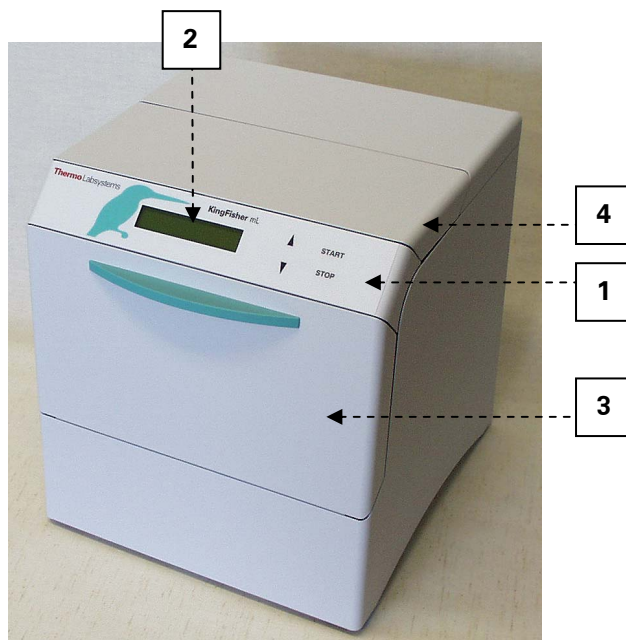
The KingFisher mL magnetic particle processor provides several advantages relating mainly to the principle of operation in that it:

- Enables automation of complicated manual steps
- Enables simultaneous processing and purification
- Enables a quicker reaction and a more efficient wash due to the simultaneous shaking during incubation and washing
- Facilitates a good collection of bead-bound sample due to the efficiency of the magnet
- Prevents carry-over due to the fact that the same tip is only used for one sample
- Facilitates the whole processing with the aid of an internal program.

4 Functional Description

4.1 Instrument layout

4.1.1 Front view



- 1 Keypad
- 2 Display
- 3 Front lid
- 4 Top lid

Fig. 4.1 KingFisher mL front view with lids closed

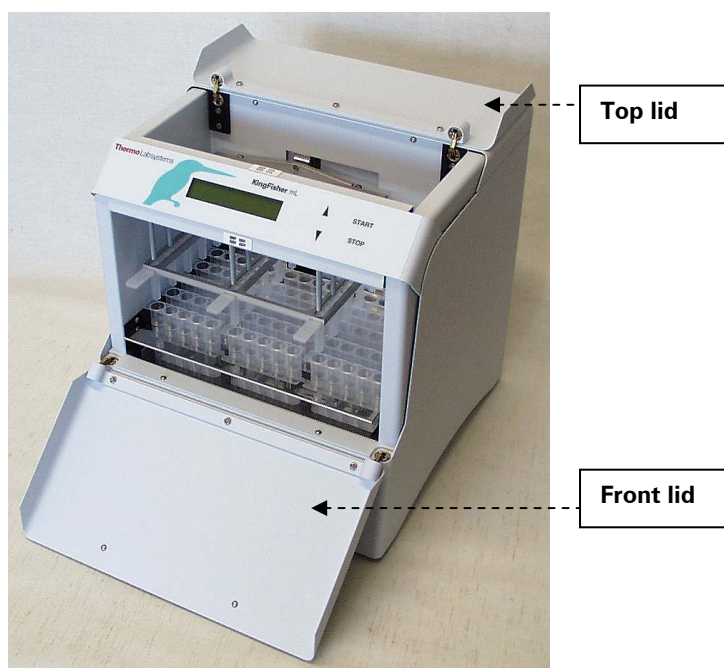
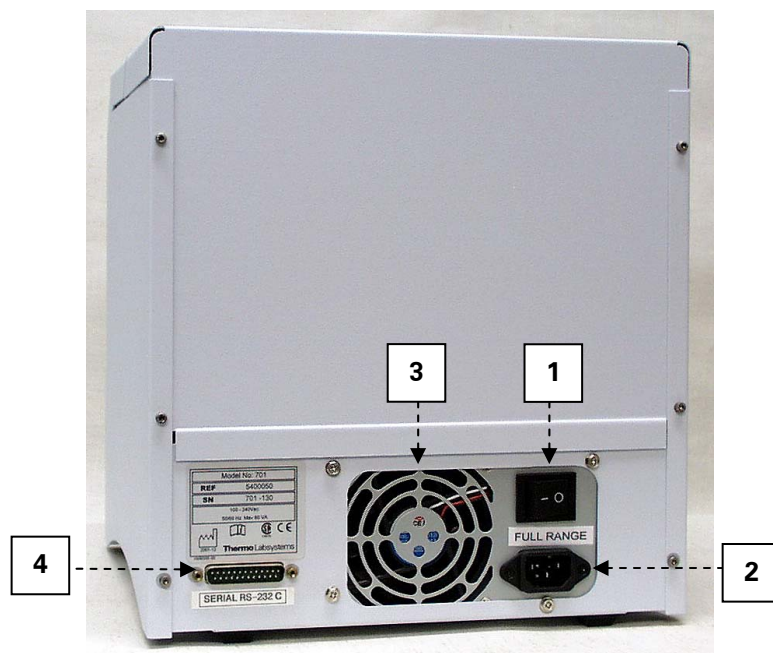


Fig. 4.2 KingFisher mL front view with lids open

4.1.2 Back view



- 1 ON/OFF switch
- 2 Mains power supply socket
- 3 Cooling-air outlet
- 4 Serial port RS-232C

Fig. 4.3 KingFisher mL rear view

4.1.3 Internal view



- 1 Tube-strip tray with tubes
- 2 Tip comb holder
- 3 Tip comb slot with tip comb
- 4 Fixed magnetic rod holder

Fig. 4.4 KingFisher mL internal view

4.2 KingFisher mL magnetic particle processor

The KingFisher mL instrument (*Fig. 4.5*) is designed for maximum 15 tube strips, which are compatible with the tip comb. The tube strip(s) is (are) kept stationary and the only moving assembly is the processing head with tip combs and magnetic rods. The head consists of two vertically moving platforms. One is needed for the magnetic rods (3 x 5 pieces) and the other one for the plastic tip combs (*Fig. 4.4*).

One tray contains fifteen separate tube strips and one sample processing uses one tube strip containing five tubes. One tip comb containing five tips is used for processing five samples at a time.

Before starting the magnetic particle processing via the keypad and display, the samples and reagents are dispensed into the tubes and the tip comb(s) is (are) loaded into its (their) slot(s). The tube strip(s) is (are) placed into the removable tray in the correct position and the tray is pushed into the end position. During the operation, the front and top lids can be closed (*Fig. 4.1*) or open (*Fig. 4.2*). Closed lids protect the processing against environmental contamination.

The operating principle employed is MPP (*inverse magnetic particle processing*) technology (*Fig. 4.6*). Rather than moving the liquids, the magnetic particles are moved from tube to tube containing specific reagents, in contrast to the external magnet method. Magnetic particles are transferred with the aid of magnetic rods covered with disposable, specially designed plastic tip combs.



