PetNADTM

Anaplasma platys Detection Kit

User Manual

For Research Use Only

Manufacturer:

GeneReach Biotechnology Corporation

TEL: 886-4-24639869 FAX: 886-4-24638255

No. 19, Keyuan 2nd Road, Central Taiwan Science Park, Taichung City, Taiwan 407

Web Site: www.petnad.com

Content

IN'	FENDED USE	1
SC	IENTIFIC MEANINGS	1
SU	MMARY AND EXPLANATION	2
PR	INCIPLES OF THE PROCEDURE	3
PR	ODUCT DESCRIPTION	3
A.	Materials Provided	3
В.	Materials and Equipments Required, but Not Provided	4
C.	Storage and Stability	4
D.	Sample Type	4
RE	COMMENDED NUCLEIC ACID EXTRACTION MET	HODS
•••••		5
PR	ECAUTIONS	5
LII	MITATIONS	6
PR	OCEDURE	7
A.	PetNAD TM Anaplasma platys Detection Kit Quick Guide	7
B.	Procedure	8
DA	TA INTERPRETATION	10

PetNADTM Anaplasma platys Detection Kit

ANALYTICAL SENSITIVITY	10
TROUBLESHOOTING	11
REFERENCE	13

INTENDED USE

PetNADTM Anaplasma platys Detection Kit is intended for *in vitro* detection of the Anaplasma platys (A. platys) DNA based on insulated isothermal polymerase chain reaction (iiPCR) technology. This kit is designed specially to be used with an insulated isothermal (iiPCR)-compatible instrument, POCKITTM Nucleic Acid Analyzer. The assay is intended for use by veterinarians or technicians with basic laboratory skills.

This kit is intended for research use only.

SCIENTIFIC MEANINGS

Antibody induced by vaccine or obtained from maternal immunity could lead to false positive interpretation in antibody-based diagnostic procedures. Detecting pathogen's nucleic acids, not antibody, PCR-based methods can avoid the false positive results described above.

Furthermore, with higher analytical sensitivity, PCR can detect lower levels of viral signals than most if not all diagnostic methods. It can reduce the chance of false negative results at early infection stage and shorten the window period between time of infection and detection.

SUMMARY AND EXPLANATION

A. platys (formerly Ehrlichia platys) was described first as a Rickettsialike, platelet-specific organism in dogs with infectious canine cyclic thrombocytopenia (ICCT) in Florida, USA, in 1978. A. platys is spread by ticks, particularly the brown dog tick. A. platys-infected dogs could be co-infected with Ehrlichia canis, Babesia canis, or other vectorborne pathogens that share the same vector.

PCR is one of the most commonly accepted methods that provide high sensitivity and specificity for *A. platys* detection. However, conventional PCR assays could take three to four hours and require sophisticated thermocyclers and well-trained technicians to perform. GeneReach has developed **PetNAD**TM *Anaplasma platys* Detection Kit based on iiPCR technology, which significantly reduces reaction time and offers sensitivity and specificity comparable to those of conventional nested PCR (Tsai, 2012; Chang, 2012). Furthermore, this simple and easy assay is completed rapidly in a portable **POCKIT**TM Nucleic Acid Analyzer.

PRINCIPLES OF THE PROCEDURE

In iiPCR, hydrolysis probe-based chemistry is used to generate fluorescent signal during amplification of target DNA. The primers and probe target the *gltA* gene and do not cross-react with nucleic acid from host and other canine pathogens.

PRODUCT DESCRIPTION

A. Materials Provided (24 tests/kit)

Component	Contents or Purpose		Amount
Premix Pack		A. platys Premix (lyophilized	24 bags (1 A. platys
		pellet) containing dNTPs,	Premix vial and 1
		primers, probe, and enzyme	desiccating
		for amplification.	agent/bag)
		Desiccating agent pack.	
Premix Buffer		Reaction buffer to re-dissolve	2 vials (1.3 ml/vial)
В		the lyophilized pellet.	
P(+) Control		Dried plasmid containing A.	1 vial
		platys partial sequence as	
		positive control.	
P(+) Control		Reaction buffer to re-dissolve	1 vial (110 μl/vial)
Buffer		P(+) Control.	
R-tube			1 bag (24 pieces/bag)

Cap	1 bag (24 pieces/bag)
User Manual	1 copy

B. Materials and Equipment Required, but Not Provided

- PetNADTM Nucleic Acid Co-prep Kit or tacoTM Automatic Nucleic Acid Extraction System.
- POCKITTM Nucleic Acid Analyzer (POCKITTM): PetNADTMcompatible instrument.
- 3) **cubeeTM** Mini-Centrifuge (**cubeeTM**).
- 4) Micropipette and filter tips.

