ELx808™

Operator's Manual

ELx808™

Absorbance Microplate Reader
Operator's Manual

Notices

BioTek® Instruments, Inc.

Highland Park, P.O. Box 998

Winooski, Vermont 05404-0998 USA

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Document Conventions

This manual uses the following typographic conventions.

Example	Description
	This icon calls attention to important **safety** notes.
Warning!	A **Warning** indicates the potential for bodily harm and tells you how to avoid the problem.
Caution:	A **Caution** indicates potential damage to the instrument and tells you how to avoid the problem.
DEFINE	Bold text in **COURIER** font represents menu options as they appear on the display of the ELx808.
Note:	**Bold text** is primarily used for emphasis.
	This icon calls attention to important information.

Revision History

Rev	Date	Changes
A	1/96	First Release
B	5/96	Revised reader specifications. Added Scanning method to software.
C	1/97	Added Panel and Reuse of Standard Curve, and revised serial port pin-out description.
D	2/98	Changed Chapter 4, Performance Verification to reflect changes to the Universal Test Plate. Added Liquid Tests 1 and 2. Changed Appendix C: Error Codes.
E	8/98	Added printer information. Updated Appendix B: Computer Control.
F	3/99	Changed European address. Corrected printer compatibility. Corrected the sequence of steps for processing a curve-fit method.
G	7/99	Added comment about blanking, P-Down and P-Across being inactive in this version of software. Clarified the need for at least one sample to be defined on a plate. Added scanning computer control commands.
H	5/00	Updated Chapter 4- Performance
		Verification to include a Liquid Test 3 to verify 340 nm instruments. Corrected the voltage range for Range 2 in the Introduction.
		Corrected Incubation Temperature Control range to 6 above ambient. Changed **Note** on page 4-7 dealing with air readings during Self-Test.
		Clarified Liquid Test 1. Changed Table 4-1 to include additional maintenance items. Reworked Liquid Test 2.
I	7/02	Updated contact information (pages iii, 1-8, 1-9, 2-8, 2-20, and 4-13).
		Added IQ/OQ/PQ procedures and updated liquid testing information (Chapter 4).
		Revised lamp replacement procedure (page 2-15).
J	11/03	Preface:
		- Updated contact information in Notices (page iii).
		- Added Document Conventions (page vi).
		- Updated Warnings section (pages vii and viii).
		- Updated Electromagnetic Compatibility section (pages ix and x).
		- Added the following safety symbols and text:
		➢ "Consult instructions for use" (page xii)
		➢ "In vitro diagnostic medical device" (page xii)
		➢ "Separate collection for (disposal of) electrical and electronic equipment" (page xiii).

Revision History (Cont'd)

Rev	Date	Changes
J	11/03	Preface (Cont.):
		- Expanded the Intended Use Statement (page xiv).
		Chapter 1:
		- Updated contact information in Technical Support.
		- Removed **About This Manual** section (page 1-5).
		- Added "Absorbance Test Plate" to the Optional Accessories list (page 1-8).
		Clarified lamp replacement procedure (Chapter 2, pages 2-14 to 2-16).
		Clarified description of cutoff formulas (Chapter 3, pages 3-42 to 3-44).
		Chapter 4:
		- Changed title to "Performance Verification and IQ, PQ, OQ Tests."
		- Added IQ/PQ/OQ test procedure information.
		- Clarified procedures for liquid tests.
		Revised decontamination instructions and added cleaning procedure (Appendix A).
		Added KC4 startup information to Appendix B.
		Added new Appendix E with two examples of assay kit instructions and directions for programming an assay.
		General:
		Edited and formatted text. Modified appearance of display screens. Standardized the presentation of significant digits. Changed "Abs" to "OD" throughout.
K	4/05	Updated the cover with a current photo of the instrument. Updated contact information and warranty.
		Changed "Automated Microplate Reader" to "Absorbance Microplate Reader" throughout. Changed "Universal Test Plate" to "Absorbance Test Plate" throughout.
		Removed references to internal barcode scanner and "R" models. Removed references to General Formulas with respect to open assays.
		Updated specifications to match product spec rev D.
		Added text from rev J1 insert (Fuse Installation/Replacement).
		Updated the installation instructions in Chapter 2 to better reflect actual practice. Updated the IQ/OQ/PQ/PM steps in Chapter 4 to better reflect actual practice.
		Created Appendix F to provide instructions for adjusting the line input voltage range and replacing fuses for instruments with an older-style power input module than the one described in chapter 2.

Revision History (Cont'd)

Rev	Date	Changes
L	4/06	Updated the manual primarily to support the introduction of Gen5™ software. In general, changed 'Bio-Tek' to 'BioTek,' and added Gen5 instructions wherever KCjunior™ and KC4™ instructions were present.

Cover: Updated BioTek logo to new graphic.

Preface: Added Contact Information section, updated Hazards, Precautions, and safety standards. Removed Registration Card and Warranty (these items ship separately).

Chapter 1: Added external power supply to Hardware Features, Package Contents, and Specifications. Also added 'Reading Speeds' section to specs. Replaced Technical Support section with one-page Product Support & Service section.

Chapter 2: Added instructions for connecting the external power supply in Connect Power section. Moved instructions in Check/Adjust the Power Input Voltage Setting section to Appendix F.

Chapter 4: Changed chapter title from 'Performance Verification, Periodic Maintenance, and IQ, PQ, OQ Tests' to 'Instrument Verification'. Moved maintenance instructions to new *Chapter 5, Preventive Maintenance* (see below). For Liquid Tests 1, 2, and 3, added recommendation to shake the plate for four minutes (or wait for 20 minutes) after pipetting the diluted test solution, *before* reading the plate.

Chapter 5 (new chapter): In Recommended Maintenance table, removed 3-month interval for cleaning the lamp contacts and changed 'Replace the Lamp' to 'Replace the Lamp and Clean the Contacts.' Added cleaning instructions from *Appendix A, Decontamination*.

Appendix B: Added new section: 'Controlling the Reader with Gen5'.

Intended Use Statement

- The ELx808™ is an 8-channel, automated, benchtop, general-purpose Enzyme Immunoassay Analyzer that performs analyses of a variety of samples. The performance characteristics of the data reduction software have not been established with any laboratory diagnostic assay. The user must evaluate this instrument and software in conjunction with the specific assay. This evaluation must include the confirmation that performance characteristics for the specific assay are met.

- The intended use of this instrument is dependent on the instrument's rear panel label. If there is an IVD label, then the instrument may be used for clinical, research and development, or other non-clinical purposes. If there is no such label, then the instrument may **only** be used for research and development or for other non-clinical purposes.

- This system is designed for use with a variety of enzyme immunoassays. Assay protocol variations are addressed by the developer of the ELISA test kit, in accordance with the test kit's procedure. A versatile curve-fitting and statistical software program is preloaded on every ELx808. If using Extensions® Define Protocol software, up to 75 assay protocols and blanking patterns can be defined, stored in memory, and instantly accessed. Plate templates and formulas are automatically combined with the protocol assay setup. Data results may be printed out, or sent to a computer running a BioTek software package, such as Gen5™, KCjunior™, or KC4™ for Microsoft® Windows®.

The ELx808's onboard software provides:

  - An easy-to-use, menu-driven interface

  - Endpoint curvilinear regressional and statistical calculations

  - Curve Fitting, with 4-parameter, cubic, quadratic, linear, 2-p, cubic spline, or point-to-point

  - Formula calculations for more complex mathematical operations

  - Ability to define controls and positive and negative cutoffs

Specimen Preparation

Samples should be obtained, treated, and stored following instructions and recommendations contained in the package kit.

Repackaging and Shipping

> If you need to ship the reader to BioTek for service or repair, contact BioTek for a **Return Materials Authorization (RMA)** number, and be sure to use the original packing. Other forms of commercially available packing are not recommended and can **void the warranty**. If the original packing materials have been damaged or lost, contact BioTek to ask for replacement packing.

Warnings

Operate the instrument on a flat surface away from excessive humidity.

Bright sunlight or strong incandescent light can reduce the linear performance range of the instrument.

Measurement values may be affected by extraneous particles (such as dust) in the microplate wells. A clean work area is necessary to ensure accurate readings.

When operated in a safe environment according to the instructions in this document, there are no known hazards associated with the instrument. However, the operator should be aware of certain situations that could result in serious injury; these may vary depending on the instrument model. See *Hazards and Precautions*.

Hazards and Precautions

Hazards

Warning! Power Rating. The instrument's power supply or power cord must be connected to a power receptacle that provides voltage and current within the specified rating for the system. Use of an incompatible power receptacle may produce electrical shock and fire hazards.

Warning! Electrical Grounding. Never use a two-prong plug adapter to connect primary power to an instrument or to a power supply. Use of a two-prong adapter disconnects the utility ground, creating a severe shock hazard. Always connect the power cord or power supply directly to a three-prong receptacle with a functional ground.

Warning! Internal Voltage. Always turn off the power switch and unplug the power cord or power supply before cleaning the outer surface of the instrument.

Warning! Liquids. Avoid spilling liquids on the instrument; fluid seepage into internal components creates a potential for shock hazard. Wipe up all spills immediately. Do not operate the instrument if internal components have been exposed to fluid.

Warning! Potential Biohazards. Some assays or specimens may pose a biohazard. Adequate safety precautions should be taken as outlined in the assay's package insert. Always wear safety glasses and appropriate protective equipment, such as chemically resistant rubber gloves and apron.

Warning! Hot Surface. The lamp is hot when the instrument is turned on. Turn off the reader and allow the lamp to cool down before attempting to replace it.

Warning! Unspecified Use. Failure to operate this equipment according to the guidelines and safeguards specified in this manual could result in a hazardous condition.

Hazards (Cont'd)

Warning! Software Quality Control. The operator must follow the manufacturer's assay package insert when modifying software parameters and establishing reading methods, using the instrument's onboard software. **Failure to conduct quality control checks could result in erroneous test data.**

Warning! Reader Data Reduction Protocol. For readers with onboard assay software, the software will flag properly defined controls when they are out of range. The software will present the data with the appropriate error flags for the operator to determine control and assay validity. For readers operated via computer control, no limits are applied to the raw absorbance data. All information exported via computer control must be thoroughly analyzed by the operator.

Precautions

The following precautions are provided to help avoid damage to the instrument:

Caution: Service. Only BioTek authorized service personnel should service the instrument. Only qualified technical personnel should perform troubleshooting and service procedures on internal components.

Caution: Environmental Conditions. Do not expose the system to temperature extremes. For proper operation, ambient temperatures should remain between 18° to 40°C. Performance may be adversely affected if temperatures fluctuate above or below this range.

Caution: Sodium Hypochlorite. Do not expose any part of the instrument to the recommended diluted sodium hypochlorite solution (bleach) for more than 20 minutes. Prolonged contact may damage the instrument surfaces. Be certain to rinse and thoroughly wipe all surfaces.

Caution: Power Supply. Only use the power supply shipped with the instrument. Operate this power supply within the range of line voltages listed on it.

Caution: Disposal. This instrument contains printed circuit boards and wiring with lead solder. Dispose of the instrument according to Directive 2002/96/EC, "on waste electrical and electronic equipment (WEEE)."

Caution: Warranty. Failure to follow preventive maintenance protocols may **void the warranty.**

	Based on the testing described below and information contained herein, this instrument bears the CE mark.

Directive 89/336/EEC: Electromagnetic Compatibility

Emissions - Class A

EN 50081-1:1992 and IEC 61326-1:1998

EN 55022:1995 Class A

Immunity

EN 50082-1:1992 and IEC 61326-1:1998

IEC 801-2: Electrostatic Discharge

EN 61000-4-3: Radiated EM Fields

EC 801-4: Electrical Fast Transient/Burst

IEC 1000-4-5: Surge Immunity

EN 61000-4-6: Conducted Disturbances

EN 61000-4-11: Voltage Dips, Short Interruptions and Variations

Directive 73/23/EEC: Low Voltage (Safety)

EN 61010-1: 2001 (2nd Edition)

"Safety requirement for electrical equipment for measurement, control and laboratory use. Part 1, General requirements."

Directive 2002/96/EC: Waste Electrical and Electronic Equipment

Disposal Notice

This instrument contains printed circuit boards and wiring with lead solder. Dispose of the instrument according to Directive 2002/96/EC, "on waste electrical and electronic equipment (WEEE)."

Directive 98/79/EC: *In Vitro* Diagnostics (some models)

- Product registration with competent authorities
- Traceability to the U.S. National Institute of Standards and Technology (NIST):

    ➢ Optical density measurements, and if equipped, incubator temperature readings, are traceable to NIST.

Specific data for a particular serial number is available on request from BioTek Instruments. See page viii for contact information.

Electromagnetic Interference and Susceptibility

USA FCC CLASS A

Warning: Changes or modifications to this unit not expressly approved by the manufacturer could **void the user's authority** to operate the equipment.

This equipment has been tested and found to comply with the limits for a Class A digital device, pursuant to Part 15 of the FCC Rules.

These limits are designed to provide reasonable protection against harmful interference when the equipment is operated in a commercial environment. Like all similar equipment, this equipment generates, uses, and can radiate radio frequency energy and, if not installed and used in accordance with the instruction manual, may cause harmful interference to radio communications. Operation of this equipment in a residential area is likely to cause interference, in which case the user will be required to correct the interference at his own expense.

Canadian Department of Communications Class A

This digital apparatus does not exceed Class A limits for radio emissions from digital apparatus set out in the Radio Interference Regulations of the Canadian Department of Communications.

Le present appareil numerique n'met pas du bruits radioelectriques depassant les limites applicables aux appareils numerique de la Class A prescrites dans le Reglement sur le brouillage radioelectrique edicte par le ministere des Communications du Canada.

User Safety

This device has been type tested by an independent laboratory and found to meet the requirements of the following:

- **Underwriters Laboratories Standard (UL) UL61010-1:2004**

  "Electrical Equipment for Laboratory Use; Part 1: General Requirements."

- **Canadian Standards Association CAN/CSA C22.2 No. 61010-1-04**

  "Safety Requirements for Electrical Equipment for Measurement, Control and Laboratory Use, Part 1: General Requirements"

  - ➢ **EC Directive 73/23/EEC**

- **EN 61010-1 (2001)**

  "Safety requirement for electrical equipment for measurement, control and laboratory use. Part 1 General requirements."

Safety Symbols

Some of the following symbols may appear on the instrument.

Alternating current
Courant alternatif
Wechselstrom
Corriente alterna
Corrente alternata

Direct current
Courant continu
Gleichstrom
Corriente continua
Corrente continua

Both direct and alternating current
Courant continu et courant alternatif
Gleich- und Wechselstrom
Corriente continua y corriente alterna
Corrente continua e corrente alternata

Earth ground terminal
Borne de terre
Erde (Betriebserde)
Borne de tierra
Terra (di funzionamento)

Protective conductor terminal
Borne de terre de protection
Schutzleiteranschluss
Borne de tierra de protección
Terra di protezione

On (Supply)
Marche (alimentation)
Ein (Verbindung mit dem Netz)
Conectado
Chiuso

Off (Supply)
Arrêt (alimentation)
Aus (Trennung vom Netz)
Desconectado
Aperto (sconnessione dalla rete di alimentazione)

Caution (refer to accompanying documents)
Attention (voir documents d'accompanement)
Achtung siehe Begleitpapiere
Atención (vease los documentos incluidos)
Attenzione, consultare la doc annessa

Warning, risk of electric shock
Attention, risque de choc électrique
Gefährliche elektrische Schlag
Precaución, riesgo de sacudida eléctrica
Attenzione, rischio di scossa elettrica

Warning, risk of crushing or pinching
Attention, risque d'écrasement et pincement
Warnen, Gefahr des Zerquetschens und Klemmen
Precaución, riesgo del machacamiento y sejeción
Attenzione, rischio di schiacciare ed intrappolarsi

Warning, hot surface
Attention, surface chaude
Warnen, heiße Oberfläche
Precaución, superficie caliente
Attenzione, superficie calda

Consult instructions for use
Consulter la notice d'emploi
Gebrauchsanweisung beachten
Consultar las instrucciones de uso
Consultare le istruzioni per uso

***In vitro* diagnostic medical device**
Dispositif médical de diagnostic *in vitro*
Medizinisches *In-Vitro*-Diagnostikum
Dispositivo médico de diagnóstico *in vitro*
Dispositivo medico diagnostico *in vitro*

Separate collection for electrical and electronic equipment
Les équipements électriques et électroniques font l'objet d'une
collecte sélective
Getrennte Sammlung von Elektro- und Elektronikgeräten
Recogida selectiva de aparatos eléctricos y electrónicos
Raccolta separata delle apparecchiature elettriche ed elettroniche

Chapter 1

Introduction

This chapter introduces the ELx808™ Absorbance Microplate Reader and describes its hardware and software features. Also included is contact information if technical assistance is needed.

This chapter contains the following sections:

Introducing the ELx808™ Absorbance Microplate Reader

BioTek's ELx808 is an eight-channel reader-assay system. The reader can serve as a standalone system, or can be controlled via PC-based software.

Designed to automatically perform endpoint and kinetic analysis, the reader can measure the optical density of solutions in 96-well microplates between 380 nm and 900 nm. (The UV option measures down to 340 nm.)

The reader features superior optical specifications, with an extended dynamic range of up to 4.000 absorbance units.

The instrument's onboard processor, 2-line x 24-character LCD screen, and membrane keys allow easy definition and management of assay protocols, templates, formulas, and data. Results can be output in a printed report format, or exported for use in a variety of ELISA-based data manipulation applications.

BioTek's ELx808 may be configured with all, or selected options for optimum performance:

- U-model instruments are capable of reading plates between 340 nm and 900 nm wavelengths.

- I-model instruments have a four-zone incubation chamber that controls temperature up to 50°C.

Quality Control

It is considered good laboratory practice to run laboratory samples according to instructions and specific recommendations included in the package insert for the test to be conducted. Failure to conduct Quality Control checks could result in erroneous test data.

Hardware Features

- Eight optics channels, with an additional reference channel

- A wavelength range of 380-900 nm (standard model), 340-900 nm (UV model)

- A user-accessible, 6-position filter wheel

- A 2-line x 24-character LCD display

- A membrane keypad with alphanumeric keys

- Adjustable plate shake frequency and times

- Reads 96-well microplates with 0.355″ well centers

- Internal, four-voltage range power input module that can be adjusted for 100, 120, 230, or 240 V~ @ 50-60 Hz (readers manufactured *before* December 2005)

- 24-volt external power supply compatible with 100-240 V~ ± 10% @ 50-60 Hz (readers manufactured *after* December 2005)

- One serial COM port (25-pin male connector) and one parallel port (25-pin female connector)

- 4-Zone™ incubation chamber option

Software Features

- Easy-to-use, menu-driven interface

- Endpoint, Kinetic, and Linear Well scanning calculations

- Curve fitting, with 4-parameter, cubic, quadratic, linear, 2-P, cubic-spline and point-to-point methods

- Transformation and formula calculations for more complex mathematical operations, including validation and cutoff formulas

- 55 assays are available onboard; up to 75 custom assays can be preprogrammed and exported to the reader using BioTek's *Define Protocol* software.

- Automatically stores results for the last 10 plates.

Specifications

Microplates:	Standard 96-well, flat or round-bottom plates
Display:	2-line x 24-character LCD
Light Source:	Tungsten halogen-filled bulb
Dimensions:	16.0″ D x 15.5″ W x 8.75" H (40.6 cm x 39.37 cm x 22.2 cm)
Weight:	35 lb. maximum (15.87 kg maximum)
Environment:	Operating temperature 18° to 40°C
Humidity:	10% to 85% noncondensing
Power Source:	Readers manufactured *before* December 2005:

Internal, adjustable power input module with four voltage ranges accommodated by the voltage selection switch:

Range 1	100 V~	90 to 110 V~, 50 to 60 Hz
Range 2	120 V~	108 to 132 V~, 50 to 60 Hz
Range 3	230 V~	207 to 253 V~, 50 to 60 Hz
Range 4	240 V~	216 to 264 V~, 50 to 60 Hz

Readers manufactured *after* December 2005:

24-volt external power supply compatible with 100-240 V~ ± 10% @ 50-60 Hz

Power Consumption:	100 VA
Reading Speeds:	*Single wavelength:* 8 seconds rapid mode; 12 seconds normal/regular mode
	Dual wavelength: 13 seconds rapid mode; 20 seconds normal/regular mode
	Single wavelength higher than 400 nm: 6-second minimum kinetic interval, rapid mode

Standard Model

Note: The following specifications apply only to standard 96-well, flat- or round-bottom microplates.

Wavelength Range:	380 to 900 nm
Filters:	10 nm half-bandwidth interference filters.
	User-accessible filter wheel. Up to 6 filters may be installed on the instrument at one time.
	Filters supplied: 405 nm, 450 nm, 490 nm, 630 nm, and two blank filters.
Absorbance Measurement Range:	0.000 to 4.000 OD

- Single-wavelength endpoint measurements with a **12-second read** (normal/regular read mode):

Accuracy:	± 1.0% ± 0.010 OD from 0.000 to 2.500 OD @ 405 nm
Linearity:	± 1.0% from 0.000 to 2.500 OD at 405 nm
	± 2.0% from 2.500 OD to 3.500 OD @ 405 nm
Repeatability:	± 0.5% ± 0.005 OD from 0.000 to 2.500 OD @ 405 nm
	± 1.5% ± 0.005 OD from 2.500 to 3.500 OD @ 405 nm
	± 2.5% ± 0.005 OD from 3.500 to 4.000 OD @ 405 nm

- Single-wavelength kinetic measurements with read intervals of less than **12 seconds***:

Accuracy:	± 2.0% ± 0.010 OD from 0.000 to 2.500 OD @ 405 nm
Linearity:	± 2.0% from 0.000 to 2.500 OD @ 405 nm
Repeatability:	± 1.0% ± 0.010 OD from 0.000 to 2.500 OD @ 405 nm

  *Plate can be read at **6-second** intervals (rapid mode) with wavelengths higher than 400 nm.

Ultraviolet/Incubator Model

Note: The following specifications apply only to standard 96-well, flat- or round-bottom microplates.

Wavelength Range:	340 to 900 nm
Filters:	10 nm half-bandwidth interference filters.
	User-accessible filter wheel. Up to 6 filters may be installed on the instrument at one time.
	Filters supplied: 340, 405, 450, 490, 630 nm
Absorbance Measurement Range:	0.000 to 4.000 OD for 400 to 900 nm range 0.000 to 3.000 OD for 340 to 400 nm range

Optical specifications for the 400 to 900 nm range **(12-second read** in normal/regular read mode):

Accuracy, Linearity, Repeatability:	Same as Standard model

Optical specifications for the 340 to 400 nm range **(12-second read** in normal/regular mode**):**

Accuracy: $\pm$ 1.0% $\pm$ 0.010 OD from 0.000 to 2.000 OD @ 340 nm

Linearity: $\pm$ 1.0% from 0.000 to 2.000 OD @ 340 nm

Repeatability: $\pm$ 1.0% $\pm$ 0.005 OD from 0.000 to 2.000 OD @ 340 nm

Incubation The following specifications apply to a 96-well sealed plate with 200 µl of liquid in all wells:

- **Temperature Control:**   Temperature controlled to 50°C
- **Temperature Variation:** $\pm$ 0.50°C @ 37°C (with the plate sealed)

Package Contents

The contents of the ELx808™ package include:

- Absorbance Microplate Reader

- Power cord (PN varies according to country of use)

- Power supply for readers manufactured after December 2005 (PN 61062, ELx808; PN 76053, ELx808IU)

- Filter wheel with four standard filters: 405 nm, 450 nm, 490 nm, 630 nm and two blank filters. UV models also include a 340-nm filter.