Fig. 4.5 KingFisher mL magnetic particle processor

4.3 Principle of magnetic particle processing

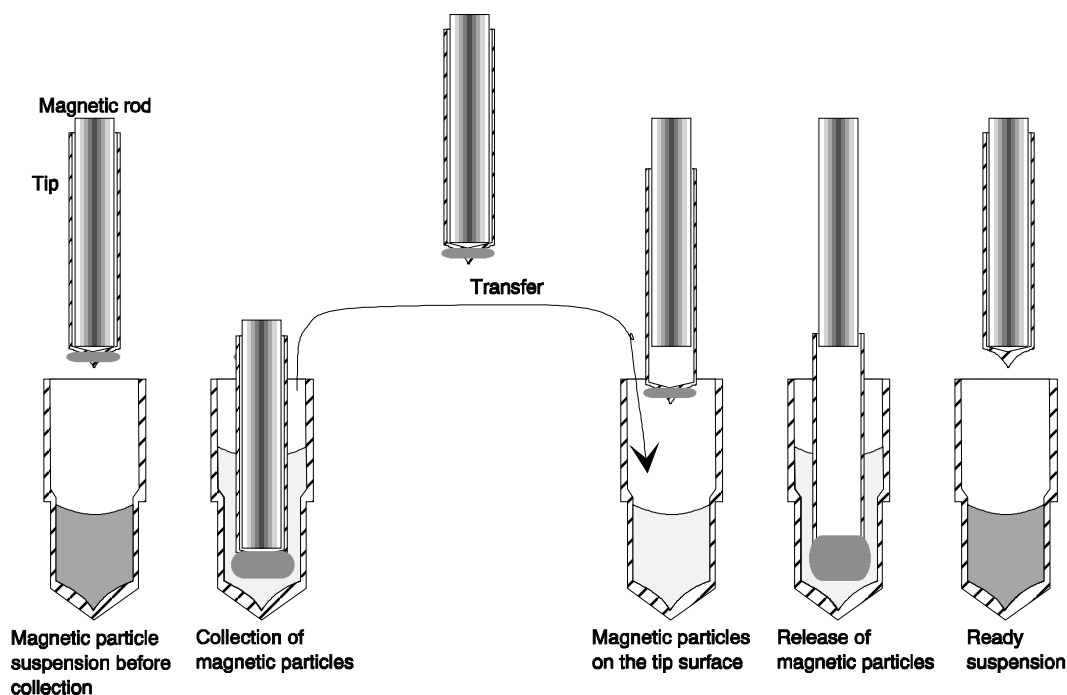


Fig. 4.6 Inverse magnetic particle processing

4.4 Working with a magnetic rod

Working with magnetic particles can be divided into five separate processes:

- collecting particles
- releasing particles
- washing particles
- incubation
- concentration

4.4.1 Collecting magnetic particles

During the collection of the magnetic particles, the magnetic rod (bar) is fully inside the tip. The magnetic rods together with the tip comb(s) move slowly up and down in the tubes and the magnetic particles are collected onto the wall of the tips. The magnetic rods together with the tip comb(s), having collected the magnetic particles, can be lifted out of the tubes and transferred into the next tubes.

4.4.2 Releasing magnetic particles

After collection of the magnetic particles, the magnetic rods together with the tip comb(s) are lifted from the tubes, the magnetic rods are lifted off and the tip comb(s) is (are) lowered into the next tubes containing a reagent.

Magnetic particles are released by moving the tip comb(s) up and down several times at considerably high speed until all the particles have been mixed with the substance in the next reaction.

4.4.3 Washing magnetic particles

Washing the magnetic particles is a frequent and an important processing phase. Washing is a combination of the release and collection processes in a tube filled with washing solution.

To maximize washing efficiency, the magnetic rods together with the tip comb(s) are designed to have minimized liquid-carrying properties.

4.4.4 Incubation

To keep the magnetic particle suspension evenly mixed in long-running reactions, the tip comb(s) can be moved up and down from time to time.

4.4.5 Changing the volume during the magnetic particle processing

The volume of the first tube can be larger than the volume of the next tube and this is used for *concentration* purposes (see Fig. 4.7 below).

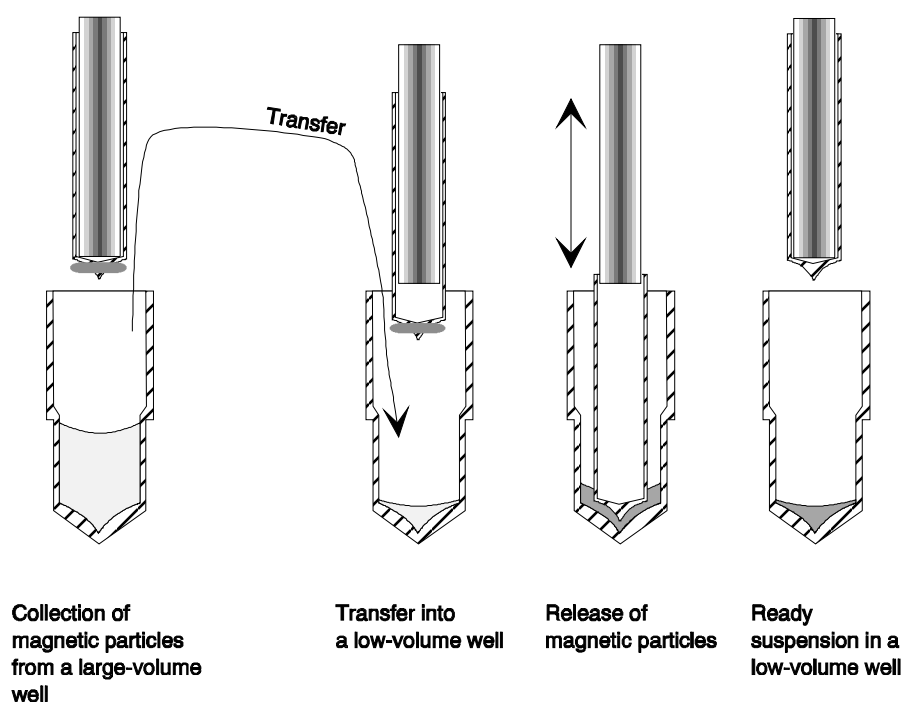


Fig. 4.7 A concentration step during magnetic particle processing

5 Installation

5.1 What to do upon delivery

5.1.1 How to unpack

Move the unpacked instrument to its site of operation. Unpack the KingFisher mL instrument and accessories carefully with the arrows on the transport package pointing upwards. The following notes and instructions are sent with the instrument and are immediately available when you open the package:

- packing instructions/packing list
- Thermo Scientific KingFisher mL Feedback Form
- Warranty Certificate card
- *KingFisher mL User Manual*.



Caution: Do not touch or loosen any screws or parts other than those specially designated in the instructions. Doing so might cause misalignment and will void the instrument warranty.

To lift the instrument, put your fingers under the bottom on either sides and lift it with your back straight.

Retain the original packaging for future transportation. The packaging is designed to assure safe transport and minimize transit damage. Use of alternative packaging materials may invalidate the warranty. Also retain all instrument-related documentation provided by the manufacturer for future use.

5.1.2 Checking delivery for completeness

Check the enclosed packing list against order. If any parts are missing, contact your local Thermo Fisher Scientific representative or Thermo Fisher Scientific Oy.