C. Storage and Stability

- 1) The kit should be stored at 4°C and is stable until the expiration date stated on the label.
- 2) Store Premix vials in sealed Premix Pack to avoid hydration of lyophilized components.
- 3) Reconstituted P(+) Control is stable for 6 months at 4°C. Aliquot reconstituted P(+) Control to avoid degradation of nucleic acid.

D. Sample Type

Nucleic acid extracted from whole blood.

RECOMMENDED NUCLEIC ACID EXTRACTION METHODS

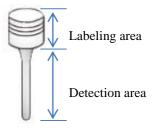
- A. PetNADTM Nucleic Acid Co-prep Kit.
- **B.** tacoTM DNA/RNA Extraction Kit, compatible instrument—tacoTM Automatic Nucleic Acid Extraction System.

Note: Please follow the instruction manual of above extraction methods to obtain optimal results. It is the user's responsibility to validate the combination of this reagent set with nucleic acids extracted by other methods for any particular application.

PRECAUTIONS

- A. Do not open R-tube(s) after reaction to prevent any carryover contamination.
- B. Perform extraction and amplification in two independent spaces to minimize contamination.
- C. Do not reuse R-tube and Premix.
- D. Include the P(+) Control to:
 - 1) Ensure **POCKIT**TM is working normally.
 - 2) Ensure detection kit performance after storage.

- E. To get optimal fluorescence detection.
 - Wear powder-free gloves to handle R-tubes.
 - Do not label in the detection area of R-tube.



LIMITATIONS

- A. The test should be used only for testing nucleic acid extracted from animal specimens. Do not add specimens (*e.g.* whole blood) directly into Premix.
- B. **PetNAD™** Nucleic Acid Co-prep Kit and **taco™ mini** Automatic Nucleic Acid Extraction System are recommended for nucleic acid extraction.
- C. Any deviations from the recommended procedure may lead to suboptimal results. Quality of the extracts should be validated by the users.
- D. For **PetNADTM** Anaplasma platys Detection Kit, it is strongly recommended to use freshly prepared nucleic acid (within 1 hour after extraction) to achieve optimal results.

PROCEDURE

A. PetNADTM Anaplasma platys Detection Kit Quick Guide



B. Procedure

Note: Before preparing the reactions for iiPCR testing, turn on POCKITTM to initiate the calibration for the instrument. The device will complete self-test within 5 minutes. Please refer to the user manual of POCKITTM for further details.

Note: Before using for the first time, add 100 μ l P(+) Control Buffer to P(+) Control. Store reconstituted P(+) Control at 4°C.

- 1) Label R-tube(s) in the label area.
- 2) Prepare one Premix for each sample. (Premix tube is in Premix Pack. Each Premix Pack contains one Premix tube.)

Note: When the pellet is not found at the bottom of the tube, spin tube briefly to bring it down.

- 3) Add 50 µl Premix Buffer B to each Premix tube.
- 4) Add 5 μl nucleic acid extract or P(+) Control to each Premix tube. Spin Premix tube for 10 seconds in a mini centrifuge (such as cubeeTM).
- 5) Transfer 50 µl Premix/sample mixture into R-tube.
- 6) Seal top of each R-tube with a cap. Make sure R-tube is capped tightly.
- 7) Place R-tube into the holder of **POCKIT**TM.
- 8) Spin tube briefly in **cubeeTM** to make sure all solution is

collected at the bottom of R-tube.

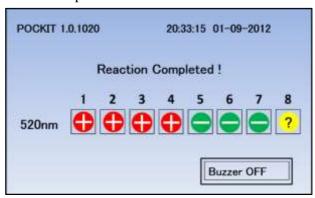
Note: Make sure there are no bubbles in the solution.

Note: Start reaction within 1 hour (to prevent nucleic acid degradation and non-specific reaction).