- Operator's Manual (PN 7341000)

- Dust cover (PN 7342066)

- Parallel cable (PN 71072)

- Serial cable (PN 75053)

- Packing instructions (PN 7341022)

- Shipping document that includes Warranty Statement and Certificate of Compliance and Calibration (PN 94075)

- Declaration of Conformity (PN 7341005, clinical models, or PN 7341031, non-clinical models)

Optional Accessories

- Additional filters (please inquire for availability):

  - ➢ 340 to 630 nm, PN 3100XXX (XXX is wavelength in nanometers)

  - ➢ 640 to 900 nm, PN 3404XXX (XXX is wavelength in nanometers)

- Replacement lamp assembly (PN 3400508)

- Filter wheel plug (PN 3122037)

- Absorbance Test Plate (PN 9000547 or PN 7260522)

- ELx808™ Service Manual for all models (PN 7341004)

- ELx808 Qualification Package (PN 7340543)

- Gen5™, KC4™, or KCjunior™ Software (PNs and versions listed on biotek.com or contact your local dealer)

- BioTek QC Check Solution No. 1 (PN 7120779, 25 ml; PN 7120782, 125 ml)

- BioTek wetting agent (PN 7773002)

Product Support & Service

A superior support staff backs all of BioTek's products. If your instrument(s) or software ever fails to function perfectly, if you have questions about how to use or maintain it, or if you need to send an instrument to BioTek for service or repair, please contact our Technical Assistance Center (TAC).

Contacting the Technical Assistance Center

Our Technical Assistance Center is open from 8:30 AM to 5:30 PM (EST), Monday through Friday, excluding standard U.S. holidays. You can send a fax or an e-mail any time.

Phone: 800-242-4685 (in the U.S.) or 802-655-4740 (outside the U.S.)

Fax: 802-655-3399

E-Mail: tac@biotek.com

Please be prepared to provide the following information:

- Your name and company information

- A daytime phone or fax number, and/or an e-mail address

- The product name, model, and serial number

- The software part number and basecode version

- For troubleshooting assistance or instruments needing repair, the specific steps that produce your problem and any error codes displayed (see also *Appendix C*, *Error Codes*).

Returning Instruments for Service/Repair

If you need to return an instrument to BioTek for service or repair, please contact the TAC for a Return Materials Authorization (RMA) number *before* shipping the instrument. Repackage the instrument properly (see *Chapter 2*, *Installation*), write the RMA number on the shipping box, and ship to this address:

BioTek Instruments, Inc.
Technical Assistance Center
100 Tigan Street
Highland Park
Winooski, Vermont 05404 USA

Installation

This chapter includes instructions for unpacking and setting up the ELx808™, and instructions for connecting printers and/or serial devices.

This chapter contains the following sections:

Unpack and Inspect the Instrument

- If the shipping box has been damaged, inspect the instrument for visible dents and scratches as you unpack it.

- If the reader is damaged, notify the carrier and your BioTek sales representative. Keep the shipping cartons and the packing material for the carrier's inspection. The manufacturer will arrange for the repair or replacement of your instrument immediately.

> **Important!** Save all packaging materials. If you need to ship the reader to BioTek for repair or replacement, you must use the original packing. Other forms of commercially available packing are not recommended and can **void the warranty**. If the original packaging materials have been damaged or lost, contact BioTek to order PN 7343000. See *Product Support & Service* in *Chapter 1* for contact information.

- Refer to the figure on the next page when unpacking the reader.

- Carefully open the top of the box, and remove any accessories. These include a power cord, a filter wheel in a padded envelope, and an Operator's Manual.

- Remove the end caps from the reader.

- Lift the reader out of the box, and place it on a level surface. Remove the reader from the plastic bag.

- Remove the filter wheel from the shipping envelope

- Place all packing material back into the shipping box for reuse if the instrument needs to be shipped again.
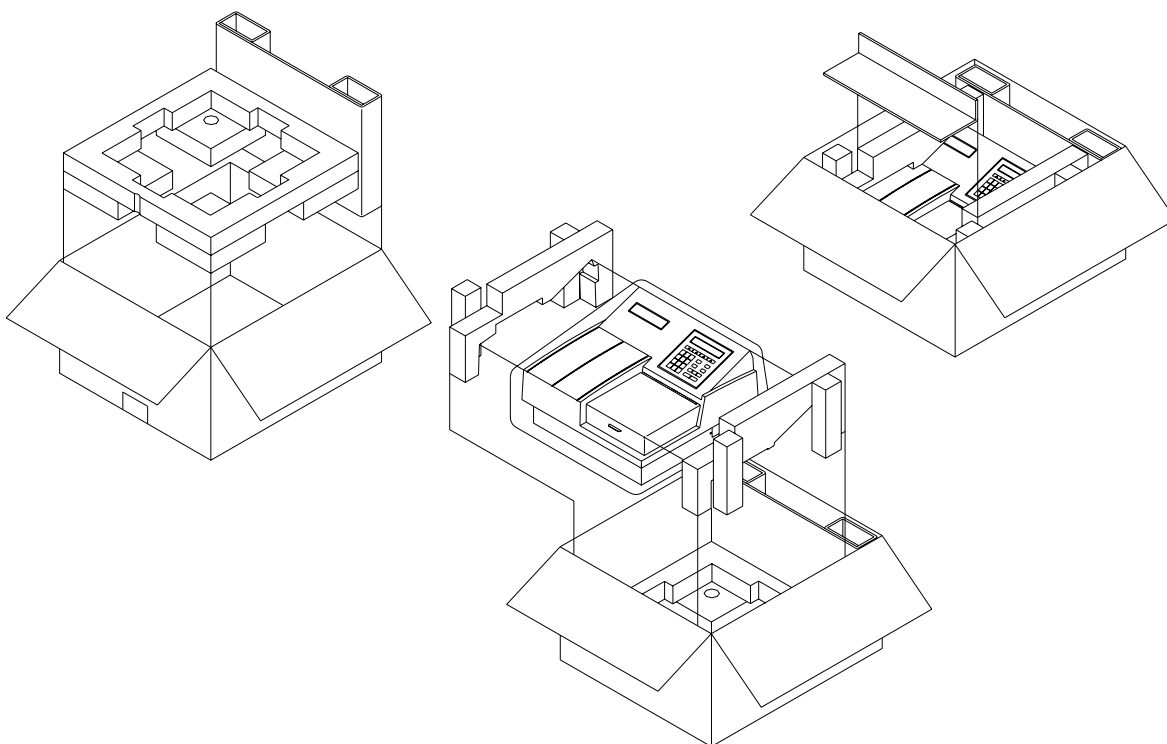
Figure 2-1: *Unpacking the ELx808™*

Select the Operating Environment

For best operation, install BioTek's ELx808 on a level surface in an area where ambient temperatures between 18°C (64°F) and 40°C (104°F) can be maintained.

The reader is sensitive to extreme environmental conditions. Conditions to avoid are:

- **Excessive humidity:** Condensation directly on the sensitive electronic circuits can cause the instrument to fail internal self-checks. The humidity must be in the range of 10% to 85%, noncondensing.

- **Excessive ambient light**: Bright sunlight or strong incandescent light can reduce the linear performance range and affect the instrument's readings.

- **Dust:** Optical density readings may be affected by extraneous particles (such as dust) in the microplate wells. A clean work area is necessary to ensure accurate readings.

Install the Filter Wheel

The filters that are shipped with the ELx808™ are installed in the six-position filter wheel. For example, the standard models have 405, 450, 490, 630 nm filters; the UV model's filter set is 405, 450, 490, 630 and 340 nm.

The filter wheel, which is packaged in a shipping envelope, must be installed before the reader is used. If you plan to install additional filters, or change the filter locations, use the following instructions to gain access to the filter wheel.

Plate access door

Filter wheel, shown installed
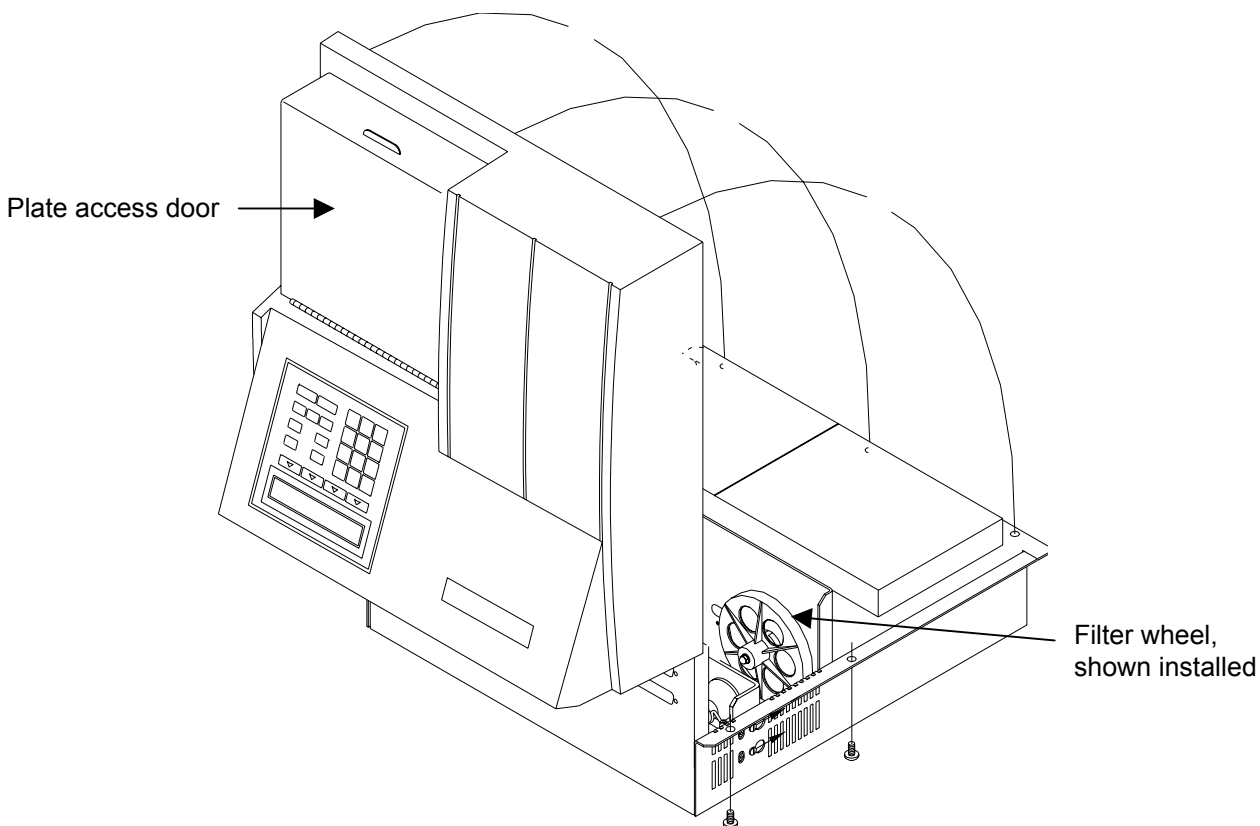
Figure 2-2: *Installing the Filter Wheel*

- If the reader is on, turn it off and disconnect the power cord or power supply.

- Remove the seven screws around the perimeter of the shroud with a flat-blade screwdriver, as shown above. *Tip*: Bring the reader to the edge of the work surface to access the screws without having to turn the reader upside down.

- Carefully lift up the shroud from the front. (Note that the shroud is hinged along its back edge.) Hold the plate access door steady, or tape it closed as the shroud is being lifted to prevent the door from moving.

- Ensure that all locations on the filter wheel contain either a filter or a blank. Each location on the filter wheel must be occupied for the reader to operate properly. Take a moment to record the filter values in each location (e.g., 405 nm in position 1).

Important! Keep track of all filter locations. The physical location of the filters must match the filter locations mapped in the reader's software filter table. The filter wheel must have no empty locations; all locations must be filled with either a filter or a blank plug. Install all filters with the arrow denoting the light direction pointing downward.

- The filter wheel mount is located in the rear left corner inside the reader. The filter wheel is held in place by a magnet on the filter wheel motor hub.

  To install the filter wheel:

  1. Line up the registration notch on the hub and the corresponding peg on the filter wheel.

  2. Apply firm pressure to attach the filter wheel to the hub. The peg must be engaged in the notch for proper installation.

  3. Ensure that the filter wheel is positioned flat against the hub, and that it rotates freely.

  4. Lower the top shroud back into position and remove tape if used to steady the plate access door.

  5. Reinstall the seven screws removed from the perimeter of the shroud.

Important! Be sure to replace the seven **perimeter screws**; they increase the reader's ability to withstand electrostatic discharges and electromagnetic interference. In addition, the screws MUST be installed to hold the shroud in place if you plan to ship the instrument

---

- To remove the filter wheel, grasp the center hub of the filter wheel and pull it toward the lamp. The wheel should easily disconnect from the hub. Lift the wheel from the instrument.

- Store unused filters in a cool, dry place away from direct sunlight. The filters can be wrapped in a piece of lens paper to protect them from scratches and dust accumulation.

Check/Adjust the Power Input Voltage Setting

ELx808™ readers manufactured before December 2005 are powered by an internal, adjustable four-voltage range power input module, instead of by the external power supply. For instructions for checking or adjusting the power input voltage setting, and for reconfiguring or replacing the fuses, refer to *Appendix F*, *Adjusting the Power Input Voltage Setting*.

Connect Power

ELx808 with the internal power input module:

1. Locate the power inlet on the right side of the reader.

2. Plug the rounded end of the power cord into the power inlet.

3. Plug the 3-prong end of the cord into an appropriate power receptacle.

ELx808 with the external power supply:

1. Connect the power cord to the external power supply.

2. Locate the power inlet on the right side of the reader.

3. Plug the rounded end of the power supply's line cord into the power inlet.

4. Plug the 3-prong end of the power cord into an appropriate power receptacle.

Warning! Power Rating. The ELx808 or power supply must be connected to a power receptacle that provides voltage and current within the specified rating for the system. Use of an incompatible power receptacle may produce electrical shock and fire hazards.

Warning! Electrical Grounding. Never use a two-prong plug adapter to connect primary power to the ELx808 or to the power supply. Use of a two-prong adapter disconnects the utility ground, creating a severe shock hazard. Always connect the ELx808 or power supply directly to an appropriate receptacle with a functional ground.

Install the Printer

If the ELx808™ will be operated in standalone mode (that is, without BioTek's Gen5™, KCjunior™, or KC4™ software running on a host PC), connect a printer directly to the reader using the supplied parallel cable.

To attach a printer to the ELx808:

(i) **Important!** To avoid system instability, make sure the printer and reader are turned OFF before connecting them.

- If the reader and/or printer are on, turn them off. Place the printer in a location adjacent to the reader.

- Attach one end of the parallel cable to the printer's parallel port.

- Attach the other end of the cable to the reader's parallel port, located on the instrument's rear panel.

- Make sure the securing screws on both ends of the cable are tightened.

- Turn on the printer.
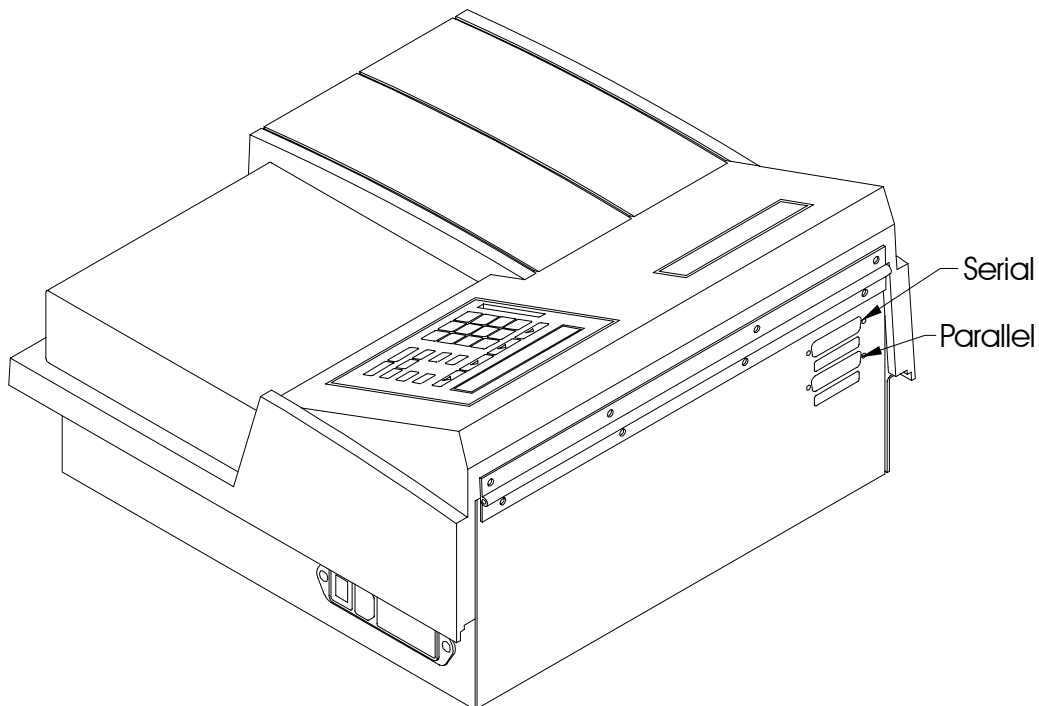
Figure 2-3: Serial and Parallel ports

Printers

The ELx808's parallel port (LPT1) allows connection to compatible printers. The reader supports printers using either HP's PCL3 language, such as the HP DeskJet series, or Epson's LQ language. For the latest list of compatible printers call BioTek's Technical Assistance Center or visit our website (refer to *Chapter 1* for contact information).

Cable

A printer cable is supplied with the reader. If this cable becomes lost or damaged, BioTek offers a variety of printer cables. Contact your authorized BioTek dealer for information on cable prices and availability.

Parallel Port Pin Definition

The table that follows illustrates the pin definitions for the reader's 25-pin (socket-female) D-sub Parallel connector.

Table 2-1
Parallel Connector Pinouts

Parallel Port Pinout			
Pin	**Signal**	**Pin**	**Signal**
1	PSTROBE	14	NC
2	D0	15	NC
3	D1	16	RESET
4	D2	17	NC
5	D3	18	GND
6	D4	19	GND
7	D5	20	GND
8	D6	21	GND
9	D7	22	GND
10	NC	23	GND
11	BUSY	24	GND
12	NC	25	GND
13	NC		

Turn on the Reader and Run a System Test

If all of the required steps preceding this one have been performed successfully, locate the reader's power switch (on the right side) and turn on the reader. The ELx808™ will automatically perform a **System Test**.

The System Test conducts a series of tests at each wavelength defined in the filter table to confirm adequate light levels, low electronic noise, adequate photodiode sensitivity, overall system cleanliness, and (if equipped) proper function of the incubator. The testing is designed to verify that the reader will give in-specification performance for each set wavelength over the specified OD range. If a printer is connected to the reader, the test results will automatically print.

Note: This test can also be run through Gen5™, KCjunior™, or KC4™ running on a host PC. Consult Gen5's Help system, or the KC4 or KCjunior User's Guide for instructions.

- **If the test passes**, the reader will beep once and the display will show the software's main menu, which will resemble the following:

```
READY          1:30PM       23.4°C
READ    DEFINE    REPORT    UTIL
```

- **If the test fails**, the reader will beep repeatedly and the display will show an error code. If this happens, write down the error code and then press the **Stop** key on the keypad to stop the beeping. Look up the error code in *Appendix C, Error Codes* to determine its cause. If the problem is something you can fix (for example, if the error code is 0201 or 0301, indicating that the filter wheel is missing), turn off the reader, fix the problem, and then turn the reader back on. If the cause is not something you can fix, contact BioTek's Technical Assistance Center.

Check/Adjust the Reader's Filter Table

After installing the filter wheel (or new filters), ensure that the ELx808's filter table accurately maps the physical location of the filters in the filter wheel. To check and/or adjust the filter table:

- Turn on the reader if it is not already on. At the Main Menu screen (shown below), press the soft key under the **UTIL** menu option.

```
READY        01:30PM  10/09/04

READ    DEFINE   REPORT   UTIL
```

- The Select Utility Option menu appears:

```
SELECT  UTILITY  OPTION:

TESTS  SETUP   OUTPUT   READ
```

- From the Select Utility Option menu, press the soft key beneath **SETUP**. The Edit Setup Information screen appears:

```
EDIT  SETUP  INFORMATION:

 DATE    TIME   FILTER   *MORE
```

- From this menu, press the soft key beneath **FILTER**. The wavelength for Filter #1 appears:

```
ENTER

FILTER#1  WAVELENGTH:405
```

- If you need to change the filter wavelength number, use the **NUMERIC** keypad to enter a number at the cursor location. To save the entry and move to the next filter on the filter table, press the **ENTER** key.

- When the last filter has been entered, the software exits the filter routine, and displays the following screen:

```
EDIT  SETUP  INFORMATION:

 DATE    TIME   FILTER   *MORE
```

- Press the **MAIN MENU** key to return to the Main Menu.

Configure Utility Options

The ELx808™ may be configured a number of ways, depending on user preference. Configuration options are accessed via the **Utility Options** menu, and include:

- **SETUP:** Set the date and time.

- **OUTPUT:** Select whether reports will be output to a Printer, the Computer, or both, choose the Column or Matrix report format, and determine if Curves will be printed.

- **READ:** Indicate whether prompts shall be displayed at run-time for Plate IDs, Sample IDs, and Sample Counts.

- **TESTS:** Access functions to test reader optics and instrument calibration. Check the version of basecode software installed. (See *Chapter 4* for more information.)

To select the SETUP, OUTPUT, and READ user-configurable options:

- At the Main Menu screen, press the soft key beneath **UTIL** to access the Utility Options menu.

```
SELECT   UTILITY   OPTION:

TESTS   SETUP   OUTPUT   READ
```

SETUP Options

- At the **UTILITY OPTIONS** screen, press the soft key beneath **SETUP**.

```
EDIT   SETUP   INFORMATION

 DATE     TIME   FILTER   *MORE
```

- At the **EDIT SETUP INFORMATION** screen, press the soft key beneath **DATE**.

```
DATE:    10/09/04          MDY

MMDDYY          DDMMYY
```

- Enter the new date, using the NUMERIC keys. The cursor is positioned under the first editable field, and advances automatically. To change the date format, press the soft key beneath **MMDDYY** or **DDMMYY**. The display updates to reflect the new format.

- Press **ENTER** to return to the **EDIT SETUP INFORMATION** screen.

```
T I M E :          0 3 : 1 1 P M          1 2 H R

1 2 H O U R          2 4 H O U R     A M / P M
```

- To edit the **TIME**, press the soft key positioned beneath **TIME**. At the **TIME** entry screen, use the NUMERIC keys to enter the correct time.

- Select a 12- or 24-hour format by pressing the soft key beneath these options. The display automatically updates with the new time / format.

- Press the **Previous Screen** key to return to the **SELECT UTILITY OPTION** menu.

OUTPUT Options

- At the **SELECT UTILITY OPTION** screen, press the soft key beneath **OUTPUT** to set report output preferences.

```
R E P O R T    O U T P U T ?    B O T H

P R I N T      C O M P U T E R     B O T H
```

- Any previously defined selection appears on the top line of the display. Press the soft key beneath the desired option (**PRINT**, **COMPUTER**, **BOTH**) to change the output device.

- Press **ENTER** to advance to the **SELECT PRINTER** menu screen. If **COMPUTER** is pressed, all results will be transferred directly to the computer screen via the RS232 serial port.

```
S E L E C T    P R I N T E R          E P S O N

E P S O N       H P
```

Compatible Printers

BTI readers support printers using either HP's PCL3 language, such as the HP DeskJet series, or Epson's LQ language.

For the latest list of compatible printers call BioTek's Technical Assistance Center (see *Chapter 1* for contact information), or visit our Web site: www.biotek.com.

REPORT Type

ⓘ   **Note:** See *Appendix D* for examples of Reports.

- At the **SELECT PRINTER** screen, press **ENTER** to advance to the **REPORT TYPE** menu screen.

```
R E P O R T    T Y P E :    M A T R I X

C O L U M N              M A T R I X      B O T H
```

- The current selection appears on the top line of the display. Press the soft key beneath the desired option (**COLUMN, MATRIX, BOTH**) to change the Report Type. The display will automatically update with the new type.