5.1.3 Checking for damage during transport

Visually inspect the transport package, the instrument and the accessories for any possible transport damage.

If the carton has been damaged in transit, it is particularly important that you retain it for inspection by the carrier in case there has also been damage to the instrument.

Visually check all interconnections in the basic instrument. Check that there are no loose parts inside the instrument.

If any parts are damaged, contact your local Thermo Fisher Scientific representative or Thermo Fisher Scientific Oy.

5.1.4 Environmental requirements

When you set up your KingFisher mL, avoid sites of operation with excess dust, vibrations, strong magnetic fields, direct sunlight, draft, excessive moisture or large temperature fluctuations.

- Make sure the working area is flat, dry, clean and vibration-proof and leave additional room for cables, lids, etc.
- Make sure the ambient air is clean and free of corrosive vapors, smoke and dust.
- Make sure the ambient temperature range is between +10°C (50°F) and +40°C (104°F).
- Make sure relative humidity is between 10% and 90% (non-condensing).

Leave sufficient space (at least 10 cm) at both sides and at the back of the unit to allow adequate air circulation.

The KingFisher mL does not produce operating noise at a level that would be harmful. No sound level measurements are required after installation.



Warning: Do not operate the instrument in an environment where potentially damaging liquids or gases are present.

5.1.5 Things to avoid

Do not smoke, eat or drink while using the KingFisher mL. Wash your hands thoroughly after handling test fluids. Observe normal laboratory procedures for handling potentially dangerous samples. Use proper protective clothing. Use disposable gloves. Be sure the working area is well-ventilated.

Never spill fluids in or on the equipment.



Warning: This product contains strong permanent magnets. People wearing a pacemaker or metallic prostheses should not use this product. A pacemaker or prostheses may be affected or damaged if it comes in very close contact with a strong magnetic field.

5.1.6 Technical prerequisites

Place the instrument on a normal laboratory bench. The net weight of the unit is approx. 10 kg (23 lbs.).

The instrument operates at voltages of 100 – 240 Vac and a frequency range of 50/60 Hz.

5.2 Setups before you put the instrument into operation

5.2.1 How to release the transport locks

Tools: 3 mm flat-head screwdriver (supplied with the instrument)

Note: THERE ARE FOUR TRANSPORT LOCKS (SCREWS) PRESENT (*Fig. 5.2*). A white tag with the red text '**CAUTION! Remove Transport Lock Before Use**' is fastened to the instrument to attract the user's attention (*Fig. 5.1*). The tag in *Fig. 5.1* and the black arrows in *Fig. 5.2* indicate where the transport locks are situated. Remove the tag and release the transport locks. Ensure that the transport locks have been released before you put the instrument into operation.



Fig. 5.1 KingFisher mL transport lock tag present before release of the locks

1. To remove the transport locks, unscrew the four screws (Fig. 5.2) with the 3 mm flat-head screwdriver supplied. Make sure that the screws are fully unscrewed in the opposite direction.
2. Remove the transport lock warning tag (Fig. 5.1).
3. Save the transport lock warning tag for future transportation.



Fig. 5.2 Location of the transport lock screws and warning tag

5.2.2 How to ensure startup



1. **Warning:** Ensure that the mains switch (*Fig. 4.3*) on the bottom left of the back panel is in the OFF position.



2. Connect the mains supply cable to the *mains power socket* (*Fig. 5.3*) at the bottom left of the back panel. If you need to use any other type of mains supply cable than supplied, use only cables certified by the local authorities. Before you plug in the power cable, ensure that the voltage on the rating label at the bottom left of the back panel (*Fig. 4.3*) corresponds to the local voltage.
3. Connect the instrument to a correctly installed line power outlet that has a protective conductor that is grounded.



Warning: Never operate your instrument from a power outlet that has no ground connection.



Fig. 5.3 Connecting the mains supply cable

5.3 Operational check

1. First switch the instrument ON. The instrument performs initialization tests and adjustments. The initialization tests are so-called self-tests.
2. The display quickly shows the internal software version and after that the display shows one of the protocols in the internal memory. This happens when the initialization tests and adjustments have been completed.
3. We recommend that you carry out an empty run (for example, a demonstration) to verify proper instrument operation.

6 Routine Operation

6.1 Switching on

Before you switch on the KingFisher mL, ensure that the voltage on the rating label at the bottom left of the back panel (*Fig. 4.3*) corresponds to the local voltage.

6.2 How to use the keypad and display

The liquid crystal display is a 20 character by 2 line LCD.

The lower line of the display can show the steps of the protocol, while the processing is running. The remaining time till the end of the process is shown in the top right corner. Furthermore, if errors occur, they will be displayed on the lower line.

The keypad and display are shown in *Fig. 6.1*.



Fig. 6.1 KingFisher mL keypad

The keys available are:

- *two cursor keys (or arrow keys, direction keys)*: to select the processing step to be run



To change to the previous/next protocol by scrolling up.



To change to the next protocol by scrolling down.

- *two function keys*

START To initiate the processing step.

STOP To pause/terminate the processing step. In short:
 STOP (paused)/START (the instrument continues after a Pause step)
 STOP (paused)/STOP (the processing is terminated).

6.3 How to START

- First of all slide the tube-strip tray out (*Fig. 6.2*).



Fig. 6.2 Loading the tube-strip tray

- A tube strip contains one sample and the reagents for a single processing. The user can select the position of the tube strip(s) on the tube-strip tray. The user must keep track of the sample order. However, the orientation of the tube strip is fixed. The tube strips have to be positioned so that the slip ends face left (*Fig. 6.3* and *Fig. 6.5*). Note that the order in the tube strip of the sample and reagents is determined by the kit used. Load only maximum fifteen special tube strips at a time into the tray for processing (*Fig. 6.5*). Slide the tray fully back in.

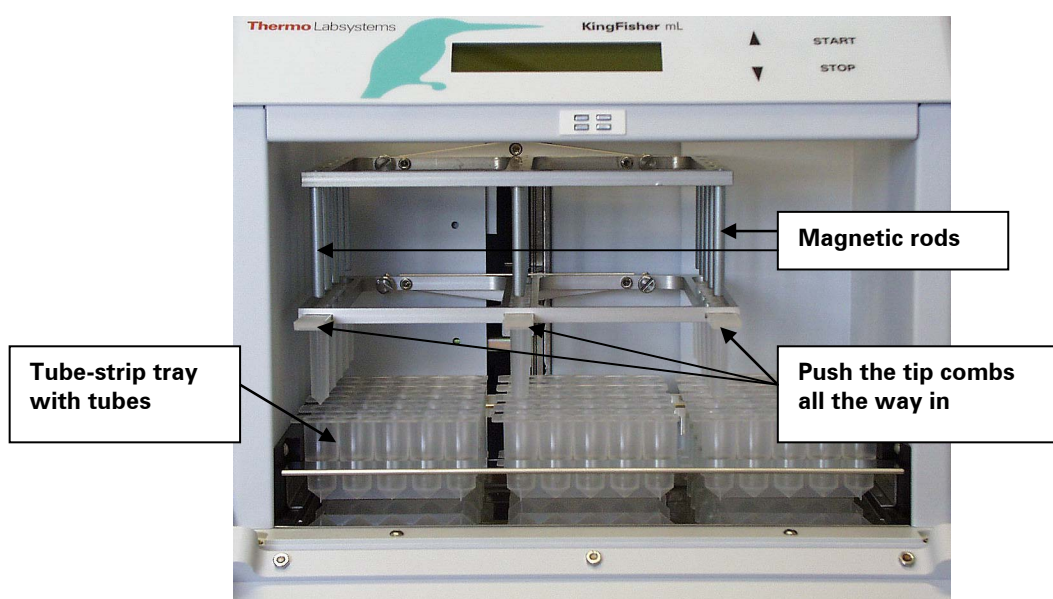


Fig. 6.3 Positioning of the tube-strip tray and insertion of the tip comb(s)