- 9) **POCKITTM** reaction:
 - a) Select "520 nm".
 - b) When "System READY" is displayed, place the holder with R-tube(s) into the reaction chamber.
 - c) Tap cap of each R-tube to make sure the tube is positioned properly.
- 10) Close lid and press "Run" to start reaction program.
- 11) Test results are shown on the monitor after reaction is completed.

DATA INTERPRETATION

* One example of results shown on the monitor.



520 nm	Interpretation		
•	A. platys Positive		
0	A. platys Negative		
?	Repeat reaction with freshly prepared nucleic		
•	acid.		

ANYLYTICAL SENSITIVITY

The detection limit of **PetNADTM** *Anaplasma platys* Detection Kit is about 10 copies/reaction.

TROUBLESHOOTING

Problems	Possible causes	Solutions
False Positive	1) Reuse of micro-	■ Micro-centrifuge tubes, tips, R-
	centrifuge tubes,	tubes and Premix are for single-use
	tips, R-tubes and	only. Reusing these accessories
	Premix.	would cause cross-contamination,
		and therefore false positive results.
		■ Used micro-centrifuge tubes, tips,
		R-tubes and Premix should be
		collected and discarded according
		to local regulation. Do not place the
		waste close to the working area to
		prevent cross-contamination.
	2) Contaminated	■ Use aerosol-free tips.
	micropipette	
	3) Contaminated	■ Consult with a GeneReach
	reagent	technical support representative or
		local distributor.
	4) Contaminated	■ Consult with a GeneReach
	working area	technical support representative on
		how to clean up working area.

Problems	Possible causes	Solutions
False	1) Nucleic acid	■ Consult manual of nucleic acid
Negative	extraction failed.	extraction kit.
	2) PCR inhibition	■ Do not overload PCR with too
		much nucleic acid.
		■ Spike nucleic acid sample (5 μl)
		into a P(+) Control reaction for a
		parallel PCR reaction. Negative
		results indicate the presence of
		inhibitors in the nucleic acid. In
		that case, prepare another nucleic
		acid extract.
Heavy	1) Leakage or spill of	■ Consult with a GeneReach
contamination	reaction from R-	technical support representative or
of amplicons	tube into reaction	local distributor.
in reaction	chamber of	
chamber of	POCKIT TM .	
POCKIT TM .		

REFERENCE

- 1. Arraga-Alvarado, C., Palmar, M., Parra, O. and Salas, P., (2003). *Ehrlichia platy* (*Anaplasma platy*) in dogs from Maracaibo, Venezuela: an ultrastructural study of experimental and natural infections. *Vet Pathol* 40:149-156.
- Beall, M.J., Chandrashekar, R., Eberts, M.D., Cyr, K.E., Diniz, P.P., Mainville, C., Hegarty, B.C., Crawford, J.M. and Breitschwerdt, E.B., (2008). Serological and molecular prevalence of *Borrelia burgdorferi*, *Anaplasma phagocytophilum*, and *Ehrlichia* species in dogs from Minnesota. *Vector Borne Zoonotic Dis* 8:455-464.
- 3. Chang, H.F. G., Tsai, Y.L., Tsai, C.F., Lin, C.K., Lee, P.Y., Teng, P.-H., Su, C. and Jeng, C.-C., (2012) A thermally baffled device for highly stabilized convective PCR. *Biotechnology Journal* 7(5): 662-666, doi: 10.1002/biot.201100453
- 4. Glaze, M.B., & Gaunt, S.D. et al. (1986). Uveitis associated with *Ehrlichia platys* infection in a dog. *J Am Vet Med Assoc* 189: 916-917.
- Olson, J.G., (1995). Ehrlichiosis. In: Zoonoses updates from the Journal of the American Veterinary Medical Association. American Veterinary Medical Association. Schaumburg, IL; 1995:74-75.
- 6. Tsai Y.L., Wang H.T. T., Chang H.F. G., Tsai C.F., Lin C.K., Teng P.H., Su C. and Jeng C.C., (2012). Development of TaqMan probe-based insulated isothermal PCR (iiPCR) for sensitive and specific on-site pathogen detection. *PLoS ONE* 7(9): e45278. doi:10.1371/journal.pone.0045278.