- Press **ENTER** to advance to the **SAMPLES ON COLUMN REPORT** screen.

```
S A M P L E S    O N    C O L    R P T ?      N O

  Y E S          N O
```

- Select **YES** to print samples on the column report, or **NO** to omit them

- Press **ENTER** to advance to the **PRINT CURVE FIT** screen.

```
P R I N T    C U R V E – F I T ?          N O

  Y E S          N O
```

- The current selection is displayed on the top line of the screen. To change the report option, press the soft key under **YES** or **NO**. The display updates to reflect the selection.

- Choose **YES** if there are quantitative assays defined on board and you wish to print the curve.

- Press **ENTER** to return to the **SELECT UTILITY OPTION** screen.

READ Options

- At the **SELECT UTILITY OPTION** screen, press the soft key beneath **READ** to set up Reader Prompt preferences. Select **YES** to prompt the operator to enter identifications and sample counts before a microplate is read. The prompts are shown below.

- Press **ENTER** after each selection to advance the display.

```
P R O M P T     F O R     P L A T E     I D ?     N O

  Y E S           N O
```

- **Prompt for Plate ID** allows the user to enter an alphanumeric name of up to 10 characters. Select **YES** if to present this prompt at run-time.

```
P R O M P T     F O R     S A M P L E     I D ?     N O

  Y E S           N O
```

- **Prompt for Sample ID** allows the user to identify samples with a 4-character alphanumeric name. The starting ID is entered and automatically incremented by the software. Select **YES** to present this prompt at run-time.

```
P R O M P T     S A M P L E     C O U N T ?       N O

  Y E S           N O
```

- **PROMPT SAMPLE COUNT** allows the user to enter the total number of samples on the plate at runtime; only those sample results will be printed. Select **YES** to present this prompt at run-time.

Read Speed

```
R E A D     I N     R A P I D     M O D E ?         N O

  Y E S           N O
```

- **RAPID MODE** reads a 96-well plate at 6-second intervals with wavelengths higher than 400 nm. Selecting **NO** means a plate with single-wavelength kinetic measurements will be read in intervals less than 12 seconds ("Regular" or "Normal" Mode). Specifications for these modes are outlined in Chapter 1.

- When selections are completed, the display returns to the **SELECT UTILITY OPTION** screen.

(Optional) Connect the Host PC

The ELx808™ has a 25-pin serial (RS232) port located on the rear panel of the instrument (see *Figure 2-3* on page 2-7). The serial port allows the reader to communicate with a computer, using BioTek's Gen5™, KC4™, or KCjunior™ software, standard communications software, and/or RS-232 protocols. *Appendix B* contains information on required protocols for computer control of the reader.

In addition, Assay Definition Files created using BioTek's Microsoft® Windows®-based Extensions® Define Reader Protocol software can be downloaded to the reader using the instrument's RS-232/serial interface. For more information about downloading custom assay files, contact BioTek.

Attach the Serial Cable

If the ELx808 will be controlled by software running on a host PC, attach the serial cable:

- Power down the computer and the reader.

- Connect the appropriate serial cable to both machines.

- Power up the reader and the computer.

- Ensure that the ELx808 and the computer are operating with the same communications settings (see below).

Install Software on the Host PC

Refer to Gen5's Getting Started Guide or Help system, or to the KC4 or KCjunior User's Guides for complete instructions for installation of the software.

Set Communications Parameters

Before serial communications are initiated between the ELx808 and another device, the communication parameters (Baud Rate, Data Bit configuration, and Parity status) must match on both devices. The reader's default communication parameters are:

- 9600 Baud

- 8 Data Bits

- 2 Stop Bit

- No Parity

The user may change the Baud rate on the reader from the recommended default setting of 9600 to 1200 or 2400 (see instructions below). Data bits, Stop Bits and Parity are not user-configurable. The communications software operating on the host computer device should be set to 8, 2, and None.

Refer to Gen5's Help system or to the KC4™ or KCjunior™ User's Guide for complete instructions for setting/testing communication parameters with the ELx808.

Change the Baud Rate on the ELx808™

To change the baud rate from the default of 9600 to either 1200 or 2400:

- Turn on the instrument if it is not already on.

```
R E A D Y        0 1 : 3 0 P M   1 0 / 0 9 / 0 4
R E A D     D E F I N E    R E P O R T    U T I L
```

- At the Main Menu screen, press the soft key beneath **UTIL**.

```
S E L E C T    U T I L I T Y    O P T I O N :
T E S T S    S E T U P    O U T P U T    R E A D
```

- At the **UTILITY OPTIONS** screen, press the soft key beneath **SETUP**.

```
E D I T    S E T U P    I N F O R M A T I O N :
  D A T E      T I M E    F I L T E R    * M O R E
```

- At the **EDIT SETUP INFORMATION** screen, press the soft key beneath ***MORE** to advance to the **EDIT SETUP / RS232** option screen.

```
E D I T    S E T U P    I N F O R M A T I O N :
R S 2 3 2        C A L P L A T E        * M O R E
```

- Press the soft key beneath **RS232** to access the **BAUD RATE** selection menu. The top line of the display shows the baud rate currently set.

```
S E L E C T    B A U D    R A T E :    9 6 0 0
  1 2 0 0      2 4 0 0      9 6 0 0      V I E W
```

- To change the **BAUD** rate, press the soft key beneath the desired baud rate. The display (top line) automatically updates to reflect the new choice.

- To view the reader's settings for parity, stop bits, and data bits, press the soft key beneath **VIEW**.

Serial Port Pinout Description

The serial port on the ELx808™ is a DTE configuration with a 25-pin (pin-male) D-sub connector. If the supplied serial cable becomes lost or damaged, contact BioTek for information on replacement cables.

Table 2-2
Serial Pinout Description

Serial Pin Description			
Pin	**Signal**	**Pin**	**Signal**
1	NC	14	NC
2	TX	15	NC
3	RX	16	NC
4	RTS	17	NC
5	CTS	18	NC
6	DSR	19	NC
7	GND	20	DTR
8	DCD	21	NC
9	NC	22	RI
10	NC	23	NC
11	NC	24	NC
12	NC	25	NC
13	NC		

After Installation and Setup, Verify Performance

Before using the ELx808™ for the first time, verify that the reader is operating properly by running a System Test and the Absorbance Plate Test. These tests and additional verification procedures are described in *Chapter 4*.

Before Repackaging the Instrument

- Decontaminate the reader prior before shipping. Refer to the Decontamination procedure in *Appendix A*.

- Once the reader is clean, pack it in its original shipping box, using original packing materials (see *Figure 2-1* on page 2-3).

- Contact BioTek's Technical Assistance Center for a **Return Materials Authorization (RMA) number** and shipping instructions.

- This shipping system was designed to be used no more than five times. If the container is damaged and/or has been used more than five times, contact the Technical Assistance Center for a new set of shipping materials.

This chapter includes instructions for operating the ELx808™ and its on-board software, and contains the following sections:

Introduction

The ELx808™ features a 25-pad keypad and a 2-line x 24-character LCD display as the user interface. The reader's bidirectional serial port allows computer control of the instrument, and provides the means for downloading additional assay protocols to the instrument.

The Keypad
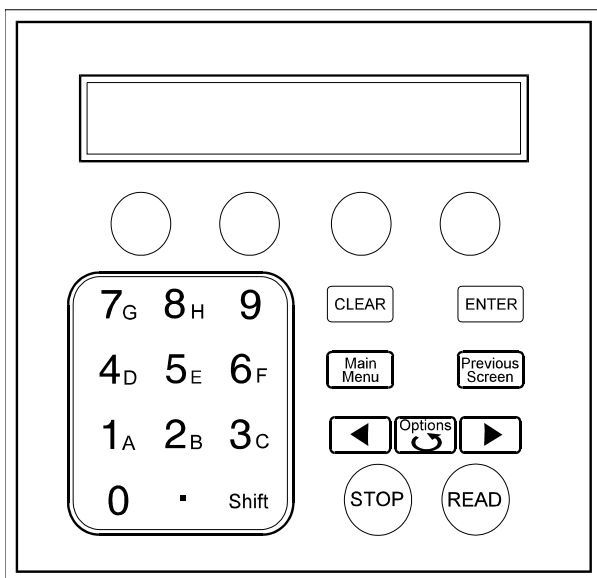
Figure 3-1: Keypad

- Use the four **"soft"** keys, located directly below the display, to select options presented on the display.

- Use the **labeled** keys to enter information, move around the reader's menu structure, specify the assay number, and so on.

- **CLEAR**: Clear/reset the current value on the reader's LCD display.

- **ENTER**: Advance the display to the next screen.

- **Main Menu**: Return the display to the Main Menu screen.

- **Previous Screen**: Return the display to the previous screen.

- **Options**: Cycle through the available options presented on the display.

- **STOP**: Halt a read in progress.

- **READ**: Select an assay and read a plate.

The Startup Screen

The ELx808™ performs a System Test when powered on, displaying the Startup screen until initialization is complete. During this period, the keyboard is inactive.

If System Test fails, the reader will beep and display an error code. Refer to *Appendix C, Error Codes* to interpret this code. For further information, please call the Technical Assistance Center (see *Chapter 1* for contact information).

```
P O W E R U P    S E Q U E N C E    V X . X X

I N I T I A L I Z I N G . . .
```

```
B i o - T e k    M o d e l

S y s t e m    S e l f - T e s t . . .
```

The Main Menu Screen

Once the system is initialized, the **Main Menu Screen** is displayed. This screen will vary slightly if the instrument has the Incubation option. The keyboard's four "soft keys," located below the on-screen menu options (**READ, DEFINE, REPORT** and **UTIL**), are activated, and may be selected.

```
R E A D Y       0 1 : 3 0 P M    1 0 / 0 9 / 0 4

R E A D    D E F I N E    R E P O R T    U T I L
```

Figure 3-2: Main Menu screen

```
R E A D Y       0 1 : 3 0 P M      3 7 . 0 ° C

R E A D    D E F I N E    R E P O R T    U T I L
```

Figure 3-3: Main Menu screen in "I " (Incubator) instruments

Note: The temperature indicated on the display of incubated models is the actual averaged temperature of the incubator's four zones. The applied setpoint of the last assay is used. To adjust the temperature, a new setpoint value must be assigned to the assay before running.

Press the "soft key" that corresponds to a displayed menu option to activate that option:

- **READ** option: Initiate a plate read. (Or, press the key labeled **READ** on the keyboard for plate reading prompts.) You will be prompted to select from a list of available assays.

- **DEFINE** option: Create a reading and data reduction protocol. You will be prompted to select/edit an assay and then define its various parameters.

- **REPORT** option: Print results reports and protocol descriptions. For results reports, you will be prompted to select a previously run assay with valid data.

- **UTIL** option: Access various onboard utilities, used for configuring and testing the reader.

(i) **Important!** On some readers, **Assay 01** has been designed to allow for quick and simple assay programming. It appears as "_Quick Read" on the display. Most of the options available in Assays 2-55, and described in this section, are unavailable for programming within Quick Read. You can access the Quick Read assay when **READ** is selected from the main menu.

The **Quick Read** assay **DEFINE** settings are shown below. They cannot be edited, except where noted:

METHOD

Single Wavelength 405 nm (editable)

MAP

96-well plate geometry

Blank on Air

Automap

Map starting location A1

Samples only (no blanks, standard, or controls)

Sample count prompted at runtime (can be turned off in UTIL | READ options)

Note: *Appendix E* provides two examples of assay kit instructions and step-by-step directions for programming each assay. The appendix includes two sample assays: one with a ratio transformation calculation and a POS/NEG cutoff determination, and another with a standard curve.

Defining Assays

The Main Menu option, **DEFINE,** allows you to customize previously defined assays stored in the reader's memory.

- From the Main Menu, press the soft key beneath the **DEFINE** menu option to advance to the **SELECT ASSAY NUMBER** screen.

Selecting an Assay

At the **SELECT ASSAY NUMBER** screen:

- Use the **NUMERIC** keys to enter the number of predefined Assay Definition files stored in the reader's memory, or use the **OPTION** key to advance one assay at a time. The cursor is positioned at the first editable field, and advances automatically. The numeric range depends on the number of assays (1-55 or more if custom programmed) programmed in the reader's memory.

- The assay's name and number are displayed on the screen.

```
SELECT   ASSAY   NUMBER: 65

NAME: HBS-AG1
```

- Press **ENTER** to select the assay and advance to the **NAME** screen. You may change the default assay name to a more descriptive one (see *Assay Name* on the next page).

- Press **ENTER** to advance to the **EDIT ASSAY NAME** screen.

Assay Name

At the **ASSAY NAME** screen, edit the name assigned to the Assay. The assay name can be up to 16 characters.

```
E D I T > H B S - A G 1

 –               /               :           S P A C E
```

- Press the **ALPHA** and **NUMERIC** keys to update the assay name. The cursor is positioned at the first editable field.

- Press the **OPTION** key to sequentially advance the character positioned above the cursor. The characters will cycle through the alphabet (A-Z), with a space character following Z.

- Press **CLEAR** to clear the assay name from the display.

- Use the **LEFT** and **RIGHT ARROWS** to move the cursor to the previous or next editable field. The cursor will wrap around the edit field.

- Press **SOFT KEYS 1, 2, 3,** and **4** when using alphanumeric characters on the display above the soft key in the assay name.

In addition:

- Press the **MAIN MENU** key to return to the Main Menu screen.

- Press **PREVIOUS SCREEN** to save the contents of the display and return to the previous screen.

- Press **ENTER** to save the contents of the display and advance to the next screen.

Defining the Method, Map, Formula, and Curve

The **DEFINE** Option screen allows you to edit the Method, Map, Formula or Curve Fit.

```
D E F I N E

M E T H O D     M A P     F O R M U L A     C U R V E
```

Press the soft key beneath the displayed option to access the following functions:

- **SOFT KEY 1: METHOD** is selected, and the user is prompted to select the read method parameters.

- **SOFT KEY 2: MAP** is selected and the user is prompted for plate mapping information.

- **SOFT KEY 3: FORMULA** is selected and the user is prompted to enter a formula.

- **SOFT KEY 4: CURVE FIT** is selected and the user is prompted for curve-fit options.

- In addition, the **MAIN MENU, PREVIOUS SCREEN** and **ENTER** keys are active, allowing you to move back and advance through the menu structure.

METHOD

The definition of a method includes selecting:

- Endpoint, Kinetic or Scanning Read Modes

- Delay First Read

- Incubation Parameters

- Filter Wavelengths Applied

- Shake Parameters

- Kinetic Analysis

Note: Some screens shown below and on the following pages may not appear on some reader models.

READ TYPE

This option allows you to enter which read type: Endpoint, Kinetic, or Scan. The following keys are active during this screen:

```
READ   TYPE:KINETIC

ENDPOINT      KINETIC   SCAN
```

- Press **SOFT KEY 1** to select Endpoint read mode. Press **SOFT KEY 2** to select Kinetic read mode.

- Press **ENTER** to save the displayed value and advance to the next screen.

DELAY IN FIRST READ

Selecting the Delay in First Read option allows you to enter a time delay before the first read.

```
DELAY   FIRST   READ

TIME:    XX:XX
```

- Enter the time in minutes and seconds, using the **numeric** keys.

- Press **ENTER** to advance to the INCUBATION TEMPERATURE screen.

INCUBATION TEMPERATURE

The incubation temperature screen allows you to set the assay incubation temperature.

```
I N C U B A T I O N   T E M P :   3 7 C

A M B I E N T         T E M P E R A T U R E
```

- Press **SOFT KEY 1** or **2** to select ambient incubation.

- Press **SOFT KEY 3** or **4** to select an adjustable temperature.

- Use the **LEFT** and **RIGHT ARROW** keys to move the cursor between the two digits on the input temperature.

- Use **NUMERIC** keys to enter the incubation temperature, up to 50°C.

- Press **ENTER** to save and advance to the next screen.

SINGLE OR DUAL WAVELENGTH

The **WAVELENGTH** selection screen allows you to select **SINGLE** or **DUAL** wavelength for the assay.

```
W A V E L E N G T H :   D U A L

S I N G L E   D U A L
```

- Press **SOFT KEY 1** to select **SINGLE** wavelength. The reader will measure the optical density of each well with a single filter.

- Press **SOFT KEY 2** to select **DUAL** wavelength. Each well will be read twice, each time with a different filter. **Note:** The microplate is not removed from the reading chamber between the two measurements. The final reported optical density is the difference between the two readings. Dual wavelength readings can significantly reduce optical interference caused by scratched or fingerprinted microplates since the scratches or fingerprints reduce the amount of light on both wavelengths.

- Press **ENTER** to save the selection and advance to the next screen.

MEAS SELECTION

The **MEAS** selection screen allows you to select the filter(s) for the assay. The currently selected filter appears on the top line of the display, and the available options appear on the bottom.

```
M E A S : 4 5 0    R E F : 6 3 0

4 0 5        4 5 0        4 9 0        6 3 0
```

- Press **SOFT KEYS 1, 2, 3,** and **4** to select the filter option displayed above the soft key. The display updates to reflect the selection.

- Press the right arrow key to select the **Reference Filter**.

- Press **ENTER** to move to the next screen.

NUMBER OF KINETIC READS / KINETIC DURATION SELECTION

This menu allows you to either select the total number of kinetic reads or the length of time the assay will run (kinetic duration). Any previously defined value is shown on the top line of the display and the options on the second.

```
K I N E T I C :    T O T A L    R E A D S

T O T A L    R E A D S    D U R A T I O N
```

- Press **SOFT KEY 1** to select the total reads option.

- Press **SOFT KEY 3** to select the duration option.

- Press **ENTER** to save the selection and advance to the next screen.

KINETIC INTERVAL

Use this screen to enter the interval of time (in hours, minutes and seconds) between each kinetic read.

```
K I N E T I C

I N T E R V A L :      0 1 : 2 3 : 5 6
```

- Use the **NUMERIC** keys to enter the time duration. Valid ranges are: 0-1 hours, 0-59 minutes and 0-59 seconds. The number of Reads = Duration / Interval must be less than or equal to 40 and more than or equal to 2.

- Use the **LEFT** and **RIGHT ARROW** keys to move to the next or previous numeric entry fields.

- Press **ENTER** to save the value and advance to the next screen.

KINETIC NUMBER OF READS

Use this screen to enter the number of kinetic reads.

```
T O T A L   N U M B E R

O F   K I N E T I C   R E A D S :        0 6
```

- Use the **NUMERIC** keys to enter the number of reads required. The range is 2 to 40 reads.

- Press **ENTER** to save the entry and advance to the next screen.

KINETIC DURATION

Use this screen to enter (in hours, minutes and seconds) the duration of the kinetic reaction.

```
K I N E T I C

D U R A T I O N :    1 1 : 2 3 : 4 5
```

- Use the **NUMERIC** keys to enter the time duration in hours, minutes, and seconds. The maximum duration time is 80 hours.

- Use the **LEFT** and **RIGHT ARROW** keys to move between entry fields.

- Press **ENTER** to save the entry and advance to the next screen.

SHAKE MODE SELECTION

Use this screen to enter the shake mode for a kinetic or endpoint assay.

```
S H A K E :    B E F O R E    E V E R Y    R E A D

F I R S T    E V E R Y    N O N E
```

- Press **SOFT KEY 1** to select shaking for the first read only or endpoint read.

- Press **SOFT KEY 2** to select shaking for every read.

- Press **SOFT KEY 3** to select **no** shaking.

- Press **ENTER** to save the selection and advance to the next screen.

- **SCREEN** key to save the selection and return to a previous screen.

SHAKE TIME

Use this screen to enter the desired shake interval. "Continuous" appears on the display when a kinetic assay with a previously specified shake has been selected.

```
S H A K E   T I M E :   0 0 : 1 2 : 3 4

C O N T I N U O U S
```

- Use the **NUMERIC** keys to enter the shake interval. Valid ranges are: 0-1 hours, 0-59 minutes and 0-59 seconds.

- Use the **LEFT** and **RIGHT ARROW** keys to move the cursor between hours, minutes, and seconds.

- Press **ENTER** to save the entry and advance to the next screen.

SHAKE SPEED

Use this screen to select the shake speed. The shake movement is a repeated 0.021-inch movement from the shake position and back.

```
S H A K E   S P E E D :     M E D I U M

L O W   M E D I U M   H I G H   V A R I
```

- Press **SOFT KEY 1** to select low-speed shaking.

- Press **SOFT KEY 2** to select medium-speed shaking.

- Press **SOFT KEY 3** to select high-speed shaking.

- Press **SOFT KEY 4** to select variable-speed shaking (1 second of each speed repeated).

- Press **ENTER** to save the entry and advance to the next screen.

KINETIC DATA ANALYSIS SELECTION

Use this screen to select the type of kinetic analysis.

```
K I N E T I C    A N A L Y S I S :    R - S Q R

  R A T E    R - S Q R    O N S E T
```

- Press **SOFT KEY 1** to select the kinetic rate calculation. This method will apply a linear fit to calculate the maximum slope based on the number of kinetic points specified.

- Press **SOFT KEY 2** to select the R-squared rate calculation. This method will calculate the R-squared value at the maximum slope, based on the linear curve fit and the number of kinetic points specified.

- Press **SOFT KEY 3** to select the time calculation, which will calculate the time for each well to reach the onset optical density.

- Press **ENTER** to save the selection and advance.

NUMBER OF KINETIC POINTS SELECTION

Use this screen to select the number of sequential kinetic points to calculate the steepest Rate, or the R squared at the steepest Rate.

```
K I N E T I C    P O I N T S :    3

A L L    P O I N T S
```

- Use the **NUMERIC** keys to input the number of points. The range is 2 to MAX where max is the total number of reads.

- Press **SOFT KEY 1** or **2** to select **ALL POINTS**.

- Press **ENTER** to save the entry and advance to the next screen.

ONSET OD SELECTION

Use this screen to enter the onset OD time.

```
E  N  T  E  R

O  N  S  E  T      O  D  :      1  .  2  3  4
```

- Use **NUMERIC** keys to enter the onset OD. 3.000 OD is the maximum value.

- Use the **LEFT** and **RIGHT ARROW** keys to move the cursor within the entered OD field.

- Press the **ENTER** key to save the entry and advance.

LINEAR SCANNING

If Scanning is chosen as the Read Type, use the following screen to enter the total number of points to be read in a line across the center of each well.

```
E  N  T  E  R      N  U  M  B  E  R      O  F

S  C  A  N      P  O  I  N  T  S  :                    1  5
```

The maximum number of points selectable is 31 (odd numbers only). The 31 scan positions are fixed in the software. You must determine the optimal number of scans per well. If, for example, 7 scans across the well is chosen, the reader will read the centermost seven points in the well. The more scan points chosen, the closer to the well sides reads will be taken.

ⓘ   **Note:** If too many scans are chosen, the reader may be reading the sides of the well.

The reader will read the chosen number of points across the well and report the calculated area under the curve.

MAP

The MAP Definition screen allows you to edit or specify the following options in the assay:

- Automatic or Manual Map Generation

- Mapping Direction

- Replication Direction

- Blank Map Selection

- Blanking Constant

- Number of Blanks

- Location of Blanks

- Number of Standards

- Number of Standard Replicates

- Averaging of Standards

- Concentration and Location of Standards

- Number of Controls

- Control Type Definition

- Number of Control Replicates

- Control Location

- Number of Samples

- Number of Sample Replicates

- Averaging of Samples

- Sample Location

Note: Some of the following screens may not appear, depending on the reader model. The valid range of the number of standards is 0-12. The valid range of valid replicate counts for standards is 1-8. The valid range of the number of controls is 0-8. The range of valid replicate counts for controls and samples is 1-12.

```
D E F I N E :

M E T H O D     M A P     F O R M U L A     C U R V E
```

- At the **DEFINE** Options screen, press **SOFT KEY 2** to begin the plate **MAP** process.