- The tip comb(s) always has (have) to be in its (their) slot(s) and fully inserted (*Fig. 6.3*). The instrument also functions with either one tube strip or one to fifteen tube strips depending on the amount of samples. At least one tip comb (*Fig. 6.4*) always has to be inserted into one of the narrow slots meant for the tip comb.

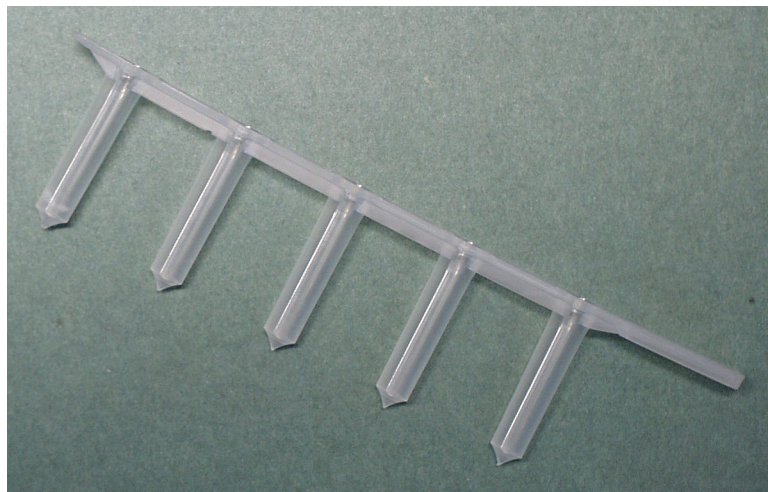


Fig. 6.4 Tip comb specially designed for the KingFisher mL

- Close the front lid if you want to. The front lid protects the instrument against environmental contamination.

Note: The front lid and/or top lid can be left open if desired. This action does not break the run.

- Select the name of the method.
- Use the **START** key to initiate the run.

For startup refer to Section 6.1 Switching on.

Note: The samples are collected immediately after the run and transferred into other tubes for longer storage.

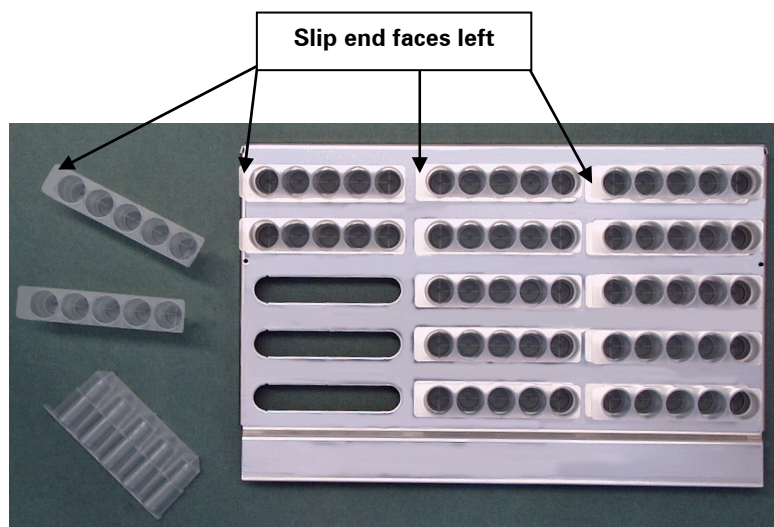


Fig. 6.5 Tube-strip tray containing tubes specially designed for the KingFisher mL

6.3.1 Usage of microtube tray in elution

The purification protocol can be chosen to use 2 ml microcentrifuge tubes in the elution step. Smaller tubes (e.g. 1.7 ml) cannot be used as they might get stuck in the tip comb.

If elution in microcentrifuge tubes is chosen in the KingFisher Software protocol with the KingFisher mL, the microtube tray is positioned on the empty KingFisher mL tray tube strip (*Fig. 6.6*). Microcentrifuge tubes are inserted and filled with buffer. This double-tray pack is then inserted into the instrument after removing the previous tray (see *Fig. 6.6*).

When the instrument pauses and **CHANGE TRAY** is displayed, the first tray is removed from the instrument and the second tray is inserted. The protocol continues by pressing the **START** key.

The following tube models have been tested and are recommended: Elkay 1.9 ml (Cat. no. 000-micr-190) and Costar 2.0 ml (Cat. no. 3213).

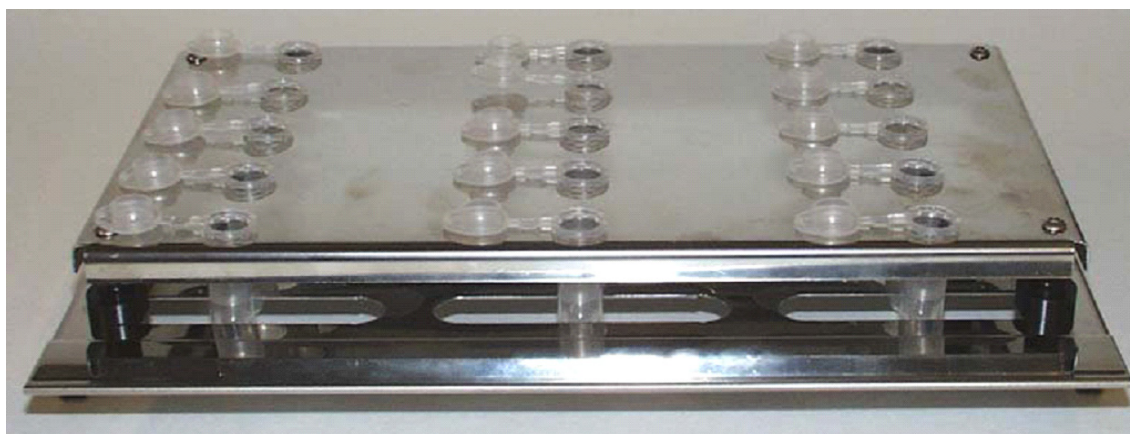


Fig. 6.6 Microtube/KingFisher mL double tray

Note: The microcentrifuge tubes should be inserted into the tray so that the caps are so near to the surface of the tray as possible and are all pointing right. This allows free tip comb movements.

6.4 Shutdown

Switch the KingFisher mL off by pressing the power switch (*Fig. 4.3*) at the bottom left of the back panel of the instrument into the OFF position.



Remove any tube strips still on the tube-strip tray. Dispose of all tube strips as biohazardous waste.