MAP GENERATION

"Map Generation" represents the method by which blanks, controls, standards, and/or samples are assigned to specific locations on the plate. The currently selected value appears on the top line of the display, and the available options appear on the bottom.

```
M A P     G E N E R A T I O N :     M A N U A L

A U T O     M A N U A L
```

Automatic Plate Map Generation: Select **AUTO** to instruct the software to automatically generate a plate map after the blanks, controls, standards, and/or samples have been defined.

Manual Plate Map Generation: Select **MANUAL** to indicate that the well assignments will be performed manually (by the user) at Define and/or Read time.

- Press **SOFT KEY 1** for **automatic** sample plate map generation. The display will update to reflect the selection.

- Press **SOFT KEY 2** for **MANUAL** plate map generation. The display updates to reflect the selection.

- Press **ENTER** to save the selection and move to the next screen.

ⓘ   **Note:** Press **SHIFT-CLEAR** to clear any previously defined manual map.

MAPPING DIRECTION

Use this option to select how the wells are mapped on the plate. Any previously defined Mapping Direction appears on the top line of the display; the available options appear on the second line.

```
MAPPING    DIRECTION:DOWN

 DOWN   ACROSS
```

- Press **SOFT KEY 1** to map **DOWN** the column.

- Press **SOFT KEY 2** to map **ACROSS** the row.

- Press **ENTER** to save the selection and move to the next screen.

REPLICATION DIRECTION

This option allows you to specify how replicates are mapped on the plate. The currently selected Replication Direction appears on the top line of the display, and the available options appear on the bottom.

```
REP   DIRECTION:     ACROSS

 DOWN   ACROSS
```

- Press **SOFT KEY 1** to map the replicates **DOWN** the column, following the direction of the map listing.

- Press **SOFT KEY 2** to map the replicates **ACROSS** (in a paired format). As an example, two replicates can be placed in A1 and A2 wells. The third replicate would follow in B1. The next standard control, or sample, would follow in B2.

- Press **ENTER** to save the selection and advance.

Examples of mapping and replication directions are shown on the next page.

EXAMPLES OF MAPPING & REPLICATION DIRECTIONS

Map Direction DOWN, Rep Direction DOWN:

	1	2	3	4	5	6	7	8	9	10	11	12
A	STD1	STD5	SMP									
B	STD1	STD5	SMP									
C	STD2	PC	SMP									
D	STD2	PC										
E	STD3	NC										
F	STD3	NC										
G	STD4	SMP										
H	STD4	SMP										

Map Direction ACROSS, Rep Direction ACROSS:

	1	2	3	4	5	6	7	8	9	10	11	12
A	STD1	STD1	STD2	STD2	STD3	STD3	STD4	STD4	STD5	STD5	PC	PC
B	NC	NC	SMP	SMP	SMP	SMP	SMP	SMP				
C												
D												
E												
F												
G												
H												

Map Direction DOWN, Rep Direction ACROSS:

	1	2	3	4	5	6	7	8	9	10	11	12
A	STD1	STD1										
B	STD2	STD2										
C	STD3	STD3										
D	STD4	STD4										
E	STD5	STD5										
F	PC	PC										
G	NC	NC										
H	SMP	SMP										

Map Direction ACROSS, Rep Direction DOWN:

	1	2	3	4	5	6	7	8	9	10	11	12
A	STD1	STD2	STD3	STD4	STD5	PC	NC	SMP	SMP			
B	STD1	STD2	STD3	STD4	STD5	PC	NC	SMP	SMP			
C												
D												
E												
F												
G												
H												

START MAPPING AT WELL LOCATION

The Start Mapping at Well Location screen is only shown if automatic mapping is selected. This option allows you to enter the location of the well that will be the starting point for automatic mapping.

```
START   MAPPING

AT   WELL   LOCATION:      A 0 1
```

- Use the **LEFT** and **RIGHT ARROW** keys to move the cursor to the previous or next editable field. The cursor will wrap around the edit field.

- Use the **NUMERIC** and **ALPHA** keys to enter a letter or number at the cursor location. For all prompts of a well location, only the **ALPHA** keys are active for the first character and **NUMERIC** for the second and third characters.

- Press **ENTER** to save the well location and advance to the next screen.

SELECTING A BLANK MAP

This option allows you to select which blanking method to apply to the assay. The currently selected Blank Map value appears on the top line of the display, and the available options appear on the bottom.

The blanking options, **AIR**, **FULL** and **CONSTANT**, **ROW** and **COLUMN**, and **P-ACROSS** and **P-DOWN** are displayed on three screens.

```
BLANK   MAP:        FULL

  AIR      FULL    CONST   * MORE
```

```
BLANK   MAP:        FULL

  ROW    COLUMN              * MORE
```

```
BLANK   MAP:        FULL

   P - ACROSS    P - DOWN   * MORE
```

- Press **SOFT KEYS 1, 2,** or **3** to select the **BLANK MAP** type above the soft key. The display updates to reflect the selection.

- Press **SOFT KEY 4** to access **MORE** options: **ROW** or **COLUMN**, and **P-ACROSS** or **P-DOWN**.

- Press **ENTER** to save the well location and advance to the next screen.

BLANK MAP DEFINITIONS:

- **AIR** performs an initial reading on "air" just prior to the plate read, and uses that value as the blank value. This value is subtracted from each well on the plate.

- **FULL** enables a single blank well or an average of blank wells to be subtracted from the whole plate.

- **CONST** (Constant) allows entry of a user-specified absorbance value. This value will be subtracted from each well on the plate. Use a blank value from the first plate, or a blanking plate to save space on subsequent assay plates.

- **ROW** enables a single blank well or an average of blank wells to be selected for each row. The blank (or average) will be subtracted from each well in the row. Use manual mapping to position blanks, controls, standards, and samples.

- **COLUMN** enables a single blank well or an average of blank wells to be selected for each column. The blank (or average) will be subtracted from each well in the column. Use manual mapping to position blanks, controls, standards, and samples.

- **P-DOWN** enables a blank in every even numbered column to be subtracted from the well to the left of it in every odd column. Manual mapping is recommended to set up the appropriate map by placing the standards, controls, and samples in only the odd columns.

- **P-ACROSS** enables a blank in the B, D, F and H rows to be subtracted from the well above in the A, C, E and G rows. Manual mapping is recommended to set up the appropriate map by placing the standards, controls, and samples in only the A, C, E, and G rows.

CONSTANT BLANK VALUE ENTRY

This entry screen only appears when a Constant Blank map is selected. Enter the value to be subtracted from each well on the plate.

```
E N T E R

B L A N K I N G    C O N S T A N T :    0 . 0 0 0
```

- Use the **NUMERIC** keys to enter the value. The range is 0.000 to 3.000. The cursor is positioned at the first editable field and advances automatically.

- Press the **CLEAR** key to clear the value on the display.

- Press the **ENTER** key to save the value and advance.

NUMBER OF BLANKS

The Number of Blanks field allows you to enter the number of blank wells in the assay. This entry screen is only displayed when Full, Column, or Row blank maps is selected.

```
E N T E R    N U M B E R    O F

B L A N K S :                          0 2
```

- Use the **NUMERIC** keys to enter the number of blanks. The range is 0 to 48.

- Press the **CLEAR** key to clear the Number of Blanks value from the display.

- Press the **ENTER** key to save the value and move to the next screen.

SELECTING A BLANK LOCATION

The Blank Location screen allows you to define where the blank well is located on the microplate. This screen only appears if manual map generation has been selected. Any previously defined value is displayed.

```
E N T E R    T H E    L O C A T I O N    O F

B L A N K  # 1 :                              A 1 2
```

- Use the **NUMERIC** and **ALPHA** keys to enter a Blank Location, based upon the plate geometry.

- Use the **ARROW** keys to move the cursor to the next or previous editable field. The cursor is positioned beneath the first editable field.

- Press the **ENTER** key to save the value, and move to the next screen.

NUMBER OF STANDARDS

Use this option to enter the number of standards that will be used in the assay. Any previously defined value will be displayed on the screen. If the number of standards is altered, the number of replicates for the standard automatically defaults to 1.

```
E N T E R    N U M B E R    O F

S T A N D A R D S :                            0 2
```

- Use the **NUMERIC** keys to enter the Number of Standards. The valid range depends on the selected curve fit method. The maximum number of standards is 12. The minimum is 4 for 4-P fit, cubic, cubic spline, and 2-P; 3 for quadratic; and 2 for linear and point-to-point.

- Press **CLEAR** to clear the value on the display.

- Press **ENTER** to save the value, and move to the next screen.

NUMBER OF STANDARD REPLICATES

This option allows you to enter the number of replicates per standard in the assay. Any predefined value appears on the display.

```
E N T E R    N U M B E R    O F

S T A N D A R D    R E P L I C A T E S :      0 2
```

- Use the **NUMERIC** keys to enter the Number of Standard Replicates. The range is 1 to 8 replicates. The software will verify that the number of replicates, multiplied by the number of standards, does not exceed the number of wells on the plate.

- Press **CLEAR** to clear the value on the display.

- Press **ENTER** to save the value and move to the next screen.

AVERAGE STANDARDS

The Average Standards option allows you to select whether or not to average the Replicates of a Standard. This average is used to calculate the standard curve instead of using the individual replicate of each standard. **Note:** If the replicate selection is 1, this option is not available.

```
A V E R A G E    S T A N D A R D S ?    Y E S

  Y E S            N O
```

- Press **SOFT KEY 1** to select **YES** (average the replicates). The top line of the display updates to reflect the selection.

- Press **SOFT KEY 2** to select **NO** (do **not** average the replicates). The top line of the display updates to reflect the selection.

- Press the **ENTER** key to save the selection, and advance to the next screen.

STANDARD CONCENTRATION

The Standard Concentration field allows you to enter the predicted or expected concentration value for each standard group. Any previously defined value is displayed.

If **Manual Map Generation** is selected, the replicate locations must also be defined.

```
C O N C N    O F          L O C A T I O N

S T D 1 : 1 . 5 0         R E P #   1 : A 0 1
```

- Use the **NUMERIC** and **ALPHA keys** and the **DECIMAL POINT** key to enter Standard Concentration values. The range is 0.00001 to 999999. The entry cannot exceed six characters including the decimal point. Valid well locations for the defined geometry are listed below.

- Use the **RIGHT ARROW** and **LEFT ARROW** keys to move to the next or previous editable field.

- Press the **CLEAR** key to clear the Standard Concentration value from the display.

- Press the **ENTER** key to save the value on the display and move to the next screen.

REUSE OF STANDARD CURVES

The ELx808™ has the ability to reuse a standard curve that has already been established.

Limitations of the Reuse of Standard Curves

- The reuse of standard curves can only be done in assay positions 31 through 55. Each of these positions can only store one standard curve.

- Standard curves cannot be reused on panels (see page 3-47 for Panel Definition).

- Standard curves will be stored in memory with the Assay Name, Standard Concentrations, Replicate Counts, and Optical Densities for each standard replicate.

- Stored standard curves can only be reused for the assay that the curve was originally generated on (i.e., the curve for assay #53 cannot be applied to samples on a plate to be run in assay #51).

- To reuse a standard curve, an assay must first be programmed (in positions 31 through 55) and run. During the defining process, you will be prompted to enter the number of standards, the number of standard replicates, and the standard concentrations. The following screen has been added after these prompts:

```
R E U S E    S T A N D A R D    C U R V E ? Y E S

  Y E S        N O
```

- After the assay has been run, the results have been calculated, and the reports have been generated, the reader will prompt if this standard curve should be stored in memory. The following display will appear:

```
S A V E    S T A N D A R D    C U R V E ?    Y E S

  Y E S        N O
```

- Select **YES** to store the curve for use at a later time. The next time a plate is to be read using this assay, the instrument will prompt if there are standards on the plate. Select **NO** to discard the curve.

```
S T A N D A R D S     O N     P L A T E ?        N O

Y E S     N O
```

- If **YES** is chosen, a new standard curve will be generated. The plate map is not changed. (If "Prompt for Sample ID" is enabled in the **UTIL** section, you will be prompted to enter the number of samples. See page 2-11 for more information on the **UTIL** options.)

- If **NO** is chosen, the stored standard curve will be used. If Auto mapping had been used to originally map the standards, blanks, controls, and samples defined for this assay, the map will be automatically regenerated without the standards, beginning in well xxx (where xxx was chosen as the Starting well in the map, usually well A01). If Manual mapping was used to map the plate, the map is NOT regenerated - the reader will NOT produce results for the well positions that originally were standards. Auto mapping is recommended, if the standards curves will be routinely reused.

NUMBER OF CONTROLS

The Number of Controls screen allows you to enter the number of controls that will be used in the assay. Any previously defined value will appear on the display.

```
E N T E R     N U M B E R     O F

C O N T R O L S :                         0 2
```

- Use the **NUMERIC** keys to enter the Number of Controls. The range depends on the number of locations on the plate that are undefined. The maximum number of controls is 8.

- Press **CLEAR** to clear the value on the display.

- Press **ENTER** to save the value and advance to the next screen.

CONTROL TYPE

This option allows you to enter the type of control used in the assay. Any previously defined Control Type will be displayed on the screen.

```
C O N T R O L # 1 :                    P C

    P C        N C        H P C      * M O R E
```

```
C O N T R O L # 1 :                    P C

  L P C        C T L 1      C T L 2    * M O R E
```

```
C O N T R O L # 1 :                    P C

  C T L 3        C T L 4            * M O R E
```

- Press the soft keys under the displayed **Control Type** to select the option (**P**ositive **C**ontrol**, N**egative **C**ontrol, **H**igh **P**ositive **C**ontrol, **L**ow **P**ositive **C**ontrol, CTL1, CTL2, CTL3, CTL4).

- Press **CLEAR** to clear the Control Type from the display.

- Press **ENTER** to save the displayed Control Type and advance to the next screen.

NUMBER OF CONTROL REPLICATES

The Number of Control Replicates screen allows you to enter the number of replicates per control group in the assay. Any previously defined number will be displayed.

```
E N T E R   N U M B E R   O F

R E P L I C A T E S   O F   P C :        0 2
```

- Use the **NUMERIC** keys to enter a value for Number of Control Replicates. The range is 1 to 12 replicates. The software performs a check to ensure the number of replicates does not exceed the number of undefined wells remaining on the plate.

- Press **CLEAR** to clear the displayed Number of Replicates value.

- Press **ENTER** to save the displayed value and advance to the next screen.

LOCATION OF CONTROLS

Use this option to enter the location of controls in the assay. The displayed location field can only be edited if manual map generation was selected.

```
CONTROL # 1                 LOCATION

TYPE : PC                   REP # 1 : A 0 1
```

- Press the **CLEAR** key to clear the value on the display.

- Use the **NUMERIC** and **ALPHA** keys to enter values for well locations.

VALID WELL LOCATIONS

For all prompts of a well location, only the **ALPHA** keys are active for the first character and **NUMERIC** for the second and third characters.

- Press the **MAIN MENU** and **PREVIOUS SCREEN** keys to move back and advance through the menu structure.

NUMBER OF SAMPLES

This option allows you to enter the number of sample groups that will be used in the assay. Any previously defined value appears on the display. If the number of samples is altered, the number of replicates for the sample reverts to a value of 1.

```
ENTER   NUMBER   OF

SAMPLES :                       2 4
```

- Use the **NUMERIC** keys to enter the number of samples in the assay. The range is 0 to the number of undefined well locations remaining on the plate. If there are no controls, blanks, or standards defined on a 96-well plate, the maximum number of samples is 96, and the minimum number of samples is 1.

- Press the **CLEAR** key to clear values on the display.

- Press **ENTER** to save the displayed value and advance to the next screen.

NUMBER OF SAMPLE REPLICATES

Use this option to enter the number of replicates per sample that will be run in the assay. Any previously defined value will be displayed.

```
E N T E R    N U M B E R    O F

S A M P L E    R E P L I C A T E S :        0 2
```

- Use the **NUMERIC** keys to enter the Number of Sample Replicates. The range is 1 to 12 replicates. The software ensures that the number of replicates multiplied by the number of samples, multiplied again by the number of dilutions, does not exceed the number of undefined wells remaining on the plate.

- Press the **CLEAR** key to clear the value on the display.

- Press the **ENTER** key to save the displayed value and advance to the next screen.

SAMPLE LOCATION

If **Manual Map Generation** is selected, this screen allows you to select the well location of the sample on the plate. Any previously defined Sample Location appears on the display.

```
S A M P L E  # 1            L O C A T I O N

                            R E P #   1 : A 0 1
```

- Use the **NUMERIC, ALPHA,** and **DECIMAL POINT** keys to enter the sample identifier and its location on the plate.

- Press the **CLEAR** key to clear the displayed value.

- Press **ENTER** to save the value and advance to the next screen.

FORMULA

Defining the Formula involves identifying the **formula type** and entering the actual **formulas**. Formula definition screens are displayed on the ELx808™ in the order detailed below. Formulas created using BioTek's Extensions® (Define Protocol software) cannot be edited by using the reader.

Formulas are processed in the following order, with the number of permitted formulas of each type:

- Blank Validation   0-1

- Control Validation 0-4

- Assay Validation   0-4

- Transformations   0-1

- Cutoff Formulas   0-1

- Curve Fit Analysis (if a curve fit method is defined)

Within any given formula type, the order of processing is the order in which the formulas are entered.

VALIDATION FORMULA EXAMPLES

- **Blank Validation:** An assay protocol states that every blank well on a plate should have an OD of less than 0.050. The formula is entered on the reader as a Blank Validation Formula: *BLK < 0.050*

- **Negative Control Validation:** An assay protocol states that every Negative Control well must have an OD of less than or equal to 0.100. The formula is entered as a Control Validation Formula: *NC < = 0.100*

- **Positive Control Validation**: An assay protocol states that every Positive Control well must have an OD higher than 1.000, but less than 2.500. Two Control Validation Formulas can be entered: *PC > 1.000 AND PC < 2.500*

  Or, one formula can be used if the formula is 24 characters or less: *PC > 1.000 AND PC < 2.500*

- **Assay Validation**: An assay protocol states that in order for an assay to be valid, the mean of the Negative Control well ODs must be less than 0.100. The Assay Validation formula that should be entered: *NC;x < 0.100* (the map identifier NC;x indicates the mean of the NCs)

From the assay **DEFINE** Menu, press the arrow corresponding to **FORMULA**.

Formula Type

The ELx808™ supports three types of formulas, as well as the ability to program variables for use within Transformation formulas.

- CUTOFF formulas are used to classify results. During data reduction, results are evaluated against the cutoff formulas, and each well is assigned a user-specified label or "call" (POS, NEG, or EQUIV).

- TRANSformation formulas are applied to the raw data in preparation for further data reduction and/or curve-fit calculation.

- VALidation formulas can be used to determine whether or not blanks and/or controls are valid. In addition, Assay Validation formulas can be used to determine whether or not the entire assay should be considered valid.

- The TRANS-VAR option allows you to define a variable to be used in transformation formulas.

- **Note:** GENERAL formulas are not used in the ELx808 open assays.

```
SELECT    FORMULA    TYPE:

CUTOFF    TRANS      VAL       *MORE
```

```
SELECT    FORMULA    TYPE:

GENERAL    TRANS-VAR      *MORE
```

- Press **SOFT KEY 1** to select CUTOFF Formula.

- Press **SOFT KEY 2** to select TRANSFORMATION Formula.

- Press **SOFT KEY 3** to select ASSAY VALIDATION Formula.

- Press ***MORE** to access additional formula types.

- Press **SOFT KEY 3** to select TRANS-VAR.

- Press **ENTER** to save the selected formula type and advance to the next screen.

VALIDATION TYPE SELECTION

If you selected VAL, this option allows you to choose which Validation Formula type (Control, Assay, or Blank Validation formulas) to enter for the assay.

```
SELECT   VALIDATION   TYPE:
CONTROL          ASSAY    BLANK
```

- Press **SOFT KEY 1** to select Control Validation Formula.

- Press **SOFT KEY 3** to select Assay Validation Formula.

- Press **SOFT KEY 4** to select Blank Validation Formula.

- Press **ENTER** to save the validation type and advance.

FORMULA ENTRY

This screen allows you to enter the formula used in the assay.

ⓘ   **Note:** In formulas, "OD" is used to represent the optical density value.

```
F O R M U L A # 1 :

 M A T H   O T H E R    M A P    F U N C T N
```

- After a moment, the **FORMULA #1:** prompt disappears, and the formula can be entered.

- Each formula can contain a maximum of 24 characters. Spaces are unnecessary.

- Use the **LEFT and RIGHT ARROW** keys to move the cursor to the previous or next editable field.

- Press **SOFT KEY 1** to place the next item on the **MATH** list at the cursor position. The following table shows the order of items on the **MATH** list:

+	Addition sign
-	Subtraction sign
*	Multiplication sign
/	Division sign
%	Percent
=	Equal
>	Greater than
>=	Greater than or equal to
<	Less than
<=	Less than or Equal to

Note: The reader software checks the formulas for errors during data reduction. A syntax error in a formula will result in a "Token Error" on results reports.

- Press **SOFT KEY 2** to place the next item on the **OTHER** list at the cursor position. See the table that follows for the order of items on the OTHER list:

(	Left parenthesis
)	Right parenthesis
AND	Logical AND
OR	Logical OR

- Press **SOFT KEY 3** to place the next defined item on the plate map list (i.e., STD, NC, PC, BLK) at the cursor position.

- Press **SOFT KEY 4** to place the next option on the **FUNCTION** list at the cursor position. The available functions are:

LOG10	Log Base 10
ALOG10	Anti Log Base 10
AB	Absolute Value
PWR	Power
ALOG	Anti Log
LOG	Log
SQRT	Square Root

EXAMPLES

LOG10: Log Base 10   **ALOG:** Anti Log
Log10 2 = 0.301029995   ALOG (0.69314718) = 2

ALOG10: Anti Log Base 10   **LOG:** Log
ALOG10 (0.30102995) = 2   LOG 2 = 0.69314718

AB: Absolute Value   **SQRT:** Square Root
AB (-1) = 1   SQRT 2 = 1.4142

PWR: Power
(10 PWR 2) = 100

- Well locations, well types, or numbers in parentheses precede functions.

- Press the **CLEAR** key to clear the item displayed at the cursor position.

- Press the **SHIFT** and **CLEAR** keys to clear the entire entry.

- Press **ENTER** to save the displayed value and advance to the next screen, or use the **PREVIOUS SCREEN** key to move backward through the menu structure.

NUMBER OF REQUIRED CONTROLS / BLANKS

If a control or blank validation formula is entered, this screen allows you to enter the number of valid controls / blanks for the assay. Any previously defined values will appear on the display.

```
P C :    N U M B E R    O F    V A L I D

R E P L I C A T E S    R E Q U I R E D ?    0 2
```

- Use the **NUMERIC** keys to enter the Number of Required Controls. The range is 1 through the number of defined replicates of a control or blank.

- Press the **CLEAR** key to clear the displayed value.

- Press the **ENTER** key to save the displayed value and advance to the next screen, or use the **PREVIOUS SCREEN** key to move backward through the menu structure.

CUTOFF FORMULAS

A cutoff formula calculates a **cutoff value** that is used for classifying samples.

During data reduction, results are evaluated against the cutoff value (with an optional greyzone), and each well is assigned a call POS (positive), NEG (negative), or EQUIV (equivocal).

- One cutoff formula may be defined per assay.

- If Transformation Formulas are defined, cutoffs are based on the transformed results. Refer to "FORMULA" on page 3-31 for the order in which formulas are processed.

- A cutoff formula can consist of a simple numeric value (1.500), a well identifier (PC or PC;x to represent the mean), or a formula combining the two (NC;x+0.050).

- A "greyzone" around the cutoff value can be defined, to indicate equivocal or indeterminate results.

- Do not use the < or > mathematical symbols in a cutoff formula.

- **Tip:** Choose to print a Column Report to see the greyzone and cutoff values as well as the equations used to assign calls to samples.