Remove any tip combs still in the instrument. Dispose of tip combs as biohazardous waste.

Wipe the tray surface and the adjacent instrument surface with a soft cloth or tissue paper moistened with distilled water, a mild detergent (SDS, sodium dodecyl sulfate) or soap solution.

If you have spilt infectious agents on the tray, disinfect with 70% alcohol or some other disinfectant (see Section 7.3 Decontamination procedure).

7 Maintenance

7.1 Regular and preventive maintenance

For reliable daily operation keep the instrument free of dust and liquid spills.

Abrasive cleaning agents are not recommended, because they are likely to damage the paint finish.

We recommend you clean the case of the instrument periodically to maintain its good appearance. A soft cloth dampened in a warm, mild detergent solution will be sufficient.

Clean the outside of the instrument and the tube-strip tray with clean low-pressure compressed air or a cloth dampened with water or a mild detergent when necessary.

Although the KingFisher mL is constructed from high-quality materials, you must immediately wipe away spilt saline solutions, solvents, acids or alkaline solutions from outer surfaces to prevent damage.



Painted surfaces can be cleaned with most laboratory detergents. Dilute the cleaning agent as recommended by the manufacturer. Do not expose painted surfaces to concentrated acids or alcohols for prolonged periods of time as damage may occur.

Clean the display areas with a mild laboratory detergent. The keypad has a wipe-clean surface.

Plastic covers and surfaces can be cleaned with a mild laboratory detergent or alcohol.



If any surfaces have been contaminated with biohazardous material, a mild sterilizing solution should be used.



Do not autoclave any part of this instrument except for the tube-strip tray.

7.1.1 How to clean the tube-strip tray

Keep the tube-strip tray surface clean to avoid dust and dirt entering into the instrument. Clean the tray surface at least once a week using a soft cloth or tissue paper soaked in a mild detergent solution (SDS), soap solution or alcohol.

If you have spilt infectious agents on the tray, you can autoclave the tray.

7.1.2 How to clean the magnetic rods

If required, wipe the magnetic rods using a soft cloth or tissue paper soaked in a mild detergent solution (SDS), soap solution or alcohol.

7.1.3 How to clean the processing chamber

If required, clean the processing chamber using a soft cloth or tissue paper soaked in a mild detergent solution (SDS), soap solution or alcohol.

7.2 Disposal of materials

Refer to local regulations for the disposal of infectious material.



The samples can be potentially infectious. Dispose of all used tube strips, disposable gloves, syringes, disposable tips, etc., as biohazardous waste.

7.3 Decontamination procedure

If there is any risk of contamination with biohazardous material, the procedure recommended below or some other corresponding decontamination procedure must be performed.

We strongly recommend that the complete decontamination procedure is performed before relocating the instrument from one laboratory to another.

Decontamination is not required for the proper functioning of the instrument.

Example of decontaminants

- Ethanol 70%
- Virkon solution 1 – 3%
- Glutaraldehyde solution 4%
- Chloramine T
- Microcide SQ 1:64



Always use disposable gloves and protective clothing and operate in a well-ventilated area.

1. Prepare the decontaminant: for example, 1 – 3% Virkon solution, or 200 ml 4% glutaraldehyde solution (or another agent recommended by your safety officer).
2. Empty the tube-strip tray.
3. Switch OFF the power and disconnect the mains supply cable (*Fig. 5.3*).
4. Disinfect the outside of the instrument using a cloth dampened with 70% ethanol.
5. Place the instrument in a large plastic bag. Ensure that the lids are open.
6. Place a cloth soaked in the prepared solution into the bag. Ensure that the cloth does not make contact with the instrument.
7. Close the bag firmly and leave the instrument in the bag for at least 24 hours.
8. Remove the instrument from the bag.
9. Clean the instrument using a mild detergent.
10. Remove any stains using 70% ethanol.

11. After performing this decontamination procedure, enclose a signed and dated Certificate of Decontamination both inside the transport package and attached to the outside of the package.

7.4 How to pack for service

When you ship the instrument for service remember to:

- Inform about the use of hazardous materials.
- Decontaminate the instrument beforehand.
- Install the transport locks.
- Pack the instrument according to the enclosed packing instructions.
- Use the original packaging to ensure that no damage will occur to the instrument during shipping. Any damage will incur additional labor charges.
- Enclose a dated and signed Certificate of Decontamination (see Section 9.4.1.1) both inside and attached to the outside of the package, in which you return your instrument (or other items).
- Enclose the return authorization number (RGA) given by the Thermo Fisher Scientific representative.
- Indicate the fault after you have been in touch with your local Thermo Fisher Scientific representative or the Thermo Fisher Scientific service department.

See Section 8.1 for details on storage and transportation temperatures.

7.5 Service contracts

We recommend you maintain and service the instrument regularly every twelve months on a contract basis by the manufacturer's trained service engineers. This will ensure that the product is properly maintained and gives trouble-free service. Contact the Thermo Fisher Scientific service department for more details.

7.6 Disposal of the instrument



- Decontaminate the instrument prior to disposal. See Sections 7.3, 9.4.1 and 9.4.1.1 on decontamination.
- Dispose of the instrument according to the legislation stipulated by the local authorities concerning take-back of electronic equipment and waste. The proposals for the procedures vary by country.



Thermo Fisher Scientific has contracted with one or more recycling/disposal companies in each EU Member State (European Country), and this product should be disposed of or recycled through them. Further information on Thermo Fisher Scientific's compliance with these Directives, the recyclers in your country, and information on Thermo Scientific products which may assist the detection of substances subject to the RoHS Directive are available at www.thermo.com/WEEERoHS.

- Regarding the original packaging and packing materials, use the recycling operators known to you.
- For further information, contact your local Thermo Fisher Scientific representative.

8 Technical Specifications

8.1 General specifications

Technical specifications

Overall dimensions	ca. 290 mm (11.4 in.) (W) x 290 mm (11.4 in.) (D) x 310 mm (12.2 in.) (H)
Weight	ca. 10 kg (23 lbs.)
Operating conditions (indoor use)	+10°C – +40°C, RH: 90% max.
Transportation conditions	-40°C – +70°C, packed in transport packaging
Storage conditions	-25°C – +50°C, packed in transport packaging
Mains power supply	100 – 240 Vac, 50/60 Hz, nominal Automatic voltage detection
Power consumption	80 VA max., 50 VA standby
Heat dissipation	273 BTU max.
Computer interface	Serial RS-232C port. Baud rate 9600. Character format 1 start bit, 8 data bits, 1 stop bit, no parity. Flow control XON/XOFF.

Performance specifications

Tube-strip tray (autoclavable)	Special design, 3 x 5 format
Magnetic rods (fixed)	3 x 5 format
Capacity	15 samples/run (3 x 5)
Vessel volumes	50 µl – 1 ml
Vessel type (disposable)	Special tube strip, 1 x 5 tubes
Tip comb (disposable)	Special design, 1 x 5 format
Magnetic particle size	ca. > 1 µm
Collecting efficiency	95%/vessel (washing buffer)
Incubating temperature	Ambient temperature (i.e., no incubation)
Keyboard / Display	START/STOP/two cursor keys/LCD
Software	Fixed internal protocols (loading through the external PC). Thermo Scientific KingFisher Software can be used to create and modify protocols.