After selecting an assay (page 3-5), define the Cutoff formula as shown below:

```
DEFINE:

METHOD   MAP   FORMULA   CURVE
```

↑

```
SELECT FORMULA TYPE:

CUTOFF  TRANS    VAL    *MORE
```

↑

```
FORMULA #1:

  MATH   OTHER    MAP    FUNCTN
```

- After the **FORMULA #1:** prompt disappears, enter the formula as described in "Formula Entry" on page 3-34. Refer to the examples on page 3-39.

GREYZONE ENTRY

The **greyzone** is a definable area around the cutoff value. Samples falling within an area defined by the greyzone (ex. ± 5.0% of the cutoff value) could be considered equivocal (EQUIV).

```
E N T E R   G R E Y Z O N E :         0 5 %



```

- Use the **NUMERIC** keys to enter the greyzone percentage.

- The valid entry range is from 00 to 99%. An entry of 00% indicates no greyzone, although a sample equal to the cutoff value will still receive the EQUIV call.

- See *POS / NEG Calls* below for information on how calls are assigned.

- Press the **CLEAR** key to clear the displayed value.

- Press the **ENTER** key to save the displayed value and advance.

POSITIVE / NEGATIVE CALLS FOR CUTOFF

After the greyzone is defined, calls for the sample wells (POSitive, NEGative, EQUIVocal) must be defined.

```
S A M P L E > C U T O F F + 0 5 % :     P O S

  P O S       N E G
```

- Select **POS** or **NEG** to select the call that will be assigned to samples greater than the cutoff value plus the greyzone.

- If, for example, POS is selected as shown in the above screen, calls will be assigned according to the following equations (SMP represents the sample wells):

  EQUIV: SMP <= (CUTOFF+(CUTOFF*GREYZONE)) AND

  SMP >= (CUTOFF-(CUTOFF*GREYZONE))

  POS: SMP > (CUTOFF+(CUTOFF*GREYZONE))

  NEG: SMP < (CUTOFF-(CUTOFF*GREYZONE))

Examples

1: The cutoff between negative and positive calls should be calculated as the average of the negative controls plus the OD value of 0.500. Samples greater than the cutoff should be labeled as positive. No greyzone is required.

- For this example, NC;x (the mean of the NC wells) equals 1.000 OD

- The cutoff formula is `NC;x+0.5`

- The greyzone is `00%`

- POS is selected for `SAMP>CUTOFF+00%`

- Calls are assigned to sample wells as follows:

    ➢ EQUIV if the sample equals 1.500

    ➢ POS if the sample is greater than 1.500

    ➢ NEG if the sample is less than 1.500

2: For a quantitative assay, samples with OD values greater than the STD2 mean plus a 10% greyzone should be labeled as positive; samples with OD values less than the STD2 mean minus the 10% greyzone should be labeled as negative. All other samples should be considered equivocal.

- For this example, STD2;x (the mean of the STD2 wells) equals 2.000 OD

- The cutoff formula is simply `STD2;x`

- The greyzone is `10%`

- POS is selected for `SAMP>CUTOFF+10%`

- Calls are assigned to sample wells as follows:

    ➢ EQUIV if the sample is greater than or equal to 1.800 *and* less than or equal to 2.200

    ➢ POS if the sample is greater than 2.200

    ➢ NEG if the sample is less than 1.800

TRANSFORMATION FORMULAS

Transformation formulas change the absorbance data of all wells defined in the Map to another format, in preparation for further data reduction.

TRANSFORMATION FORMULA DEFINITION

- From the assay **Define** Menu, press the arrow corresponding to **Formula**.

```
D E F I N E :

M E T H O D    M A P    F O R M U L A    C U R V E
```
⬆

- This will bring you to a screen asking to **Select Formula Type**. At this screen, select **TRANS** and then enter the formula using the **Math, Other, Map** and **Function** keys.

```
S E L E C T    F O R M U L A    T Y P E :

C U T O F F    T R A N S    V A L    * M O R E
```
⬆

Example: Divide all ODs on the plate by 2 and multiply by 100.

Enter the formula: **(OD/2)*100**

This formula will be applied to the ODs of all samples, standards, controls, and blanks that are present on the plate map.

TRANSFORMATION SCOPE VARIABLE

For more complex transformations, a *Transformation Scope Variable* (TVar) can be defined for use with a transformation formula. This variable defines the *scope* of the transformation: whether to apply the transformation to just the samples (SMP) or to all wells defined on the plate (OD).

From the assay **Define** Menu, press the arrow corresponding to *Formula*.

```
D E F I N E :

M E T H O D    M A P    F O R M U L A    C U R V E
```
↑

This will bring you to a screen prompting you to **Select Formula Type**. Press ***MORE** at this screen*.

```
S E L E C T    F O R M U L A    T Y P E :

C U T O F F    T R A N S    V A L      * M O R E
```
↑

The options displayed now include **TRANS-VAR**. Press **VAR** at this screen.

```
S E L E C T    F O R M U L A    T Y P E :

G E N E R A L    T R A N S - V A R     * M O R E
```
↑

The following screen will appear, asking you to choose the *scope* of this transformation.

```
S C O P E    V A R I A B L E :          O D

  S M P       O D
```

If **SMP** is chosen, the transformation formula will be applied *only* to the samples defined in the plate map. If **OD** is chosen, the formula definition screen will appear. Use the formula keys (Math, Other, Map and Function) to define the *transformation variable (TVar)*. Once the variable has been defined, it can be used in a transformation formula. The TVar will be available as a MAP option when writing the transformation formula.

Example: An assay plate map has 2 blanks, 1 control well in duplicate (CTL1), 1 negative control well in triplicate (NC), and 5 standards in duplicate (STD1-STD5) with varying concentrations.

The assay data reduction states:

- Subtract the mean of CTL1 from the mean of the NC. Subtract the difference from all ODs on the plate.

- Divide the result of the above by the means of the NC less the means of CTL1, and then multiply by 100.

On paper, the formula reads:

(OD-(NC;x-CTL1;x))/(NC;x-CTL1;x)*100

- On the reader, define the formula *(NC;x-CTL1;x)* as the *Transformation Variable,* since the transformation will apply to all standards, controls and samples on the plate.

- At the SCOPE VARIABLE screen, choose **OD** and press **ENTER**. Now enter the formula **(NC;x-CTL1;x)** by using the **MATH, OTHER, MAP** and **FUNCTION** keys. Press the **ENTER** key.

- The formula definition screen is displayed. Choose **TRANS**.

- Enter the following formula: **(OD-(TVar))/(TVar)*100** using the **MATH**, **OTHER**, **MAP,** and **FUNCTION** keys. (TVar is included in Map options on the formula entry screen.) The transformation formula has been added to the assay definition.

ANOTHER TRANSFORMATION EXAMPLE:

In the case of competitive reactions, converting absorbance data to percent B/B_0 can be: (OD/Std1)*100. This divides all the wells by **STD1** (presumably the 0 standard), and multiplies the results by 100.

- At the SCOPE VARIABLE screen, choose OD and press **ENTER**.

- Select STD1 from **MAP** and Press **ENTER**.

- Choose **TRANS**.

- Enter (**OD/TVAR**) * 100

CURVE

The **CURVE** entry screens allow editing and entry of:

- Curve-Fit Type

- Editing of Outliers

- Axis Identification

- Extrapolation of Unknowns

These screens are displayed on the ELx808™ in the order in which they appear in the assay. If a closed variable (i.e., an element of the assay definition that you cannot access or modify) is being used in the assay, the entry screen is omitted.

Curve Fit

The **Curve-Fit** screen allows you to select the curve-fit method that will be applied to the assay. Any previously defined curve-fit type appears on the top line of the display, and available options on the second line.

The **Curve-Fit** screen has three sub-menu screens. Each sub-menu screen provides different curve-fit options for selection. These options include C-Spline, Linear, Quadratic, Cubic, 4-P, 2-P (Logit/Log), PT to PT (point to point), and None.

- **Linear curve fit:** A simple best-fit straight line is plotted using the values of the standards.

- **Quadratic or "Quad" curve fit:** A curve fit which uses the Quadratic equation "$ax^2 +bx + c = y$" to plot the standard's values. Utilizing this curve, any data point for a standard that deviates from the ideal value will not affect the entire curve.

- **Cubic curve fit:** A curve fit that uses the equation "$ax^3 + bx^2 + cx + d = y$" to plot the standard's values. This type of curve fit is affected even less than the quadratic fit when any particular standard has a poor value.

- **2-P (LOGIT/LOG):** A curve fitted to the standard values, which is characterized by a skewed sigmoidal (S-shaped) plot that eventually becomes asymptotic to the upper and lower standard values. The logistic equation is algebraically transformed to a simpler form in which experimentally determined values are used for the responses at concentrations of zero and infinity.

- **Cubic Spline (C-Spline) curve fit:** A piecewise polynomial approximation consisting of joining a set of data points by a series of straight lines, which is then smoothed by using a cubic fit.

- **4-Parameter Logistic or "4-P":** A curve fitted to the standard values, which is characterized by a skewed sigmoidal (S-shaped) plot that eventually becomes asymptotic to the upper and lower standard values. The 4 parameters are: Left asymptote, Right asymptote, Slope and Value at the Inflection point. This fit is most recommended for immunoassay data, and is more exact than Logit/Log.

- **Point to Point or "PT to PT":** A plot that connects each standard point with a line, with no averaging of the values to "smooth" the curve at each standard.

```
C U R V E - F I T    T Y P E :    C - S P L I N E

  N O N E    L I N E A R    Q U A D     * M O R E
```

```
C U R V E - F I T    T Y P E :    C - S P L I N E

   C U B I C      4 - P      2 - P     * M O R E
```

```
C U R V E - F I T    T Y P E :    C - S P L I N E

    C - S P L I N E      P T - P T    * M O R E
```

- Press **SOFT KEYS 1, 2, 3,** or **4** to select the curve-fit type that is displayed above the soft key. Select the soft key below the menu option **MORE** to display additional options. The top line of the display updates to reflect this selection.

- Press **ENTER** to save the selection and advance to the next screen, or use the **MAIN MENU** and **PREVIOUS SCREEN** keys to move backward through the menu structure.

EDIT STANDARD OUTLIERS

This screen allows you to select which method (None or Manual) will be used to edit Standard Outlier values. After the standard curve has been calculated, one or more standards can be excluded from the recalculation of the curve. Any previously defined edit method is displayed.

```
E D I T    S T D    O U T L I E R S : M A N U A L

 N O N E    M A N U A L
```

- Press **SOFT KEY 1** or **2** to select the edit option displayed above the soft key. The display updates to reflect your selection.

- Select **NONE** to suppress the Edit Standard Outliers capability for this assay.

- Choose **MANUAL** to enable the capability.

  ➢ If **AVERAGE STANDARDS** is set to **NO**, the individual standard replicates are available for editing. If set to **YES**, the standard *groups* are available for editing.

  ➢ After the assay is run and reports are generated, press **REPORT** from the Main Menu. Press **RESULT**, select the assay, and then press **ENTER**. The **EDIT STD OUTLIERS? YES/NO** prompt will appear. See *Editing Standard Outliers* later in this chapter for further instructions.

- Use the **ENTER** key to save the selection and advance to the next screen, or use the **MAIN MENU** and **PREVIOUS SCREEN** keys to move backward through the menu structure.

AXIS SELECTION

This screen allows you to select the X and Y Axis Type. Any previously defined axis type will be displayed. This option screen appears only if Manual Map Generation has been selected.

```
X / Y   A X I S   T Y P E :   L I N

L I N    L I N / L O G   L O G   L O G / L I N
```

- Press **SOFT KEY 1, 2, 3,** or **4** to select the method by which the X and Y-axes will be scaled. The top line of the display updates to reflect the selection.

- This option is not available for the 2-P and 4-P curve-fit types. The X/Y scaling for these curves is always LIN/LIN.

- Press **ENTER** to save the selection and advance to the next screen.

EXTRAPOLATION OF UNKNOWNS

This screen allows you to choose whether to extrapolate the curve to evaluate samples outside of the absorbance range defined by the standards. Any previously defined decision appears on the screen.

```
E X T R A P O L A T E   U N K N O W N S ? Y E S

  Y E S        N O
```

- Press **SOFT KEY 1** to select **YES** (extrapolate the unknowns). The top line of the display updates to reflect this selection.

- On the printed reports, extrapolated concentrations (RSLT values) are surrounded by < > (e.g., <44.425>).

- Press **SOFT KEY 2** to select **NO**. The top line of the display updates to reflect this selection.

(i) **Note:** If extrapolation is chosen for the Point-to-Point curve fit, unknown concentrations will be extrapolated linearly from the nearest segment of the curve. If the plot includes both increasing and decreasing segments, the curve printout will be labeled "Ambiguous." The resulting values, which actually are extrapolated, may not be indicated as such. All calculated results for an "Ambiguous" curve should be considered unreliable.

Panel Assays

A Panel assay is a collection of up to 8 assays to be run on one plate.

- The most common reason to use a Panel assay is for confirmatory tests based on a screening test in clinical applications.

- Only one panel assay can be defined on the reader at any time.

- The assays specified within the Panel must be predefined in any of the assay positions 1-55.

- The assays specified within the panel must all use the Endpoint read method.

- The assays specified within the panel must all read at the same wavelength(s).

- Any curve-fit type, formulas, or standard concentrations previously defined for each assay will be used when the assay is selected for a Panel.

- Panel assays cannot reuse standard curves.

- The type and number of controls, blanks, standards, and replicates in the assays chosen for the Panel will be "copied" into the Panel definition. To change any of the map or assay parameters in the Panel, they must be changed in the predefined assay first.

- Consider printing a Map Report for each assay that will be included in the panel, for use with mapping the Panel.

To create a panel assay, start at the Main Menu, select **DEFINE**, then choose assay number **99**. Enter the panel assay name.

```
N A M E :        P A N E L

   –              /            :         S P A C E
```

- The default name is **"PANEL".**
- Use the **ALPHA** and **NUMERIC** keys to update the Assay name, if desired.
- Press **ENTER** to continue. The Number of Assays entry screen will appear.

```
┌─────────────────────────────────────────────────────────────────┐
│                                                                   │
│  N U M B E R    O F    A S S A Y S :              2               │
│                                                                   │
│                                                                   │
└─────────────────────────────────────────────────────────────────┘
```

- Specify the number of assays to include in the panel (1 to 8).

- Press **ENTER** to continue. The Mapping Direction selection screen will appear.

```
┌─────────────────────────────────────────────────────────────────┐
│                                                                   │
│  M A P P I N G    D I R E C T I O N : D O W N                     │
│                                                                   │
│  D O W N      A C R O S S                                         │
└─────────────────────────────────────────────────────────────────┘
```

- This option ensures that all assays will be mapped in the same direction.

- Select **DOWN** or **ACROSS**.

 (i)   **Note:** The original mapping directions for the predefined assays are
        overridden by the Panel's mapping direction. If the assays include
        replicates, they will follow the Panel mapping direction.

After selecting the mapping direction of the assays, choose which assays to include in the panel.

```
┌─────────────────────────────────────────────────────────────────┐
│                                                                   │
│  S E L E C T    A S S A Y    N U M B E R :      2 2               │
│                                                                   │
│  N A M E :    H B S - A G 1                                       │
└─────────────────────────────────────────────────────────────────┘
```

- Press **OPTION** to cycle through the assay numbers and names, or use the
  NUMERIC keys to enter an assay number. Press **ENTER** to make a selection.

- After selecting an assay, you must define its starting location.

```
S T A R T    M A P P I N G

A T    W E L L    L O C A T I O N :        A 0 1
```

- Use the **ALPHA** and **NUMERIC** keys to choose the well location to begin the assay. Wells A01 through H01 are valid for ACROSS mapping; A01 through A12 are valid for DOWN mapping.

- This process will be repeated for each assay within the panel. Remain aware of the total number of controls, standards, and blanks that were originally mapped in each individual assay while mapping for the Panel assay.

- For example, to include Assays 1, 8, and 22 in the Panel assay (DOWN mapping is selected for the Panel):

  ➢ Assay 1 has a total of 12 wells defined for controls, blanks, and standards. In the Panel, the mapping for Assay 1 begins in well A01. The user wants to run 6 samples in Assay 1. Assay 1 now fills wells A01 through B03.

  ➢ The mapping for Assay 8 can begin in well B04, or any well other than A01 to B03. The reader will "beep" if you try to map into a well that is already assigned for use with the Panel.

  ➢ The mapping for Assay 22 may begin at the next available well location after Assay 8 mapping is complete.

  ➢ After all the assays have been entered into the Panel, consider printing the Panel's Map Report to verify the map before reading the plate. Choose Report (from the Main Menu), Map, and Assay 99. The reader will print the map of each assay configured in the Panel.

  ➢ The Panel Assay results are sorted by Sample (unless a custom assay has been programmed by BioTek). **Note:** The Interpretation of Results reports for each assay in the panel will print first, and then the Sample results will print.

Reading a Microplate

Press the **READ** option, found at the Main Menu, to read a microplate.

- From the **MAIN MENU** screen, press the soft key beneath the **READ** menu option to access the **SELECT ASSAY NUMBER** screen.

- Alternately, press the red **READ** key on the lower right of the keyboard.

SELECT ASSAY

At the **Select Assay Number** screens**:**

- Use the **NUMERIC** keys to enter the number of any predefined Assay Definition Files stored in the reader's memory, or the **OPTION** key to advance one assay at a time. The cursor is positioned at the first editable field and advances automatically. The numeric range depends on the number of assays (1-55) programmed in the reader's memory.

  The assay's name and number are displayed on the screen.

```
S E L E C T    A S S A Y    N U M B E R :      2 5

N A M E : H B S - A G
```

- Press **ENTER** to advance to the Run-Time prompts.

- **MAIN MENU**: Returns the display to the Main Menu screen.

RUN-TIME PROMPTS

After the assay is selected, you may be prompted for information (depending on preferences selected in the reader's Utilities), whether a manual map was set up, or if the assay was created and downloaded from BioTek's Extensions® Define Reader Protocol software.

If the assay was created using Extensions® software, prompts might include:

- The number of samples

- Standard Concentrations

- Assay ID

- Fill Pattern

- Blank Method

- First Well Location

- Replicate Count for each well type

- Wavelength Mode

- Report preferences, etc.

Refer to the Extensions Define Protocol Software User's Manual for more information on user prompts that might be encountered.

Other **READ** prompts might include:

- Enter number of samples

- Plate ID

- Enter Sample ID

If a **Manual Map** is used, prompts for information might include:

- Well locations for each sample

ENTER NUMBER OF SAMPLES

Use this screen to enter from 00 to the maximum number of samples permitted by the previously created well map. If there are no controls, standards, or blanks defined, the minimum number of samples is 1.

This value controls the number of samples reported if Matrix or Column reports are requested.

```
ENTER    NUMBER    OF

SAMPLES:                          9 1
```

ENTER PLATE ID

You can enter a 10-character (maximum) identifier to assign to the plate. Since this Plate ID will be stored in the reader's memory, each plate ID should be unique.

(i) **Note:** Use caution when creating multiple Plate IDs. The reader does not warn you that you are about to exceed the maximum of 10 plate IDs stored in memory. If an eleventh Plate ID is added, it will overwrite the first Plate ID stored in memory.

(i) **Note:** If the internal bar code scanner option is installed, the reader will automatically scan the plate/bar code label and use this as the Plate ID.

```
PLATE   ID:

     –          /       :            SPACE
```

- Use the **KEYPAD** to enter numbers, and the **LEFT / RIGHT** arrow keys to move the cursor. **CLEAR** clears the display.

ENTER SAMPLE ID

You can start entering a starting sample identification number from 0001 to 9999. The software will automatically increment each subsequent sample identification number by 1. Sample IDs will be assigned according to the previously defined mapping order.

```
E N T E R

S A M P L E   I D :   0 0 0 1
```

- Use the **KEYPAD** to enter numbers and the **LEFT / RIGHT** arrow keys to move the cursor. **CLEAR** clears the display.

PROMPTS FOR WELL LOCATION

Well locations can be changed at run time if a Manual Map has been specified, and you have requested a sample count at run time via the Utilities menu.

```
S A M P L E #   1           L O C A T I O N

                            R E P #   1 : A 0 1
```

- Use the **KEYBOARD** to enter the well location, using the **SHIFT-LETTER** sequence to key in letters, and press **ENTER** to specify the desired location.

BEGINNING THE PLATE READ

When the following screen appears on the display, the reader is ready to read a plate:

```
P L A C E   P L A T E   I N   C A R R I E R

A N D   P R E S S   < R E A D >   K E Y
```

- Place the plate in the carrier and press the **READ** key to initiate the plate read. After the plate has been read, all requested reports will be generated.

- To halt in read in progress, press the **STOP** key.

Note: If using the incubation option, the reader will wait for the incubator to reach temperature before reading the plate.

Printing Reports and Assay Lists

Reports are automatically generated after a plate has been read (see *OUTPUT Options* and *REPORT Type* in *Chapter 2* for information on selecting reports). To manually regenerate results reports, use the **REPORT** option from the Main Menu. You can also print Map, Assay, and Assay List reports.

Note: See *Appendix D* for sample reports.

```
┌─────────────────────────────────────────────────────┐
│ R E A D Y        9 : 4 5 P M      0 5 / 0 9 / 9 5    │
│                                                       │
│ R E A D    D E F I N E    R E P O R T    U T I L     │
└─────────────────────────────────────────────────────┘
```

- Press **SOFT KEY 3, REPORTS** to access additional reports.

```
┌─────────────────────────────────────────────────────┐
│ P R I N T    R E P O R T :                           │
│                                                       │
│ R E S U L T    M A P    A S S A Y    L I S T         │
└─────────────────────────────────────────────────────┘
```

- Select the **RESULT** option to print an exact copy of results from the plate reading. The form in which the results are presented is determined by the report settings (Matrix, Column, Curve-Fit) specified in the **UTILITIES** menu option.

- Select **MAP** to print a matrix showing the locations of the Blanks, Standards, Controls, and Samples for a selected assay.

- Select **ASSAY** to print a plate map and a listing of all of the assay's settings, such as wavelengths, numbers of well types, formulas, and curve-fit parameters.

- Select **LIST** to print a list of all assays (name and number) currently programmed in the ELx808.

(i)　　**Note:** The reader stores measurement values for the last 8 plates in memory.

Result

```
R E P O R T :  H B S - A G

I D :    0 0 1                    0 7 / 1 7 / 9 5
```

- Use the **OPTION** key to select the appropriate **Plate ID** and **Report**. Note that the **Assay ID** will change if the selected **Plate ID** was read with a different assay. Once you have found the correct **Plate ID**, press **ENTER**.

Editing Standard Outliers

If a standard curve was generated and if **EDIT STANDARD OUTLIERS** was set to **MANUAL** in the assay definition, the option to edit outliers is presented.

```
E D I T    S T D    O U T L I E R S :

  Y E S      N O
```

- Select **NO** to include all standards in the curve-fit calculations.

- Select **YES** to indicate that one or more standard replicates or groups should be temporarily excluded from curve fit-calculations.

  ➢ If **AVERAGE STANDARDS** was set to **NO** in the assay definition, one or more standard *replicates* can be chosen for exclusion.

```
E D I T    S T D 1    R E P 1 ?    Y E S

  Y E S      N O
```

- Select **YES** to exclude the replicate from curve-fit calculations.

- Select **NO** to retain the replicate.

- Press **ENTER** to advance to the next replicate.

> ➢ If **AVERAGE STANDARDS** was set to **YES** in the assay definition, one or more standard *groups* can be chosen for exclusion.

```
E D I T    S T D  1 ;  X ?    Y E S

  Y E S      N O
```

- Select **YES** to exclude the group from curve-fit calculations.

- Select **NO** to retain the group.

- Press **ENTER** to advance to the next group.

(i) **Note:** Each curve-fit type requires a minimum number of standards for curve generation: 4 for 2-P, 4-P, cubic, and cubic-spline, 3 for quadratic, and 2 for linear and point-to-point. Exercise caution when editing outliers. If the assay is left with insufficient standards, the curve fit will fail.

Printing Results

After the assay is selected and standard outliers are edited (if necessary), the results report can be printed.

```
P R I N T    R E S U L T S ?

  Y E S      N O
```

- Ensure that the printer is connected, turned on, and filled with paper.

- Press **YES** to print reports, or **NO** to return to the Main Menu.

Map

- Select **REPORT** at the Main Menu, and then select **MAP.**

```
S E L E C T    A S S A Y    N U M B E R : 0 1

N A M E :    H B S - A G
```

- Use the keyboard to type the assay number, or the **OPTION** key to cycle through the list of available assays. Press **ENTER** to enter the assay and begin printing the map of programmed well locations in the selected geometry.

Assay

- Select **REPORT** at the Main Menu, and then select **ASSAY**.

```
S E L E C T    A S S A Y    N U M B E R : 0 1

N A M E :    H B S - A G
```

- Use the keyboard to type the assay number, or the **OPTION** key to cycle through a list of available assays. Press **ENTER** to enter the assay and begin printing the map and other assay parameters.

List

- Select **REPORT** at the Main Menu, and then select **LIST**. The entire list of assays stored in the ELx808's memory will be sent to the printer.

Chapter 4

Instrument Qualification

This chapter discusses the tasks and procedures necessary for qualifying instrument performance initially and on an ongoing basis. A convenient Recommended Qualification Schedule arranges tasks into Operational and Performance Qualification categories.

This chapter contains the following sections:

Recommendations for Achieving Optimum Performance

- Microplates should be perfectly clean and free of dust or bottom scratches. Use new microplates from sealed packages. Do not allow dust to settle on the surface of the solution; use microplate covers when not reading the plate. Filter solutions to remove particulates that could cause erroneous readings.

- Although the ELx808™ supports standard flat, U-bottom, and V-bottom 96-well microplates, the reader achieves optimum performance with optically clear, flat-bottomed wells.

- Non-uniformity in the optical density of the well bottoms can cause loss of accuracy, especially with U- and V-bottom polyvinyl microplates. Check for this by reading an empty microplate. Dual wavelength readings can eliminate this problem, or bring the variation in density readings within acceptable limits for most measurements.

- Inaccuracy in pipetting has a large effect on measurements, especially if smaller volumes of liquid are used. For best results, use at least 100 μl per well.

- The inclination of the meniscus can cause loss of accuracy in some solutions, especially with small volumes. Agitate the microplate before reading to help bring this problem within acceptable limits. Use Tween® 20, if possible (or some other wetting agent) to normalize the meniscus. Some solutions develop menisci over a period of several minutes. This effect varies with the brand of microplate and the solution composition. As the center of the meniscus drops and shortens the light path, the density readings change. The meniscus shape will stabilize over time.

- Although the effect of ambient light is mathematically quantified and subtracted from each absorbance reading, the illumination of the instrument by strong ambient light should be avoided. If interference from ambient light is suspected, read a microplate of high-density solutions under the suspect conditions, and again with all ambient light blocked (in a dark room, for example), then compare results. The blocked results will appear as an increase in optical density readings if light is influencing the readings. Because of the mathematical correction, this difference under normal conditions should be slight or nonexistent.

- A 10-minute warm-up of the instrument is suggested, prior to reading, to achieve the best repeatability from microplate-to-microplate measurements.

Installation Qualification (IQ)

The **IQ** is conducted when unpacking and setting up the ELx808™ according to the following steps. Refer to *Chapter 2, Installation* where necessary for illustrations and additional information.

1. Unpack the instrument as outlined in *Chapter 2* and ensure that all required items (and any purchased optional items) are present. Check for any possible shipping damage.

2. Record the Instrument Model Number, Serial Number, BioTek Sales Order Number, and the date of shipment from BioTek.

3. Locate the reader on a level, stable surface where ambient temperatures between 18ºC (64ºF) and 40ºC (104ºF) can be maintained. Environmental conditions to avoid include excessive humidity, excessive ambient light, and dust.

4. Install the reader's 6-position filter wheel as described in *Chapter 2*. This involves removing the perimeter screws from the reader's top shroud to access the internal components. Record the filter wavelength values and their locations in the wheel.

5. **ELx808 with the internal power input module:** Check the voltage setting on the power input module located on the right side of the reader. Adjust it if necessary, according to the directions in *Appendix F*.

6. Locate the power inlet on the right side of the reader. Connect the power:

   ELx808 with the internal power input module:

   - Plug the rounded end of the power cord into the power inlet.

   - Plug the 3-prong end of the power cord into an appropriate power receptacle.

   ELx808 with the external power supply:

   - Connect the power cord to the external power supply, and plug the rounded end of the power supply's line cord into the power inlet.

   - Plug the 3-prong end of the power cord into an appropriate power receptacle.

7. If the reader will be run in "standalone" mode (that is, without any controlling software running on a host computer), connect the printer to the reader, using the supplied parallel cable. Turn on the printer.

8. Turn on the reader using the switch located on the right side.

9. Verify that the ELx808's internal filter table matches the physical locations of the filters in the filter wheel (**UTIL|SETUP|FILTER**).

10. Set the current time and date.

11. If a printer is connected, set the data output and reporting options. Select the proper printer type.

12. Set the read time prompts and read speed.

13. Run a System Test and print the results.

14. Run a Checksum Test and record the checksum, part number(s), and version number(s) of the software. (See page 4-9 for instructions.)

15. If the instrument will be controlled via software running on a host PC:

a. Connect the PC to the reader using the serial cable.

b. If you will be using Gen5™, KC4™, or KCjunior™, install the software on the PC.

c. Ensure that the communication parameters match on both devices (e.g., the baud rate may need to be adjusted on the ELx808™).

d. Launch the PC program and ensure communications are established. Run a System Test and print the results.

The Installation Qualification is now complete.

Recommended Qualification Schedule (OQ/PQ)

The schedule shown below defines the factory-recommended intervals for performance testing for a microplate reader used two to five days a week.

Note: The risk factors associated with your tests may require that the Qualification procedures be performed more or less frequently than shown below.

Table 4-1
Recommended Qualification Schedule

	Operational Qualification: Initially and Annually	**Performance Qualification:** Monthly
System Test, p. 4-7	✔	✔
Checksum Test, p. 4-9	✔	✔
Absorbance Plate Test, p. 4-11	✔	✔
Empty Carrier Test, p. 4-18	✔	
Liquid Test 1*, p. 4-22 or **Liquid Test 2*,** p. 4-24	✔	Every three months
Liquid Test 3**, p. 4-27 (optional, for 340 nm)	✔	

* Regarding Liquid Tests 1 and 2:

    If you have an Absorbance Test Plate, run Liquid Test 1.

    If you <u>do not</u> have an Absorbance Test Plate, run Liquid Test 2.

** Liquid Test 3 is optional; it is provided for sites requiring verification at wavelengths lower than those attainable with the Absorbance Test Plate.

Qualification Procedures

Your ELx808™ reader was fully tested at BioTek prior to shipment and should operate properly upon initial setup. The tests outlined in this section may be utilized to confirm initial and ongoing performance of the ELx808.

Note: An instrument qualification package (PN 7340543) for the ELx808 is available for purchase. The package contains thorough procedures for performing Installation Qualification, Operational Qualification, and Performance Qualification (IQ/OQ/PQ) and Preventive Maintenance (PM). Extensive checklists and logbooks are included for recording results. Contact your local dealer for more information.

Set up the reader according to the Installation Qualification instructions on page 4-3 and confirm that it powers up and communicates with any peripherals. After Installation Qualification, conduct the Operational Qualification tests:

- **System Test:** Verifies proper gains, bulb operation, and low electronic noise.

- **Checksum Test:** Compares the basecode software to the internally recorded checksum values to ensure that the code has not been corrupted. Displays software part number and version information.

- **Absorbance Plate Test:** Confirms the optical accuracy/linearity and repeatability, alignment, channel-to-channel variation, and wavelength accuracy of the instrument.

    - **Accuracy** of the optical density (OD) readings: The comparison of the OD readings with those given with the Absorbance Test Plate insert will confirm the accuracy of the optical density values at that wavelength.

    - **Alignment** of the plate carrier and standard microplates is confirmed by the positional accuracy check of wells F01, C12 and H08.

    - **Channel-to-channel variation** can be tested by completing the turnaround test. This tests the reader's ability to read the glass filters accurately in different plate positions.

    - **Linearity** of the OD readings is confirmed by default if the wavelength readings are accurate.

- **Empty Carrier Test:** Confirms the ELx808's read capabilities at the 100% light level.

- **Liquid Tests:** Verify that the reader will perform to specification with liquid samples.

Conduct these tests again monthly/quarterly as outlined in the schedule, as a part of Performance Qualification.

The following pages describe how to run these tests via the reader's keypad, and include basic instructions for running the tests via Gen5™, KCjunior™, and KC4™ software. Refer to Gen5's Help system or to the KC4 or KCjunior User's Guide for more detailed instructions.

Notes: KC4 and KCjunior refer to the Absorbance Plate Test by its *former* name, "Universal Plate Test."

Gen5's Reader Diagnostics Utility (PN 5320201) must be installed for performance of the Absorbance Plate Test in Gen5.

Initiating Tests Via the Utility Option

```
R E A D Y        0 1 : 3 0 P M    1 0 / 0 9 / 0 4
R E A D     D E F I N E    R E P O R T    U T I L
```

- From the Main Menu, select **UTIL**.

```
S E L E C T    U T I L I T Y    O P T I O N ?
T E S T S    S E T U P    O U T P U T    R E A D
```

- Select **TESTS**.

```
S E L E C T    T E S T
S Y S T E M    C H K S U M       C A L P L A T E
```

- Select **SYSTEM** to run the **SYSTEM TEST**.
- Select **CHKSUM** to run the **CHECKSUM TEST**.
- Select **CALPLATE** to run the **ABSORBANCE PLATE TEST**.

  Note: Before running this test, ensure that the Test Plate values are entered. See *Defining the Test Plate Parameters* on page 4-13 for more information.

"SYSTEM" (System Test)

The System Test conducts a series of tests at each wavelength defined in the filter table to confirm adequate light levels, low electronic noise, adequate photodiode sensitivity, overall system cleanliness, and (if equipped) proper function of the incubator. The testing is designed to verify that the ELx808™ will give in-specification performance for each set wavelength over the specified OD range.

The reader automatically runs an internal System Test each time it is powered on. The reader will "beep" repeatedly if the power-on System Test results do not meet the internally coded Failure Mode Effects Analysis (FMEA) criteria established by BioTek. A system test should then be initiated via the keypad, Gen5™, KC4™, or KCjunior™ to try to retrieve an error code from the reader.

To run the System Test using Gen5: Select **System|Diagnostics|Run System Test**. Click the **Help** button for guidance.

To run the System Test using KC4: Select **System|Diagnostics|Run Optics Test**. Click the **Help** button for guidance.

To run the System Test using KCjunior: Select **Utilities|Diagnostics|Reader System Test**. Click the **Help** button for guidance.

To obtain a printout of the System Test values for either periodic testing and documentation or troubleshooting, select **SYSTEM** from the **TESTS** submenu. The results are sent to the printer via the reader's parallel port, and a printout in pass/fail format, similar to *Figure 4-1* (on the following two pages) is produced.

Note: If a System Test shows one or more channels that have air readings falling below 30,000 for filter wavelengths of 405 nm or higher, Liquid Test 1 should be performed. This test is applicable to the eight reading channels only, not the reference channel.

Note: The incubation portion of the System Test is only performed on incubation-equipped instruments.

```
                     Gen5 System Test Report

Reader:                    ELx808  (Serial Number: 118943)
Basecode:                  P/N 7340201  (v3.15)
Date and Time:             11/28/2005 3:06:11 PM
User:                      Administrator
Company:                   BioTek
Comments:                  System Test run during the IQ

Test Results

Operator ID:_____

Notes:_____


03:11PM   02/28/05    SYSTEM SELF TEST

Filter: 405  Gain: 15.06
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       24272 36673 54326 49036 37302 41392 37956 46880 42789
Dark:       1627  1790  1738  1983  1922  1969  2155  1978  2041
Delta:     22645 34883 52588 47053 35380 39423 35801 44902 40748

Filter: 450  Gain:  5.12
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       26520 40514 54985 51166 41113 45699 43595 50055 47401
Dark:        587   644   626   708   688   705   767   707   729
Delta:     25933 39870 54359 50458 40425 44994 42828 49348 46672

Filter: 490  Gain: 16.00
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       29585 40785 53183 50389 42324 46927 45225 51112 48181
Dark:       1806  1981  1927  2187  2121  2172  2369  2182  2247
Delta:     27779 38804 51256 48202 40203 44755 42856 48930 45934

Filter: 620  Gain:  8.26
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       28973 40210 53315 49573 42596 47522 46312 50677 49083
Dark:        922  1012   983  1116  1084  1111  1211  1114  1150
Delta:     28051 39198 52332 48457 41512 46411 45101 49563 47933

Filter: 630  Gain:  5.57
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       28127 42576 54097 52142 44139 48634 47638 52007 50305
Dark:        630   691   672   762   739   758   826   760   784
Delta:     27497 41885 53425 51380 43400 47876 46812 51247 49521

Filter: 340  Gain: 64.00
Channel:     Ref     1     2     3     4     5     6     7     8
Air:       18961 28041 46573 41370 27548 30291 25144 36515 31946
Dark:       7163  7855  7642  8677  8411  8630  9413  8655  8927
Delta:     11798 20186 38931 32693 19137 21661 15731 27860 23019
```

Figure 4-1: *Sample System Test Report (Sheet 1 of 2)*

Note: The format varies depending on the software used.

```
Channel:      Ref      1      2      3      4      5      6      7      8
Noise Max:   3952   4351   4225   4817   4668   4790   5237   4807   4963
Noise Min:   3951   4346   4220   4814   4664   4785   5233   4802   4957
Delta:          1      5      5      3      4      5      4      5      6


03:11PM   02/28/05    INCUBATOR SELF TEST

Temperature Setpoint: 37.0    Current Average: 36.9      A/D Test: PASS

Zone 1: 37.0   Min: 36.9   Max: 37.0     Range: PASS   Thermistor: PASS
Zone 2: 37.0   Min: 37.0   Max: 37.1     Range: PASS   Thermistor: PASS
Zone 3: 37.0   Min: 36.9   Max: 37.0     Range: PASS   Thermistor: PASS
Zone 4: 36.9   Min: 36.9   Max: 37.0     Range: PASS   Thermistor: PASS


AUTOCAL ANALYSIS


Upper Left Corner:  x=5344   y=0
Lower Left Corner:  x=6156   y=0
Lower Right Corner: x=14850  y=0
Upper Right Corner: x=14032  y=0
Delta 1 : 5344-6156=-812
Delta 2 : 14032-14850=-818


SYSTEM TEST PASS
0000


Reviewed/Approved By: _____  Date: _____


For Technical Support

In the U.S.:                      In Europe:
  BioTek Instruments, Inc.          BioTek Instruments GmbH
  Tel: 800 242 4685                 Tel: 49 (0) 7136-9680
  Fax: 802 655 3399                 Fax: 49 (0) 7136-968-111

All Others:
  Tel: 802 655 4040
  Fax: 802 655 3399

email: TAC@biotek.com
Product support center: www.biotek.com/service
```

Figure 4-1: *Sample System Test Report (Sheet 2 of 2)*

Note: The format varies depending on the software used.

"CHKSUM" (Checksum Test)

The Checksum Test compares the basecode software to the internally recorded checksum values to ensure that the programming has not been corrupted. To verify the checksum, revisions, and version of software currently loaded onto your reader, select **CHKSUM** from the **TESTS** submenu. If an error is detected, the display will show an error code.

Note: This test also runs automatically when the reader is turned on.

When the CHKSUM Test is selected, the software versions and the software's checksum appear on the display, as shown in the **example** below:

Software P/N	Software Version
7330203	Version 3.14
Code Checksum:	(6B87)

The second screen shows:

Configuration P/N	Configuration Version
7330203	Version 2.82

"CALPLATE" (Absorbance Plate Test)

The Absorbance Plate Test, also referred to as the Calibration Plate Test, confirms the alignment, accuracy/linearity, and repeatability of the ELx808™.

To initiate this test, select **CALPLATE** from the **TESTS** submenu.

More information on the test is provided in the next section, *Absorbance Plate Test*.

Absorbance Plate Test

(i) **Note:** Gen5™ Software's Reader Diagnostics Utility (PN 5320201) must be installed before you can perform the Absorbance Plate Test in Gen5.

Description

This test uses BioTek's **Absorbance Test Plate** to confirm the Mechanical Alignment, Accuracy and Linearity, Repeatability, and Channel-to-Channel uniformity of the ELx808™. The Absorbance Plate Test compares the reader's optical density measurements and mechanical alignment to NIST-traceable values.

BioTek's **6-Filter** (PN 9000547) or **7-Filter** (PN 7260522) Absorbance Test Plate may be used for the test.

The Absorbance Plate Test confirms the following:

- **Mechanical Alignment:** The Test Plate has several groups of precisely machined holes to confirm the mechanical alignment of different microplate readers. The amount of light that shines through these holes is an indication of how well the reader is aligned. A reading of more than 0.015 OD for any of the designated alignment holes indicates that the light is being "clipped" and the reader may be out of alignment.

- **Accuracy/Linearity:** The Test Plate contains neutral-density glass filters of known OD values at several wavelengths. Actual measurements are compared against the expected values provided in the Test Plate's Standards Certificate. Since there are several filters with differing OD values, the accuracy across a range of ODs can be established. Once it is proven that the reader is accurate at these OD values, the reader is also considered to be linear.

- **Repeatability:** This test ensures the reader meets its repeatability specification by reading each neutral-density filter on the Test Plate twice with the filter in the same location. Note that there may not be a Pass/Fail indication for filter values that are beyond the specified accuracy (and thus repeatability) range of the instrument.

- **Channel-to-Channel Variation:** This test ensures that selected channels read the same value for a filter as their paired channel when the plate is "turned around" 180° in the plate carrier. It is a way to verify that each channel's Delta values are adjusted relative to the signal strength of the reference channel.

Note: If the Absorbance Test Plate is not available, an alternative method that may be used to determine relative accuracy, repeatability, and linearity is **Liquid Test 2**, described on page 4-24.

Peak Wavelength Test

The 7-Filter Absorbance Test Plate contains a glass filter in position C6. This filter is used to check wavelength accuracy in readers with monochromators. A "Peak Wavelength Test" is performed when the glass filter is scanned across a specified wavelength range in 1-nm increments. The wavelength of the maximum absorbance is compared to the expected peak wavelength supplied on the 7-Filter Test Plate's Peak Wavelength Certificate. Since the ELx808 does not have a monochromator, the Peak Wavelength Test will be not be performed.

Requirements

To run the Absorbance Plate Test, you'll need BioTek's **6-Filter** or **7-Filter Absorbance Test Plate** with its accompanying **Standards Certificate**.

The Standards Certificate contains standard OD values for the filters at several different wavelengths (see sample below). These values must be entered via the keypad, or via Gen5™, KC4™, or KCjunior™ Software *before* the Absorbance Plate Test can be performed. This information must only be entered once.

This test plate can be used for testing the reproducibility, linearity, and alignment of your BioTek autoreader. The following calibration data has been recorded by a N.I.S.T. traceable spectrophotometer.

WAVELENGTH (nm)

Well	405nm	450nm	490nm	550nm	620nm	630nm	690nm	750nm
C1	0.147	0.140	0.135	0.130	0.136	0.136	0.127	0.134
E2	0.618	0.575				0.568	0.485	0.434
G3	1.133	1.052	S A M P L E			1.040	0.881	0.783
H6	1.701	1.578				1.560	1.323	1.179
F5	2.279	2.024	1.976	1.956	1.893	1.865	1.537	1.272
D4	2.945	2.604	2.545	2.513	2.437	2.400	1.972	1.632

Set # 2453   Serial # 161259

Figure 4-2: Sample Standards Certificate for the Absorbance Test Plate

The 7-Filter Test Plate also includes a **Peak Wavelength Certificate**. The Peak Wavelength Certificate contains one or more Peak Wavelength values for the glass filter located in position C6 on the plate.

When defining the 7-filter test plate in Gen5, KC4, or KCjunior software, you must also enter values for the "Peak Wavelength Test," even though the ELx808™ does not have a monochromator, and the Peak Wavelength Test will not actually be performed. Refer to the Gen5 Help system or to the KC4 or KCjunior User's Guides for more information on this topic.

Defining the Test Plate Parameters

Using the Standards Certificate provided with the Absorbance Test Plate, follow the steps below to define the Absorbance Test Plate parameters.

To define the Absorbance Test Plate parameters via the instrument keypad:

```
R E A D Y        0 1 : 3 0 P M   1 0 / 0 9 / 0 4

R E A D    D E F I N E   R E P O R T   U T I L
```

- From the Main Menu, select **UTIL|SETUP |*MORE|CAL PLATE**.

- The **CALIBRATION FILTER** selection screen will appear.

```
C A L I B R A T I O N   F I L T E R :   4 0 5

  4 0 5      4 5 0      4 9 0      6 3 0
```

- Select the filter you wish to enter data for.
- A data entry screen will appear, requiring the entry of a Well Location and corresponding Calibration Value. Carefully enter the information from the Standards Certificate.

```
W A V E L E N G T H : 4 0 5   W E L L : C 0 1

C A L I B R A T I O N   V A L U E S : 0 . 0 0 0
```

- After each entry, press **ENTER** to advance to the next well location. Continue to enter the values listed on the Certificate for each well location at each filter (wavelength) value you wish to test.

- When all values have been entered, press the **MAIN MENU** key.

ⓘ **Gen5 customers:** The Reader Diagnostics Utility must be installed before you can perform the Absorbance Plate Test.

To define the Absorbance Test Plate parameters in Gen5™: Select **System|Diagnostics|Test Plates| Add/Modify Plates**, and click **Add**. Click the **Help** button for guidance.

To define the Absorbance Test Plate parameters in KC4™: Select **System|Diagnostics|Define Universal Plates|Add**. Click the **Help** button for guidance.

To define the Absorbance Test Plate parameters in KCjunior™: Select **Utilities|Diagnostics| Universal Plate Test|New Data Sheet**. Click the **Help** button for guidance.

Running the Absorbance Plate Test

To run the Absorbance Plate Test from the instrument keypad, at the Main Menu select **UTIL| TESTS|CALPLTE**.

```
S E L E C T     T E S T ?

S Y S T E M     C H K S U M           C A L P L T E
```

```
C A L I B R A T I O N     F I L T E R :     4 0 5

  4 0 5         4 5 0         4 9 0         6 3 0
```

- The test is run using one filter wavelength. Select the desired wavelength from the **CALIBRATION FILTER** screen.

- When prompted, insert the Test Plate into the ELx808's plate carrier, and press the **READ** key to begin the calibration program.

- The plate will be read twice and the reader will prompt you to rotate the plate 180°. Rotate the plate so that H12 is in the A1 position, and then press the **READ** key to complete the test.

- The Absorbance Plate Test Report (*Figure 4-3* on the next page) will print when the test is complete.

To run the Absorbance Plate Test in Gen5™, select **System|Diagnostics|Test Plates|Run**. Click the **Help** button for guidance.

To run the Absorbance Plate Test in KC4™, select **System|Diagnostics|Run Universal Plate Test**. Click the **Help** button for guidance.

To run the Absorbance Plate Test in KCjunior™, select **Utilities|Diagnostics|Universal Plate Test**. Click the **Help** button for guidance.

(i) **Important!** The ELx808's Absorbance Plate Test tests the accuracy and repeatability specifications from *0.000 to 2.500 OD only*. The Absorbance Plate Test report (*Figure 4-3* on the following page) displays the OD value read in well position D04, but does not indicate PASS or FAIL, because the value is higher than 2.500 OD and therefore is not within the software test range.