8.2 Safety specifications

The KingFisher mL fulfills the following requirements:

EN 61010-1:1993 + A2:1995/IEC 61010-1:1990 + A1:1992 + A2:1995

The safety specifications are also met under the following environmental conditions in addition to or in excess of those stated in the operating conditions:

Altitude	up to 2000 m
Temperature	+5°C – +40°C
Mains supply fluctuations	± 10% from nominal
Installation category (overvoltage category)	II according to IEC 60664-1 (see Note 1)
Pollution degree	2 according to IEC 60664-1 (see Note 2)

Notes

1. The *installation category* (overvoltage category) defines the level of transient overvoltage, which the instrument is designed to withstand safely. It depends on the nature of the electricity supply and its means of overvoltage protection. For example, in CAT II, which is the category used for instruments in installations supplied from a supply comparable to public mains, such as hospital and research laboratories and most industrial laboratories, the expected transient overvoltage is 2500 V for a 230 V supply and 1500 V for a 120 V supply.
2. The *pollution degree* describes the amount of conductive pollution present in the operating environment. Pollution degree 2 assumes that normally only nonconductive pollution, such as dust, occurs with the exception of occasional conductivity caused by condensation.

Both of these affect the dimensioning of the electrical insulation within the instrument.

8.3 In conformity with the requirements

The KingFisher mL bears the following markings:

Type 701
100 – 240 Vac 50/60 Hz, 80 VA
CE mark
CSA monogram

The KingFisher mL conforms to the following requirements:

2006/95/EC (Low Voltage Directive)
2004/108/EC (Electromagnetic Compatibility Directive, EMC)
2002/96/EC (Waste of Electrical and Electronic Equipment)

Safety performance:

EN 61010-1:1993 + A2:1995/IEC 61010-1:1990 + A1:1992 + A2:1995
including CENELEC Common Modifications, US and CA National differences
-Installation Category (Overvoltage Category) II; Pollution Degree 2
EN 61010-1:2001 (Ed. 2)

EMC performance:

EN 50081-1:1992	Generic emission standard. Residential, commercial and light industry.
EN 50082-1:1997	Generic immunity standard. Residential, commercial and light industry.
EN 61326-1:1997 + A1:1998 + A2:2001 + A3:2003	Product family standard.

Test standards

Performance limits

EN 55022:1998	Class B, 150 kHz – 1 GHz
EN 61000-3-2:1995 + A1:1998 + A2:1998 + A13:1997 + A14:2000 EN 61000-3-3:1995	Class A
EN 61000-4-2:1995 + A1:1998	4 kV CD, 8 kV AD, Criteria B
EN 61000-4-3:1996 + A1:1998	3 V/m, 80 MHz – 2 GHz, Criteria A
EN 61000-4-4:1995	1 kV, Criteria B
EN 61000-4-5:1995	2 kV line to ground, 1 kV line to line, Criteria B
EN 61000-4-6:1996	3 V _{rms} , 150 kHz – 80 MHz, Criteria A
EN 61000-4-11:1994	30%/10 ms, Criteria B 60%/100 ms, Criteria C 100%/20 ms, Criteria B > 95%/5 s, Criteria C

9 Troubleshooting Guide

9.1 Error messages and warnings

<i>Error message</i>	<i>Warning</i>
Command error Cannot move track Head position error Head timeout Track position error Track timeout Magnets pos. error Magnets timeout Serial number error Serialno already set Too high pos. offset Invalid parameter Out of memory No acceleration tbl NV parameters lost Progmem full	Time already expired

9.2 Troubleshooting guide

Error	Cause	Action
Cannot move track	The track cannot be moved because the head is not up.	Press STOP . If the fault cannot be remedied, then switch the power on and off.
Head position error	The calculated head position does not agree with the head home sensor signal.	Check that there is nothing preventing the head from moving up and down.
Head timeout	The head has not arrived at the home sensor within the allowed time.	Check that there is nothing preventing the head from moving up and down.
Track position error	The calculated track position does not agree with the track home sensor signal.	Check that there is nothing preventing the track from moving left or right.
Track timeout	The track has not arrived at the home sensor within the allowed time.	Check that there is nothing preventing the track from moving left or right.
Magnets position error	The calculated magnetic rod position does not agree with the rod home sensor signal.	Check that there is nothing preventing the magnetic rods from moving up and down. Ensure that the tip comb is fully in.
Magnets timeout	The magnetic rods have not arrived at the home sensor within the allowed time.	Check that there is nothing preventing the magnetic rods from moving up and down.
Nonvolatile parameters lost	The nonvolatile parameters of the instrument are lost.	Contact authorized service.

9.3 Frequently asked questions (FAQ) about KingFisher mL processing

Q1: What does the KingFisher mL system do?

A1: Thermo Fisher Scientific now offers a complete KingFisher mL system – the magnetic particle processor – for the purification and processing of DNA, RNA, cells and proteins. The processor handles particles automatically according to the preloaded purification protocols. For more information on these applications, contact Thermo Fisher Scientific Oy.

Q2: Do I need a PC to control the processor?

A2: Normally the protocol(s) is/are preloaded by Thermo Fisher Scientific and no PC is needed for routine use. KingFisher Software can be used to create new protocols or modify existing protocols if needed. PC-modified protocols are then downloaded into the processor for routine use.

Q3: Will the magnet get weaker? If so, how long can they be used? Can they be replaced?

A3: The KingFisher mL magnets are made of material that is very stable. The magnetic field will not get weaker.

Magnets can be replaced if needed. Usually there is no need to replace them, but it can be done, for example, if they are mechanically damaged during runs. Changing the magnets can be carried out by local technical personnel.

Q4: How strong are the magnets? Can they, for example, disturb some sensitive equipment?

A4: The magnetic field is situated just around the magnetic tips, so it is very local. Therefore, the magnetic field should not cause any problems to surrounding equipment.

However, if you touch your floppy disk with the magnetic rod, you will lose all your data. Generally, a normal PC screen may also detect the magnetic field with a minor distortion of the image.

Q5: What should I do if I forget to insert the tip combs into the tip comb holder and the magnetic particles stick to the magnetic rods?

A5: Wipe the magnetic rods with a soft cloth or tissue paper soaked in a mild detergent solution, soap solution or alcohol.

Q6: What if the magnetic particles remained in the sample well?

A6: If the starting material is too viscous, the magnetic rods will not be able to collect the particles. Dilute the sample and check that the sample is properly homogenized/lysed.

Q7: What if the magnetic particles are attached to the tip combs?

A7: This happens sometimes but it will not affect the yield because the sample has been released from the particles.

Q8: Are the volumes of reagents in each well critical?

A8: We strongly recommend that you keep the given volumes within certain limits to avoid failure in performance of the chemical reactions and the processor.

9.4 Service request protocol

If the KingFisher mL requires service, contact your local Thermo Fisher Scientific representative or the Thermo Fisher Scientific service department. Do not under any circumstances send the instrument for service without any prior contact. It is imperative to indicate the fault and nature of the required service. This will ensure a faster return of the instrument to the customer.

The Thermo Fisher Scientific representative or distributor will take care of sending a complaint form (that is, the Warranty Claim Technical Sheet) to the Thermo Fisher Scientific service department. The Warranty Claim Technical Sheet contains a more detailed description of the fault, symptom or condition. Give all the necessary information to the distributor, who will fill in and forward the Warranty Claim Technical Sheet to the Thermo Fisher Scientific service department.

Check Section 7.4 How to pack for service. You will find instructions on how to proceed before shipping the instrument for service.

Check that any necessary decontamination procedure has been carried out before packing. See Sections 9.4.1 and 9.4.1.1 on decontamination. Refer also to Section 7.3 Decontamination procedure. Ensure that the Certificate of Decontamination as well as the return authorization number (RGA) are sent with the instrument.

The Thermo Fisher Scientific service department will keep you up to date with the progress of service and provide you with any further details you might need, for example, on maintenance, serviceability, troubleshooting and replacement.

9.4.1 Decontamination

Decontamination should be performed in accordance with normal laboratory procedures. Any decontamination instructions provided with the reagents used should be followed.

A decontamination procedure is only recommendable when infectious substances have been in direct contact with any part(s) of the instrument.

9.4.1.1 Certificate of Decontamination

The decontamination procedure is required prior to shipping the instrument to Thermo Fisher Scientific Oy, e.g., for repair. If, for any reason, the instrument is shipped back to Thermo Fisher Scientific Oy, it must be accompanied by a dated and signed Certificate of Decontamination, which must be attached to the outside of the package containing the instrument. See Section 7.3 Decontamination procedure.

Failure to confirm decontamination will incur additional labor charges or at worst the items will be returned for proper cleaning.

Before returning any instrument(s) or item(s), ensure that they are fully decontaminated. Confirm A or B status:

Name: _____

Address: _____

Tel./Fax: _____

Name: _____ Serial no.: _____

A)

I confirm that the returned items have not been contaminated by body fluids, toxic, carcinogenic or radioactive materials or any other hazardous materials.

B)

I confirm that the returned items have been decontaminated and can be handled without exposing the personnel to health hazards.

Materials used in the unit: Chemicals + Biological • Radioactive *)

Specific information
about contaminants: _____

Decontamination
procedure¹: _____

Date and place: _____

Signature: _____

Name (block capitals): _____

*) The signature of a Radiation Safety Officer is also required when the unit has been used with radioactive materials.

This unit is certified by the undersigned to be free of radioactive contamination.

Date and place: _____

Signature: _____

Name (block capitals): _____

¹ Please include decontaminating solution used.

PHOTOCOPIABLE

10 Warranty Certificate

Thermo Fisher Scientific Microplate Instrumentation Business products are fully guaranteed against defective parts and materials, including defects caused by poor workmanship, for a period of one year from the date of delivery.

Thermo Fisher Scientific will repair or replace defective parts or materials during the term of warranty at no extra charge for materials and labor provided that the products were used and maintained in accordance with Thermo Fisher Scientific's instructions. The warranty is invalid if products have been misused or abused.

For the warranty to be effective, the product must have been purchased either directly from Thermo Fisher Scientific or from an authorized Thermo Fisher Scientific distributor. The guarantee is not transferable to a third party without prior written approval from Thermo Fisher Scientific.

This guarantee is subject to the following exclusions:

- Any defects caused by normal wear and tear.
- Defects caused by fire, lightning, flood, earthquake, explosion, sabotage, war, riot, or any other occurrence of the type listed above.
- Refurbished products that are subject to different warranty conditions.

THIS WARRANTY IS IN LIEU OF ALL OTHER EXPRESSED OR IMPLIED WARRANTIES, INCLUDING BUT NOT LIMITED TO ANY IMPLIED WARRANTIES OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE. The seller is not liable for any loss or damage arising out of or in connection with the use of the product or other indirect damages.

These warranty terms and conditions can be obtained from your local Thermo Fisher Scientific dealer.

10.1 Warranty limitations

- Consumables are not included in the warranty.
- The warranty does not extend beyond the expiration date of the reagent kit.

11 Ordering Information

Code	Instrument/System
-------------	--------------------------

5400050	KingFisher mL, 100 – 240 V
---------	----------------------------

11.1 List of accessories

Code	Item	Quantity
1508260	<i>KingFisher mL User Manual</i>	1
97 002 111	KingFisher mL tip comb	800 pcs/bag
97 002 121	KingFisher mL tube	900 pcs/bag (15 x 60 pcs)
97 002 131	KingFisher mL Combi 60 (tubes and tip combs for 60 samples)	box
97 002 141	KingFisher mL Combi 240 (tubes and tip combs for 240 samples)	box
4540000	FP-Stepper 10 µl – 5 ml	1
9404190	Finntip 2.5 ml	100 pcs/box

11.2 List of spare parts

Code	Item	Quantity
11273100	Tube-strip tray = Tube rack	1
24073180	Microtube tray	1
2305290	Serial cable F9/F25 (for RS-232C port)	1
5186980	KingFisher Software, version 1.0	1

11.3 List of recommended spare parts

Code	Item *	1 – 2 unit(s) / year	10 units / year
1091910	Tooth belt T2.5/330		1
1092250	Tooth belt T2.5/265		1
2004460	PCB PSP-01		1
2004470	PCB CSU-02	1	2
2004480	PCB CSU-01		1
21040811	Power supply	1	2
21040820	H-motor assembly		1
21040850	M-motor assembly		1
24070740	Free wheel assembly	1	3
24072550	Magnetic plate assembly		1

* = Contact the Thermo Fisher Scientific service department

12 Glossary and Abbreviations

Magnetic rod	The rods which are magnetic and collect magnetic particles. The rods do not collect the particles on their own, the magnetic rods must always be protected by a tip comb.
Tip comb	Protects the magnetic rod. A disposable tip comb always has to be fitted into the tip comb holder slot during processing.
Tube strip	The disposable tubes where all the reagents and samples are located and where the processing takes place.
Tube-strip tray (Tube rack)	A detachable tube-strip holder or tube rack that can be placed on the laboratory bench to facilitate pipetting (<i>Fig. 6.3</i> and <i>Fig. 6.5</i>).

12.1 Keywords for web pages

affinity separation	PCR
amplification	PCR amplification
beads	PCR product
cloned DNA	post PCR purification
cloning	primer
DNA	probes
DNA complex	processing
DNA immobilization	processor
DNA purification	purification
DNA strand separation	RNA
DNA template	mRNA
double-stranded DNA	RT-PCR
dsDNA	separation
encapsulated magnetic microspheres	separator
genome	silica-coated beads
genomic DNA	silica-coated microspheres
hybride selection	silica-coated particles
KingFisher (or Kingfisher)	streptavidin-coated beads
magnetic beads	streptavidin-coated microspheres
magnetic microspheres	streptavidin-coated particles
magnetic particles	subtraction libraries
magnetic separation	superparamagnetic beads
microspheres	superparamagnetic microspheres
nucleotide	template
oligo-dT	Thermo Fisher Scientific
particles	Thermo Scientific

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15 Appendix A. Application Protocols for KingFisher mL

A.1 Example of a cell or protein purification protocol using KingFisher mL

Cell separation and protein purification based on paramagnetic particles can be automated with the KingFisher mL magnetic particle processor.