(i) **Important!** The Absorbance Plate Test and CALPLATE utility in the reader are used only to verify the reader's performance. The Test Plate and utility DO NOT correct or calibrate the instrument.

```
CALIBRATION PLATE ANALYSIS

08:51AM 06/21/02   Read Mode:Normal   Filter: 405

Operator ID:_____

Notes:_____  _____

_____


A1 Top Left
Alignment Results
  F01=0.002  PASS     C12=0.002  PASS     H08=0.003  PASS

A1 Top Left
Accuracy Results
             C01      D04      E02      F05      G03      H06
   STANDARD  0.156    2.812    0.620    2.264    1.192    1.754
   DATA      0.147    2.824    0.618    2.276    1.197    1.757
   RESULT    PASS              PASS     PASS     PASS     PASS

Repeatability Results
             C01      D04      E02      F05      G03      H06
   READ 1    0.147    2.824    0.618    2.276    1.197    1.757
   READ 2    0.147    2.817    0.618    2.275    1.197    1.757
   RESULT    PASS              PASS     PASS     PASS     PASS

H12 Top Left
Accuracy Results
             C01      D04      E02      F05      G03      H06
   STANDARD  0.156    2.812    0.620    2.264    1.192    1.754
   DATA      0.146    2.812    0.618    2.274    1.196    1.756
   RESULT    PASS              PASS     PASS     PASS     PASS

Repeatability Results
             C01      D04      E02      F05      G03      H06
   READ 1    0.146    2.812    0.618    2.274    1.196    1.756
   READ 2    0.146    2.815    0.618    2.275    1.196    1.756
   RESULT    PASS              PASS     PASS     PASS     PASS
```

Figure 4-3: *Sample Absorbance Plate Test Report*

Note: The format varies depending on the software used.

Results & Troubleshooting Tips

The Absorbance Plate Test Report contains results for the following:

- **Alignment:** This portion of the test measures the alignment of the microplate carrier with the optical path. A reading greater than 0.015 OD for alignment wells F01, C12, or H08 represents an out-of-alignment condition. If the test fails:

  - ➢ Examine the microplate carrier to ensure that it is clear of debris.

  - ➢ Make sure the Test Plate is properly seated in the microplate carrier.

  - ➢ Check the Test Plate's alignment holes to ensure they are clear of debris.

- **Accuracy:** Accuracy is a measure of the absorbance (optical density) of Test Plate wells C01, D04, E02, F05, G03 and H06 compared to known standard values contained in the Test Plate Standards Certificate. If the test fails:

  - ➢ Examine the glass filters on the test plate to see if they are dirty. If needed, clean them with lens paper. **Note:** Do not remove the filters from the test plate, and do not use alcohol or other cleaning agents to clean them.

  - ➢ Check the filter calibration values entered in the ELx808™ software to make sure they match the values on the Test Plate Standards Certificate.

  - ➢ Check the Calibration Due Date on the Test Plate; if it has expired, the plate must be re-calibrated. Contact BioTek.

- **Repeatability:** This test ensures that the reader meets its repeatability specification by reading each Test Plate filter twice with the filter in the same location. Note that there may not be a Pass/Fail indication for filter values that are beyond the specified accuracy (and thus repeatability) range of the instrument. If the test fails:

  - ➢ Examine the glass filters on the test plate to see if there are any loose particles that may have shifted between readings and caused changes. If needed, clean the filters with lens paper. **Note:** Do not remove the filters from the test plate, and do not use alcohol or other cleaning agents to clean them.

  - ➢ Check the microplate carrier to ensure that it is clear of debris.

- **Regarding Linearity:** Linearity of the Optical Density readings is confirmed by default if the readings are accurate (that is, if the Accuracy portion of this test passed). To further prove linearity, you can perform a regression analysis on the Test Plate OD values in a program such as Microsoft® Excel as follows:

  1. Launch Excel.

  2. Open a spreadsheet and label one column "Assigned" and the next column "Observed".

  3. Enter the Assigned OD data for each glass filter in the first column from the Standards Certificate provided with the Test Plate. (Analyze one wavelength at a time.)

  4. Enter the Observed OD values for the same glass filters in the adjacent column.

5. Under Tools, select Data Analysis and then Regression. Use the Regression "Input" box to enter the Assigned values as the "Input Y Range" and the Observed OD as the "Input X Range".

   Note: If the Data Analysis command is not available on the Tools menu, you may need to install the Analysis ToolPak in Microsoft® Excel. Consult Microsoft® Excel Help for assistance.

6. Click **OK** and the Summary Output sheet will be displayed. An R Square value of at least 0.99 is expected.

If any portion of the test continues to fail, contact BioTek's Technical Assistance Center (refer to *Chapter 1* for contact information). Please have a copy of the test report and the reader's serial number available when you call.

Empty Carrier Test

The Empty Carrier Test confirms the ELx808's read capabilities at the 100% light level, and can help to identify deteriorating interference wavelength filters and other optical problems.

Perform these steps (do *not* place a microplate on the carrier):

1 For each filter in the filter wheel, run a simple endpoint protocol and then print the results. (Do not apply blanking or transformations.)

2 Examine the OD values on the printouts. Every OD should read 0.000 ± 0.004 OD.

Note: These limits are required by BioTek and cannot be changed.

3 Sign and date the printouts, and store them with your test documentation.

If the Empty Carrier test fails <u>at more than one wavelength</u>:

- Check the microplate carrier to ensure that it is clear of debris.

- Verify that the lamp is properly aligned, and then run the test again. If the lamp has been in use for approximately 600 hours, it may need to be replaced, and the contacts in the lamp socket may need to be cleaned. Refer to ***Replace the Lamp and Clean the Contacts*** in ***Chapter 5***, ***Preventive Maintenance***.

- Follow the steps in ***Inspect and Clean the Wavelength Filters*** in ***Chapter 5*** to remove the filter wheel and clean the filters.

If the Empty Carrier test fails <u>at just one wavelength</u>:

- Remove the filter wheel and examine the filter that exhibited the problem; it may need to be replaced (contact BioTek). Check for spotting or a halo effect.

Liquid Testing

Conducting "Liquid Tests" confirms the ELx808's ability to perform to specification with liquid samples. Liquid testing differs from testing with the Absorbance Test Plate in that liquid in the wells has a meniscus, while the Test Plate's neutral density filters do not. The optics characteristics may differ in these two cases, alerting you to different types of problems. The Absorbance Test Plate will indicate the absolute amount of light absorbed, which will effectively test the linearity of the electronics. The liquid tests will help detect optical defects such as dirt on the lenses or other contamination that can contribute to errant readings.

- **Liquid Test 1** tests the alignment, accuracy, repeatability, and channel-to-channel variability of the reader, making evident any problems with the optics of the system. If you have the Absorbance Test Plate, you will only need to run Liquid Test 1 for routine testing.

- If you do not have an Absorbance Test Plate, test the linearity, repeatability, and alignment of the reader by preparing a series of solutions of varying absorbances as described in **Liquid Test 2**.

- **Liquid Test 3** is an optional test offered for sites that must have proof of linearity at wavelengths lower than those attainable with the Absorbance Test Plate. This test is considered optional because the reader has good "front end" linearity throughout its wavelength range.

BioTek offers a dye solution (PN 7120779, 25 ml; or 7120782, 125 ml) that may be used in the stock solution formulation for Liquid Tests 1 and 2, or, if you prefer, you may use the dye solution described in *Table 4-3* on the following page. The purpose of the formulation is to create a solution that absorbs light at ~ 2.000 OD full strength when dispensed at 200 μl/well in a flat-bottom microplate.

Alternatively, any solution that gives a stable color will suffice. (This includes substrates incubated with an enzyme preparation and then stopped with an acidic or basic solution.) Some enzyme/substrate combinations that may be used as alternates to the described dye are shown in the table below:

Table 4-2

Typical Enzyme-Substrate Combinations and Stopping Solutions

Enzyme	Substrate	Stopping Solution
Alkaline Phosphate	o-nitrophenyl phosphate	3N sodium hydroxide
beta-Galactosidase	o-nitrophenyl -beta-D galactopyranoside	1M sodium carbonate
Peroxidase	2,2'-Azino di-ethylbenzothiazoline-sulfonic acid (ABTS)	citrate-phosphate buffer, pH 2.8
Peroxidase	o-phenylenediamine	0.03N sulfuric acid

Stock Solution Formulation

The stock solution for Liquid Test #1 and #2 may be formulated from the ingredients listed below (**Solution A**), or by diluting a dye solution available from BioTek (**Solution B**).

SOLUTION A

Required Materials:

- FD&C Yellow No. 5 dye powder (typically 90% pure)
- Tween® 20 (polyoxyethylene (20) sorbitan monolaurate), **or** BioTek wetting agent, PN 7773002
- Deionized water
- Precision balance with readability of 0.001 g
- Weigh boat
- 1-liter volumetric flask

Table 4-3

Stock Solution Formulation for Liquid Tests 1 and 2

FD&C Yellow No. 5 powder	0.092 g
Tween® 20	0.5 ml
DI Water to bring volume to:	1000 ml

Preparation of Stock Solution:

1. Weigh out 0.092 gram of FD&C No. 5 yellow dye powder into a weigh boat.
2. Rinse the contents into a 1-liter volumetric flask.
3. Add 0.5 ml of Tween 20, or 5 ml of BioTek's wetting agent.
4. Make up to 1 liter with DI water; cap and shake well.

   ➢ This should create a solution with an absorbance of about 2.000 when using 200 µl in a flat-bottom microwell. The OD value will be proportional to the volume in the well and the amount of FD&C No. 5 dye used. You can use a larger or smaller well volume, or add more dye or water to adjust the solution. Note that too small a well volume may result in increased pipetting-related errors.

SOLUTION B

Required Materials:

- BioTek QC Check Solution No. 1 (PN 7120779, 25 ml; 7120782, 125 ml)
- Deionized water
- 5-ml Class A Volumetric Pipette
- 100-ml Volumetric Flask

Preparation of Stock Solution:

1. Pipette a 5-ml aliquot of BioTek QC Check Solution No. 1 into a 100-ml volumetric flask.

2. Make up to 100 ml with DI water; cap and shake well.

   ➢ This should create a solution with an absorbance of about 2.000 when using 200 µl in a flat-bottom microwell. The OD value result will be proportional to the volume in the well and the amount of QC Check Solution No. 1 used. You can use a larger or smaller well volume, or add more Check Solution or water to adjust the stock solution. Note that too small a well volume may result in increased pipetting-related errors.

Liquid Test 1

Note: A 96-well, flat-bottom microplate is required for this test (Corning® Costar® #3590 is recommended). Use a new microplate; any fingerprints or scratches may cause variations in the readings.

(i) **Important!** Before running the liquid tests, ensure that the reader is *not* running in Rapid mode. To check this, start at the main menu and select **UTIL > READ** and cycle through the prompts until **READ IN RAPID MODE?** is displayed. Set to **NO**.

---

1. Using a freshly prepared stock solution A or B (see page 4-20 or 4-21), prepare a 1:2 dilution using deionized water (one part stock, one part deionized water; the resulting solution is a 1:2 dilution).

   ➢ The concentrated stock solution should have an optical density of approximately 2.000 OD or lower.

2. Pipette 200 μl of the *concentrated* solution into the first column of wells of the microplate.

3. Pipette 200 μl of the *diluted* solution into the second column of wells.

(i) **Note:** After pipetting the diluted test solution into the microplate and *before* reading the plate, we strongly recommend shaking the plate at Variable speed for four minutes. This will allow any air bubbles in the solution to settle and the meniscus to stabilize. Alternatively, wait 20 minutes after pipetting the diluted test solution before reading the plate.

---

4. Read the microplate <u>five times</u> at 405 nm using Normal mode, single wavelength, no blanking ("Normal" plate position).

5. Without delay, rotate the microplate 180° so that well A1 is now in the H12 position. Read the plate five more times ("Turnaround" plate position).

6. Print the ten sets of raw data, or export the data to an Excel spreadsheet using Gen5™, KC4™, or KCjunior™. The mathematical computations described below may then be performed and the template kept for future data reduction.

CALCULATIONS:

7. Calculate the mean OD value for each well location in columns 1 and 2 for the five plates read in the Normal position, and then again for the five plates read in the Turnaround position. This will result in 32 mean values.

8. Perform a mathematical comparison of the mean values for each microwell in its Normal and Turnaround positions (that is, compare A1 to H12, B1 to G12, … H1 to A12, and so on). To pass this test, the differences in the compared mean values must be within the accuracy specification for the instrument.

    For example:

    If the mean value for well A1 in the Normal position is 1.902, where the specified accuracy is ± 1.0% ± 0.010 OD, then the expected range for the mean of the same well in its Turnaround (H12) position is 1.873 to 1.931 OD.

    1.902 * 0.010 + 0.010 = 0.029; 1.902 - 0.029 = **1.873**; 1.902 + 0.029 = **1.931**

    Note: If any set of mean values is out of the expected range, review the other three sets of mean values for the same channel pair. For example, if the A1/H12 comparison fails (the wells are not within the expected range of each other), review the comparisons of A2/H11, H1/A12, and H2/A11. If two or more sets of mean values for a channel pair are out of the expected range, there is a problem with one of the eight read channels. If only one of the four mean values results in a failure, check the well for debris and the plate for scratches or fingerprints.

ACCURACY SPECIFICATION:

For comparison in this test, the following accuracy specification is applied, using Normal mode and a 96-well microplate.

± 1.0% ± 0.010 OD from 0.000 to 2.500 OD @405 nm

Liquid Test 2

Note: A 96-well, flat-bottom microplate is required for this test (Corning® Costar® #3590 is recommended). Use a new microplate; any fingerprints or scratches may cause variations in the turnaround reading.

REQUIRED MATERIALS

- A new 96-well, flat-bottom microplate (Corning® Costar® #3590 is recommended)
- Ten test tubes, numbered consecutively, stored in a rack
- Calibrated hand pipette (Class A volumetric pipette recommended)
- Stock solution A or B (see page 4-20 or 4-21)

PREPARATION OF DILUTIONS:

Create a percentage dilution series, beginning with 100% of the original concentrated stock solution (A or B) in the first tube, 90% of the original solution in the second tube, 80% in the third tube, all the way to 10% in the last tube. Dilute using amounts of the remaining 0.05% solution of deionized water and Tween 20, as shown below:

Table 4-4

Test Tube Dilutions

Tube Number	1	2	3	4	5	6	7	8	9	10
Volume of Original Solution (ml)	20	18	16	14	12	10	8	6	4	2
Volume of 0.05% Tween Solution (ml)	0	2	4	6	8	10	12	14	16	18
Absorbance expected if original solution is 2.0 at 200 μl	2.0	1.8	1.6	1.4	1.2	1.0	0.8	0.6	0.4	0.2

Note: The choice of dilutions and the absorbance of the original solution can be varied. Use **Table 4-4** as a model for calculating the expected absorbances of a series of dilutions, given a different absorbance of the original solution.

PLATE PREPARATION:

1. Pipette 200 μl of the concentrated solution from Tube 1 into each well of the first column, A1 to H1, of the microplate.

2. Pipette 200 μl from each of the remaining tubes into the wells of the corresponding column of the microplate (Tube 2 into wells A2 to H2, etc.).

Note: After pipetting the diluted test solution into the microplate and *before* reading the plate, we strongly recommend shaking the plate at Variable speed for four minutes. This will allow any air bubbles in the solution to settle and the meniscus to stabilize. Alternatively, wait 20 minutes after pipetting the diluted test solution before reading the plate.

LINEARITY – TEST A

1. Read the microplate prepared on the previous page <u>five times</u> using Normal mode, dual wavelength at 450 nm with 630 nm as the blank. **Note:** Do not discard the plate; you will use it again for Test C.

2. Print the ten sets of raw data or export it to an Excel spreadsheet using Gen5™, KC4™, or KCjunior™. The mathematical computations described below may then be performed and the template kept for future data reduction.

Calculations:

3. Calculate the mean absorbance for each well, and then average the means for each concentration.

4. Perform a regression analysis on the data to determine if there is adequate linearity.

   For example, using Microsoft® Excel:

   - In a spreadsheet, create two columns labeled 'X' and 'Y'. Enter the actual absorbance values in column X. Enter the expected absorbance values in column Y.

   - Select **Tools|Data Analysis|Regression**. Identify column X as the 'Input X Range' and column Y as the 'Input Y Range' and then click OK to perform the analysis.

   - Click **OK** to perform the analysis, the results of which will be output in a separate sheet.

   - **Note:** If the Data Analysis command is not available on the Tools menu, you may need to install the Analysis ToolPak in Excel. Consult Excel's help system for assistance.

Expected Results:

Since it is somewhat difficult to achieve high pipetting accuracy when conducting linear dilutions, an R Square value of at least 0.99 is considered adequate.

REPEATABILITY – TEST B

1. Calculate the mean and standard deviation for the five readings taken above at each concentration. Only one data set needs to be analyzed. The well that shows the most variation for any concentration is selected for data reduction.

2. For each mean below 2.500 OD, calculate the allowed deviation using the repeatability specification for a 96-well plate of ± 0.5% ± 0.005 OD from 0.000 to 2.500 @405 nm.

3. The standard deviation for each set of readings should be less than the allowed deviation.

For example: Absorbance readings of 1.950, 1.948, 1.955, 1.952, and 1.950 will result in a mean of 1.951, and a standard deviation of 0.0026. The mean (1.951) multiplied by 0.5% (1.951 * 0.005) = 0.0098, which, when added to the 0.005 (0.0098 + 0.005) = 0.0148 OD, which is the allowable deviation. Since the standard deviation is less than this value, the reader meets the test criteria.

Repeatability Specification:

± 0.5% ± 0.005 OD from 0 to 2.500 OD @405 nm

CHANNEL-TO-CHANNEL VARIATION AND ALIGNMENT – TEST C

1. Using the plate prepared for Test A on the previous page, conduct a Turnaround test by reading the plate with the A1 well in the H12 position <u>five times</u>. This test results in 2 comparisons of each channel to its corresponding channel.

2. Calculate the means of the wells in column 1 in the Normal plate position (data is from Test A) and in the turnaround position (from Step 1 above). Compare the mean reading for well A1 to its mean reading when in the H12 position. Next, compare the mean values for the other wells to their corresponding mean values with the well in the turnaround position. (Compare B1 to G12, C1 to F12, D1 to E12, E1 to D12, F1 to C12, G1 to B12, H1 to A12). The difference in the values for any two corresponding wells should be within the accuracy specification for the instrument.

For example:

If the mean of well A1 in the normal position is 1.902, where the specified accuracy is ± 1.0% ± 0.010 OD, then the expected range for the mean of the same well in the H12 position is 1.873 to 1.931 OD. (1.902 * 1.0% = 0.019 + 0.010 = 0.029, which is added and subtracted from 1.902 for the range.)

If any set of well values is out of the expected range, review the other set for the same channel pair. Thus, if A1 and H12 are not within range of each other, review the compliance of H1 to A12, A2 to H11, and H2 to A11. This will confirm that there is a problem in one of the eight read channels, or indicate that the result of one set of wells was in error. If any two sets of well values for a channel pair are out of the allowed accuracy range, there may be contamination on one of the lenses.

3. If the four corner wells are within the repeatability range, the reader is also in alignment.

Liquid Test 3

Note: A 96-well, flat-bottom microplate is required for this test (Corning® Costar® #3590 is recommended). Use a new microplate; any fingerprints or scratches may cause variations in the turnaround reading.

REQUIRED MATERIALS:

- 340 nm filter installed in the reader
- New 96-well, flat-bottom microplate (Corning® Costar® #3590 recommended)
- Calibrated hand pipette(s)
- Beakers and graduated cylinder
- Precision balance with readability of 0.01 g
- Buffer Solution A or B

SOLUTION A: 10X CONCENTRATE PBS

- Deionized water
- Ingredients shown in *Table 4-5*
- β-NADH Powder (β Nicotinamide Adenine Dinucleotide, Reduced Form) Sigma® bulk catalog number N 8129, or pre-weighed 10-mg vials, Sigma® number 340-110

  Note: Store the β-NADH powder according to the guidelines on its packaging

1. Prepare the stock buffer solution using the ingredients below:

Table 4-5

Phosphate Buffered Saline 10X Concentrate Solution

KH_2PO_4 anhydrous	0.2 grams
NaCl	8.0 grams
Na_2HPO4 anhydrous	1.15 grams
KCl	0.2 grams
Tween® 20	0.5 ml
Add Deionized water to bring to	100 ml

2. Mix 5 ml of the stock buffer solution with 45 ml of deionized water.
3. Add 10 mg of the β-NADH powder and mix thoroughly. This is the **10x Concentrate PBS Solution**.

SOLUTION B: SIGMA PBS

- Deionized water

- Phosphate-Buffered Saline (PBS), pH 7.2-7.6, Sigma® tablets, #P4417
  (or equivalent)

- β-NADH Powder (β Nicotinamide Adenine Dinucleotide, Reduced Form) Sigma® bulk
  catalog number N 8129, or pre-weighed 10-mg vials, Sigma® number 340-110

  Note: Store the β-NADH powder according to the guidelines on its packaging

1. Prepare a PBS solution from the Sigma tablets.

2. In a beaker, mix 50 ml of the PBS solution with 10 mg of the β-NADH powder and mix
   thoroughly. This is the **Sigma PBS Solution**.

PROCEDURE:

1. Check the absorbance of a sample of either buffer solution at 340 nm on the microplate
   reader. This solution, which will be referred to as the **100% Test Solution**, will have an
   optical density (absorbance) of approximately 0.700 to 1.000. This value is not critical,
   but it should be within this absorbance range.

   If low, adjust up by adding β-NADH powder until the high-level test solution is at least at
   the lower end of this range. Do not adjust if slightly high.

2. Carefully prepare a **75% Test Solution** by diluting 15 ml of the 100% Test Solution:

   - If using the <u>Sigma PBS solution</u>, use 5 ml as the diluent.

   - If using the <u>10x Concentrate PBS Solution</u>, mix one part of the concentrate with
     nine parts of deionized water. Then use 5 ml of this solution as the diluent.

3. Carefully prepare **50% Test Solution** by diluting 10 ml of the 100% Test Solution:

   - If using the <u>Sigma PBS solution</u>, use 10 ml as the diluent.

   - If using the <u>10x Concentrate PBS Solution</u>, mix one part of the concentrate with
     nine parts of deionized water. Then use 10 ml of this solution as the diluent.

4. Pipette the three solutions into the **new** 96-well microplate:

   - 150 μl of the <u>100% Test Solution</u> into all wells of columns 1 and 2

   - 150 μl of the <u>75% Test Solution</u> into all wells of columns 3 and 4

   - 150 μl of the <u>50% Test Solution</u> into all wells of columns 5 and 6

Note: After pipetting the diluted test solution into the microplate and *before*
reading the plate, we strongly recommend shaking the plate at Variable speed
for four minutes. This will allow any air bubbles in the solution to settle and
the meniscus to stabilize. Alternatively, wait 20 minutes after pipetting the
diluted test solution before reading the plate.

---

5. Read the microplate five times using Normal mode, single wavelength at 340 nm, no blanking (or blank on air).

6. Print the five sets of data, or export the data to an Excel spreadsheet using Gen5™, KCjunior™, or KC4™. The mathematical computations described below may then be performed and the template kept for future data reduction.

REPEATABILITY – TEST A

1. For each well, calculate the mean and standard deviation of the five readings. Only one data set needs to be analyzed for each concentration. The well that shows the most variation for each concentration is selected for data reduction.

2. For each mean calculated in step 1, calculate the allowed deviation using the repeatability specification for a 96-well plate of ± 1.0% ± 0.005 OD from 0.000 to 2.000 OD @340 nm.

3. For each well, compare the standard deviation calculation in step 1 with the allowed deviation calculation in step 2. The standard deviation should be less than the allowed deviation.

   For example:

   Five readings in well A1 of 0.802, 0.802, 0.799, 0.798, and 0.801 result in a mean of 0.8004 and a standard deviation of 0.0018. To calculate the allowed deviation for well A1, multiply the mean by 1.0% and add 0.005 (0.8004 * 0.010 + 0.005) to get 0.013. Since the standard deviation for well A1 is less than 0.013, the well meets the test criteria.