Generate a protocol with Thermo Scientific KingFisher Software for cell or protein isolation called **General_protocol** following the instructions below. Load the protocol into the internal memory of the instrument. Pipette the reagents into the Thermo Scientific KingFisher mL tube strips. After you have filled the KingFisher mL tube strips, insert the tray into the KingFisher mL processor and choose the correct program by using the arrow keys. The name of the KingFisher mL protocol is **General_Protocol2**. Start the protocol by pressing the **START** button. Purified cells or proteins are released into a 200 µl volume into the last tube 5.

The description of the KingFisher mL protocol is as follows:

Program

1. The sample is mixed with particles in tube 1.
2. The sample is incubated with particles for 3 minutes in tube 1.
3. The particles are washed in washing buffer in tube 2.
4. The particles are washed in washing buffer in tube 3.
5. The particles are washed in washing buffer in tube 4.
6. The purified sample is released into the elution buffer in tube 5.

Volumes

1. Sample + particles = **1000 µl**
2. Washing buffer **1000 µl**
3. Washing buffer **1000 µl**
4. Washing buffer **1000 µl**
5. Elution buffer **200 µl**

The protocol can be modified according to customer needs.

A.2 Example of a KingFisher mL total RNA purification protocol

The total RNA purification kit which is developed for the purification of total RNA from cultured cells or tissue homogenates can be used with this protocol. Normally the kit is based on paramagnetic silica particles that bind RNA in the presence of chaotropic salts. After the binding step, the RNA/magnetic particle complex is washed extensively in washing buffers to remove any remaining contaminants. Finally the purified RNA is released into the elution buffer. During the elution step the samples are heated outside the KingFisher mL instrument. The approximate processing time for 15 (KingFisher mL) samples is 40 minutes.

The KingFisher mL total RNA purification procedure provides high-quality RNA for direct use in downstream applications, such as RT-PCR and Northern blotting.

You can use ca. $2 - 5 \times 10^6$ cells or 30 mg of tissue to purify RNA. Yields will vary according to the kit, cell and tissue type used in the assay.

The name of the purification protocol is **Total_RNA_mL_1**.

Pipetting the reagents / KingFisher mL

Tube no.	KingFisher mL tube strip
1	1000 µl cell lysate OR tissue homogenate + 50 µl magnetic particles
2	Washing buffer 1 1000 µl
3	Washing buffer 2 1000 µl
4	Washing buffer 2 1000 µl
5	Elution buffer 200 µl

Protocol Total_RNA_mL_1

1. The sample is incubated with magnetic particles in tube 1 for 19 minutes.
2. The particles are washed in washing buffer 1 in tube 2.
3. The particles are washed in washing buffer 2 in tube 3.
4. The particles are washed in washing buffer 2 in tube 4.
5. The particles are released into the elution buffer in tube 5.
6. The protocol is paused for the heating step. The text marked "PAUSED" appears on the screen.
7. The tray is removed OR the samples are transferred into microtubes for the heat incubation.
8. After incubation, the tray is returned to the KingFisher mL OR the samples are transferred back into tube 5.
9. The protocol will continue after you press **START**.
10. The particles are removed and returned into tube 4.

Sample preparation from cultured cells

- Transfer the cultured cells suspended in PBS into a suitable centrifuge tube.
- Collect the cells by centrifugation for 5 minutes at 6000 rpm and discard the supernatant.
- If frozen cells are used, thaw the cells on ice, centrifuge the cells for 5 minutes at 1000 rpm and discard the supernatant.
- Add lysis buffer (including 10 μ l β -ME/1000 μ l buffer) so that the final concentration ranges from 2 – 5 $\times 10^6$ cells/1000 μ l.
- Mix well by vigorously pipetting up and down several times until the lysate is homogenous. Vortex for an additional 30 seconds at full speed.

Note: If a lot of foam has formed, keep the sample on ice while filling the KingFisher mL tube strips with other reagents or centrifuge the sample for 1 minute at 1000 g (~ 3000 rpm in a microcentrifuge).

Note: If a centrifuge is used, ensure that you follow the centrifugation instructions. Do not exceed the time and speed, as this will drop the RNA yield drastically.

- Mix the homogenate gently by pipetting and add the cell mixture into the KingFisher mL tube strip according to the instructions.

Sample preparation from tissue

- Weigh the needed amount of tissue (up to 30 mg) and add 900 μ l of lysis buffer (including 10 μ l β -ME/1000 ml buffer β -ME). Depending on the homogenizer and the tool used, change the amount of tissue and lysis buffer using the same ratio as above (see the instructions in the previous section).
- Homogenize the sample on ice in the presence of lysis buffer for 5 – 90 seconds, depending on the toughness of the tissue.
- Centrifuge the homogenate for 1 minute at 1000 g (~ 3000 rpm in a microcentrifuge).

Note: Ensure that you follow the centrifugation instructions. Do not exceed the time and speed, as this will drop the RNA yield drastically.

Note: If a rotor-stator homogenizer is not available, other homogenization methods can be used instead. For example, soft tissues, such as liver, can be homogenized using, for example, a pestle and needle/syringe method. Always follow the centrifugation instructions above, despite the homogenization method used.

- Add up to 1000 μ l of the supernatant into the KingFisher mL tube according to the instructions.

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- Switch the KingFisher mL on (p. 17).
- Pipette the samples, reagents and wash buffers into the tube strips (p. 19, 41).
- Load the tray(s) onto the instrument (p. 19).
- Insert the tip comb(s) into the tip comb holder slot(s) (p. 19).
- Select the desired protocol from the keypad and display (p. 18).
- Press the **START** button (p. 19).
- After the end of the run, remove the tube strips and tip combs (p. 21).
- Maintain your KingFisher mL instrument regularly (p. 22).



Fig. 16.1 KingFisher mL keypad

Please send to Thermo Fisher Scientific Oy: Fax +358-9-2910415

17 Appendix C. Thermo Scientific KingFisher mL Feedback Form

Cat. no. PURCHASED BY Company/Institute Department Address Tel. Fax Internet home page Date of purchase Your research area Dr. ☐ Mr. ☐ Mrs. ☐ Ms. ☐ Job title/Position Surname (block capitals) Internet e-mail address 	Serial no. PURCHASED FROM Distributor Address Tel. Date of delivery
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	Excellent	Above expectations	As expected	Below expectations	Comments
Reagent kit/Instructions	☐	☐	☐	☐	
Instrument/User manual	☐	☐	☐	☐	
Operational reliability	☐	☐	☐	☐	
Design	☐	☐	☐	☐	
Ease of use	☐	☐	☐	☐	
Operational costs	☐	☐	☐	☐	
Customer support	☐	☐	☐	☐	

Further instrument/system developments desired:

Further applications desired:

Where did you first learn about the product?

Would you like to receive information about other Thermo Scientific products?

18 Appendix D. Addresses

For the latest information on products and services, visit our websites at:

<http://www.thermo.com>

<http://www.thermo.com/kingfisher>

Manufactured by:

Thermo Fisher Scientific Oy
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19 Appendix E. Supplementary Application Protocols

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