Repeatability Specification

± 1.0% ± 0.005 OD from 0.000 to 2.000 OD @340 nm

LINEARITY – TEST B

1. For each of the three Test Solutions, calculate the mean absorbance for the wells containing that solution (mean of wells A1 to H2, A3 to H4, and A5 to H6).

2. Perform a regression analysis on the data to determine if there is adequate linearity.

   Example using Microsoft® Excel:

   In a spreadsheet, enter the three mean values in ascending order and label the column as the Y values. Enter 0.50, 0.75, and 1.00 and label the column as the X values.

   Select Tools > Data Analysis > Regression. Identify column Y as the "Input Y Range" and column X as the "Input X Range" and then click OK to perform the analysis.

   Note: If the Data Analysis command is not available on the Tools menu, you may need to install the Analysis ToolPak in Microsoft® Excel. Consult Microsoft® Excel Help for assistance.

Expected Results:

Since it is somewhat difficult to achieve high pipetting accuracy when conducting linear dilutions, an R Square value of at least 0.99 is considered adequate.

Chapter 5

Preventive Maintenance

This chapter provides step-by-step instructions for maintaining the ELx808™ in top condition, to ensure that the reader continues to perform to specification.

This chapter contains the following sections:

Recommended Maintenance Schedule

Overview

A general **Preventive Maintenance** (**PM**) regimen for all ELx808™ models includes periodically cleaning all exposed surfaces, inspecting/cleaning the wavelength filters, replacing the lamp, and decontamination. Robotic models should have certain components lubricated.

The following chart recommends Preventive Maintenance tasks and the frequency with which each task should be performed..

Note: The risk factors associated with your tests may require that some or all of the procedures be performed more or less frequently than shown below.

Table 5-1
Recommended Maintenance Schedule

Task	Frequency			
	Every three months	Every six months	As needed	Before storage or shipment
Clean Exposed Surfaces, p. 5-4			✔	
Inspect/Clean Wavelength Filters, p. 5-5	✔			
(Optional) Lubricate Robotic Components, p. 5-6		✔		
Replace Lamp and Clean Contacts, p. 5-8			✔ (after ~ 500 hours)	
Decontamination (see *Appendix A)*				✔

Warnings and Precautions

Please read the following before performing any maintenance procedures:

Important! Turn off and unplug the instrument for all maintenance and repair operations.

Wear protective gloves when handling contaminated instruments. Gloved hands should be considered contaminated at all times; keep gloved hands away from eyes, mouth, nose, and ears.

Mucous membranes are considered prime entry routes for infectious agents. Wear eye protection and a surgical mask when there is a possibility of aerosol contamination. Intact skin is generally considered an effective barrier against infectious organisms; however, small abrasions and cuts may not always be visible. Wear protective gloves when handling contaminated instruments.

Do not immerse the instrument, spray it with liquid, or use a "wet" cloth. Do not allow the cleaning solution to run into the interior of the instrument. If this happens, contact the BioTek Service Department.

Do not soak the keypad—this will cause damage. Moisten a clean cloth with deionized or distilled water and wipe the keypad. Dry immediately with a clean, dry cloth.

Clean Exposed Surfaces

Exposed surfaces may be cleaned (not decontaminated) with a cloth moistened (not soaked) with water or water and a mild detergent. You will need:

- Mild detergent

- Deionized or distilled water

- Clean cotton cloths

To clean the exposed surfaces:

1. Turn off and unplug the instrument.

2. Moisten a clean cotton cloth with water, or with water and the mild detergent. Do not soak the cloth.

3. Wipe the plate carrier, the inside of the plate carrier door, and all exposed surfaces of the instrument.

4. If detergent was used, wipe all surfaces with a cloth moistened with water.

5. Use a clean, dry cloth to dry all wet surfaces.

Inspect and Clean the Wavelength Filters

Laboratory air is used to cool the lamp, and the wavelength filters can become dusty as a result. The filters must be inspected and cleaned at least every three months. You will need:

- Tape

- Flat-blade screwdriver

- Isopropyl, ethyl, or methyl alcohol

- Lens-cleaning tissue

To inspect and clean the wavelength filters:

1. If you have not already done so, turn off and unplug the reader.

2. Temporarily tape the reader's plate access door shut.

3. Using your fingertips, locate the seven perimeter screws around the sides and front where the reader's top shroud meets the base. Remove the screws using a flat-blade screwdriver. **Tip:** Bring the reader to the edge of the work surface to access the screws without having to turn the reader upside down.

4. Carefully raise the front of the shroud and tip it toward the back of the instrument.

5. The filter wheel is located in the left rear quadrant inside the reader. Remove the filter wheel by pulling it off its magnetic hub.

6. Inspect the glass filters for speckled surfaces or a halo effect. This may indicate deterioration due to moisture exposure over a long period of time. If you have any concerns about the quality of the filters, contact your BioTek representative.

7. Clean the filters using lens-cleaning tissue moistened with a small amount of isopropyl, ethyl, or methyl alcohol. Ensure that the filters remain in their current locations.

8. Replace the filter wheel on its magnetic hub. Ensure that the peg on the wheel lines up with the notch on the hub, and that the filter wheel is positioned flat against the hub and rotates freely.

9. Close the top shroud and replace the seven perimeter screws.

Notes:

- Store filters in a dry environment, such as a desiccator, if they will not be used for an extended period.

- If the reader continues to fail tests in *Chapter 4*, *Instrument Qualification*, after you have cleaned the filters and the lamp contacts, or if one or more filters appear to have deteriorated, contact your BioTek Representative or BioTek.

(Optional) Lubricate Robotic Components

This section applies to ELx808 robotic models only. You will need lubricant, BioTek PN 66039.

1. Remove the top shroud of the instrument (see instructions in *Chapter 2*).

2. Lubricate the components as shown in *Figures 5-1* and *5-2* every six months, or after 10,000 cycles, according to the instructions in the table below.

Table 5-2
Instructions for Lubrication of ELx808™ Robotic Models

Component Location	Procedure
A	Lubricate the underside of the bracket.
B	Lightly lubricate the shaft indicated.
C	Apply a heavy coating of lubricant to both sides of the motor shaft. Run the motor shaft back and forth through the motor to ensure that the internal drive nut is heavily lubricated. Use the robotic motor adjustment procedure described on the next page to work the lubricant into the internal drive nut.
D	Lightly lubricate the surface of the roller.
E	Lightly lubricate the underside of the hook-in bracket.

3. Reattach the top shroud.
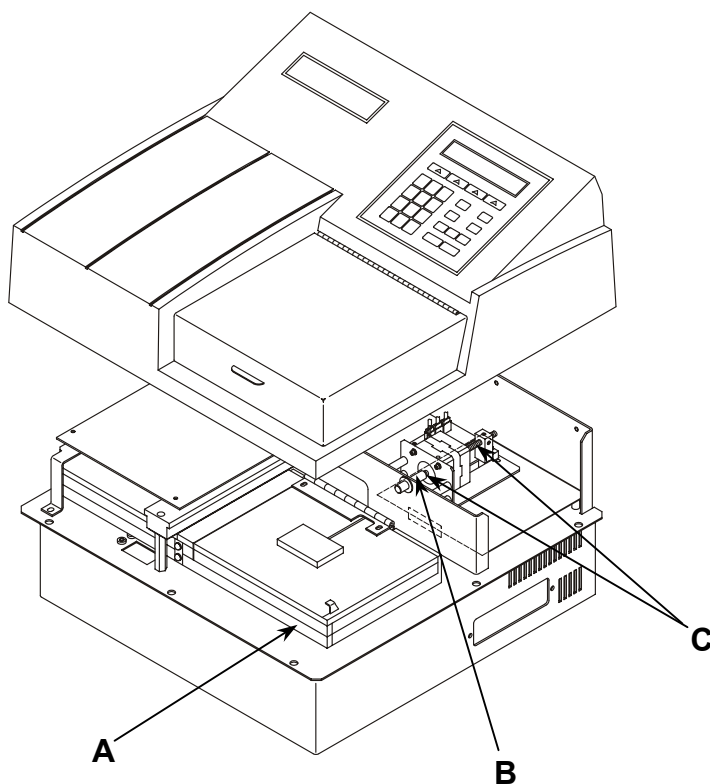
Figure 5-1: *Location of bracket, motor shaft, and main PCB/motor assembly*
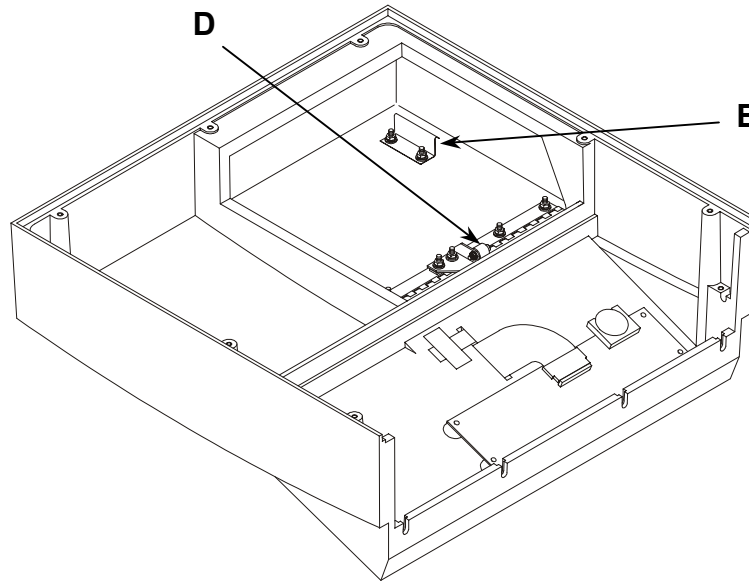
Figure 5-2: *Roller surface and hook-in bracket*

Robotic Motor Adjustment Procedure

After lubrication/reassembly, the robotic door needs to be set to the open and closed positions. Press the following keys to adjust the door.

At the **Main Menu**:

- Press the **UTIL** soft key.

- Press the **SETUP** soft key.

- Press the **hidden key** between the **Main Menu** and **Previous Screen** keys.

You should see the **Door Adjustment** screen.

- Press the **CLOSE** soft key.

- Press the **UP** soft key until you just see the door move up. You may have to press the key many times before the door moves. When the door does move, press the **DOWN** soft key 4 times. This positions the door lift ram just behind the door roller in the closed position.

- Press the **OPEN** soft key.

- Press the **UP** soft key to raise the door to the desired height. Press the **DOWN** soft key if you want to adjust the door downward.

  Important: Do not open the door to its maximum open position.

- Press the **Main Menu** key.

- Press the **CLOSE** soft key. This will close the door. The door should rest closed on the top cover.

Replace the Lamp and Clean the Contacts

The reader's lamp, on average, should provide 500 hours of total service before it needs to be replaced. This equates to approximately six months to one year of service under routine conditions. The intensity of the lamp will slowly drop over time until the reader's run-time self-check detects a low signal and warns the user with a displayed error code. The bulb should be replaced at this time (PN 3400508), and the electrical contacts in the lamp socket cleaned. The figure below provides a detailed view of access to the lamp.
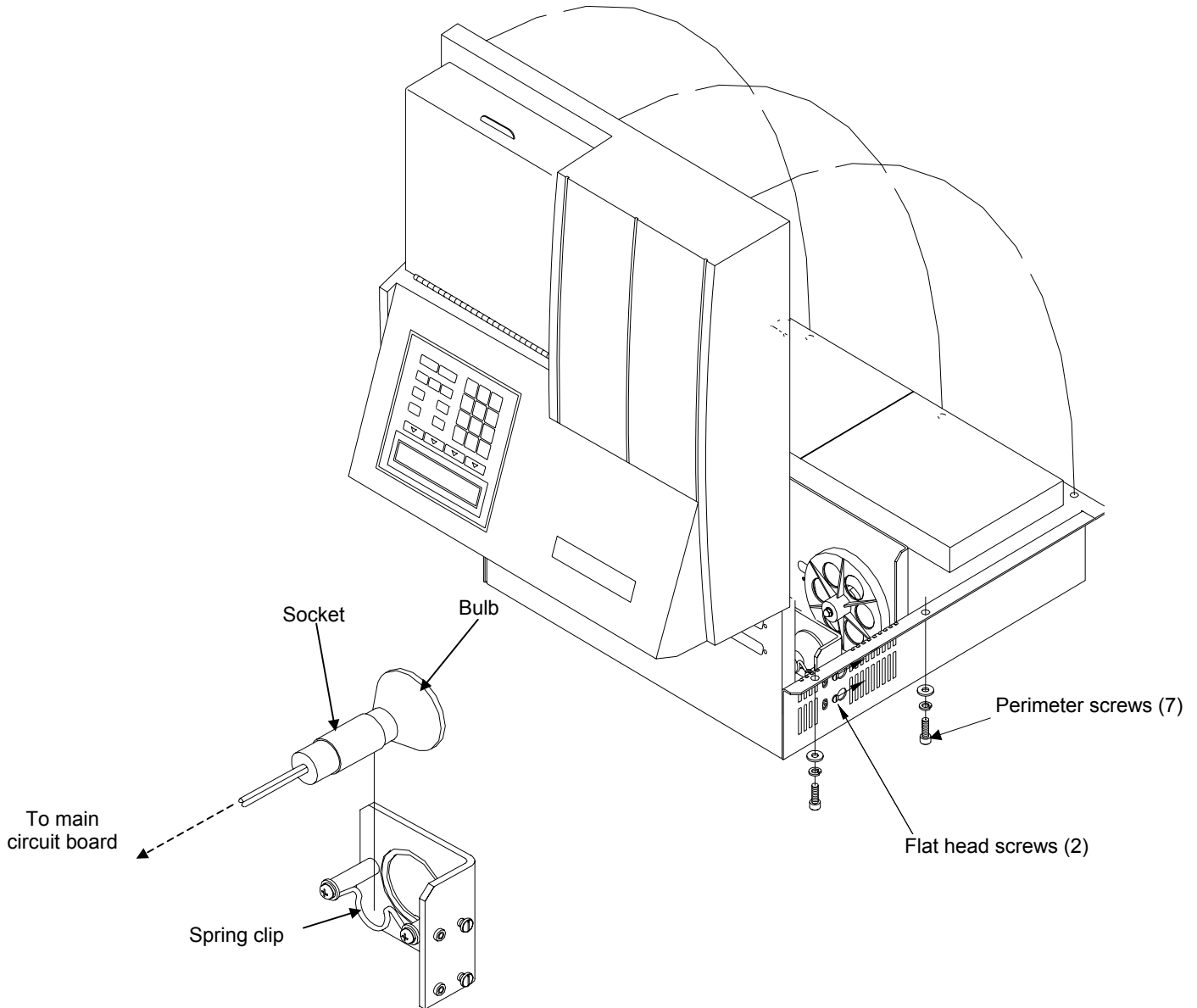
Figure 5-3: *Location of the lamp in the ELx808™*

Warning! Hot surface. The lamp is hot when the instrument is turned on. Turn off and unplug the reader. Allow the lamp to cool down before attempting to replace it.

Oxide coating on the electrical contacts can reduce lamp output. Clean the contacts in the socket whenever the lamp is replaced (approximately every 500 hours).

You will need:

- Tape
- Flat-blade screwdriver
- Cotton swabs or pencil eraser
- Isopropyl alcohol
- Lamp, BioTek PN 3400508

To replace the lamp:

1. Turn off and unplug the reader. Allow the lamp to cool for at least ten minutes.

2. Temporarily tape the reader's plate access door shut.

3. Using your fingertips, locate the seven securing (perimeter) screws around the sides and front where the reader's top shroud meets the base. Remove the screws using a flat-blade screwdriver. *Tip:* Bring the reader to the edge of the work surface to access the screws without having to turn the reader upside down.

4. Carefully raise the front of the shroud and tip it toward the back of the instrument.

5. The lamp is located in the left rear quadrant inside the reader. Unplug the lamp connector from the circuit board.

   Note: Do not touch the glass lens in the lamp bracket.

6. The lamp bracket is attached to the side of the instrument with two screws, which are accessed from the outer left side of the instrument. Using a flat-blade screwdriver, loosen (do not remove) these two screws. Slide the bracket out of the instrument.

7. Remove the bulb from its socket by pushing the socket and the bulb together and rotating the socket counterclockwise.

8. Clean the two contacts in the socket, using a cotton swab moistened with isopropyl alcohol, and allow them to dry for a few minutes. An alternative cleaning technique would be to rub the contacts with a pencil eraser.

To **install** the lamp, reverse steps 1 through 7. Keep your fingers out of the reflector interior and away from the bulb. Ensure that the bulb is set flat against its housing before reinstallation in the instrument.

Decontamination

This appendix contains a procedure for decontaminating the ELx808™.

This appendix contains the following sections:

Purpose

Any laboratory instrument that has been used for research or clinical analysis is considered a biohazard and requires decontamination prior to handling.

Decontamination minimizes the risk to all who come in contact with the instrument during shipping, handling, and servicing. Decontamination is required by the U.S. Department of Transportation regulations.

Persons performing the decontamination process must be familiar with the basic setup and operation of the instrument.

BioTek Instruments, Inc. recommends the use of the following decontamination solutions and methods based on our knowledge of the instrument and recommendations of the Centers for Disease Control and Prevention (CDC). Neither BioTek nor the CDC assumes any liability for the adequacy of these solutions and methods. Each laboratory must ensure that decontamination procedures are adequate for the Biohazard(s) they handle.

Wear prophylactic gloves when handling contaminated instruments. Gloved hands should be considered contaminated at all times; keep gloved hands away from eyes, mouth, and nose. Eating or drinking while decontaminating instruments is not advised.

Mucous membranes are considered prime entry routes for infectious agents. Wear eye protection and a surgical mask when there is a possibility of aerosol contamination. Intact skin is generally considered an effective barrier against infectious organisms; however, small abrasions and cuts may not always be visible. Where protective gloves when performing the decontamination procedure.

Tools and Supplies

- Sodium hypochlorite (NaClO, or bleach)

- 70% isopropyl alcohol (alternative to bleach)

- Deionized or distilled water

- Safety glasses

- Surgical mask

- Protective gloves

- Lab coat

- Biohazard trash bags

- 125 ml beakers

- Clean cotton cloths

Decontamination Procedure

⚠️ The bleach solution is caustic; wear gloves and eye protection when handling the solution.

Do not immerse the instrument, spray it with liquid, or use a "wet" cloth. Do not allow the cleaning solution to run into the interior of the instrument. If this happens, contact the BioTek Service Department.

Do not soak the keypad – this will cause damage.

ⓘ **Important!** Turn off and unplug the instrument for all decontamination and cleaning operations.

1. Turn off and unplug the instrument.

2. Prepare an aqueous solution of 0.5% sodium hypochlorite (bleach). As an alternative, 70% isopropyl alcohol may be used if the effects of bleach are a concern.

    ➢   Be sure to check the percent NaClO of the bleach you are using; this information is printed on the side of the bottle. Commercial bleach is typically 10.0% NaClO; if this is the case, prepare a 1:20 dilution. Household bleach is typically 5.0% NaClO; if this is the case prepare a 1:10 dilution.

3. Moisten a cloth with the bleach solution or alcohol. Do not soak the cloth. Wipe down the carrier and all exposed instrument surfaces with the bleach solution.

    ➢   Wipe the keypad (do not soak). Wipe again with a clean cloth moistened with deionized or distilled water. Dry immediately with a clean, dry cloth.

    ➢   Wipe the plate carrier, the inside of the carrier door, and all exposed surfaces of the instrument.

4. Wait 20 minutes. Moisten a cloth with deionized or distilled water and wipe all surfaces of the instrument that have been cleaned with the bleach solution or alcohol.

5. Use a clean, dry cloth to dry all wet surfaces.

6. Reassemble the instrument as necessary.

7. Discard the used gloves and cloths using a Biohazard trash bag and an approved Biohazard container.

Computer Control

The ELx808™ can be controlled either from the reader's front panel or from a computer connected to the reader via the computer's serial port. This appendix describes the features of computer control, and explains the information necessary to program the computer to control the reader.

This appendix contains the following sections:

Overview

With the computer control feature, the user is provided even more power and flexibility. For example, the ELx808™ can define and run computer-controlled kinetic assays using up to three wavelengths on the same microplate.

When using computer control, the ELx808's onboard blanking and data reduction calculations are suppressed and the raw data is returned to the controlling software for evaluation. Readings higher than 3.000 OD may be transmitted.

This appendix details the protocols necessary for communicating with the ELx808 reader. In addition, instructions are provided for controlling the reader with BioTek's Gen5™, KCjunior™, or KC4™ software packages.

Controlling the Reader With Gen5™

⁞⁞⁞	Before installing Gen5™, verify that your computer meets the minimum system requirements specified in Gen5's Help system or Getting Started Guide.

Setting Up Gen5

The ELx808™ can be operated via BioTek's Gen5 software installed on a host computer. The following instructions *briefly* show you how to set up Gen5 for operation of the reader. Refer to Gen5's Getting Started Guide or Help system for more detailed instructions.

1. Turn off the computer and the reader. Connect the appropriate serial cable (PN 75053) between the two machines.

2. Power up both machines.

3. Install Gen5 on the computer's hard drive and register the software with BioTek.

    - If you purchased Gen5's Reader Diagnostics Utility (required for the Absorbance Plate Test), install this software on the computer's hard drive.

4. Start Gen5.

5. Login if prompted. The default System Administrator password is **admin**.

6. When the 'Welcome to Gen5' screen appears, select **System Menu**.

7. From Gen5's main screen, select **System|Reader Configuration** to open the 'Reader Configuration' dialog.

8. Click the **Add** button to open the 'Reader Settings' dialog.

    - Gen5 and Gen5 Secure: Up to two readers may be added in Gen5.

9. Use the drop-down list in **Reader Type** to select **ELx808.**

10. Enter the appropriate **Com Port**.

    - The **Baud Rate** is set to the default transmission speed of **9600** for the ELx808 and may not be changed.

11. Click **Test Comm.** Gen5 will attempt to communicate with the reader.

    - If you receive "The reader is communicating!" message, click **OK** and then click **OK** again to save the settings. Click **Close** at the Reader Configuration dialog to return to the main screen.

- If the test is not successful and you receive an error message, refer to the Problems section below or to the Troubleshooting section of Gen5's Help system for assistance.

- Gen5 Secure only: An 'Audit Trail' dialog will appear after exiting Reader Configuration, whenever you add, modify, or delete a reader. If desired, enter any comments, then click **Close**.

Problems

If Gen5™ fails to communicate with the reader and displays a serial communications error, try the following troubleshooting suggestions:

- Confirm that the correct Reader Type was selected in step 9.

- Try a different COM port.

- Check the serial cable connections. Ensure that the cable is properly attached to the port defined in step 10, and is not a Null cable. If this is suspected, add another Null and try again.

- Confirm that the reader has passed its Self Test. The reader will not communicate if it fails an internal system test.

If the test still fails, refer to the Troubleshooting section in Gen5's Help system for further assistance.

Getting Started With Gen5

The following instructions *briefly* show you how to perform a "Quick Read" in Gen5™ (**File|New Experiment | Default Protocol**). It's called "Quick" because you can perform a reading without having to take the time to create a new protocol.

If the reading is part of an experiment or assay that you will perform numerous times, you will need to create a new protocol (**File|New Protocol**).

Refer to Gen5's Help system early and often to learn how to create protocols, assign well identifiers, read plates, print reports, perform data reduction, and more.

To perform a "Quick Read":

1. At Gen5's Welcome screen, select **System Menu|File|New Experiment**. (Alternative: select **Read a Plate** at the Welcome screen, then proceed to step 4.)

2. Click **Default Protocol**, then click **OK**. Gen5 will open the Experiment workspace, which includes the Protocol menu tree and Plate screen.

3. Select **Plate|Read** or click the **Read** button. The 'Procedure' dialog will open.

- Gen 5 and Gen5 Secure: If more than one reader was added in Gen5, the 'Instrument Selection' dialog will appear instead of the Procedure dialog. Select **ELx808**, then click **OK**. The Procedure dialog will then appear.

4. Select a **Plate Type**.

5. Click **Read** to open the'Read Step' dialog.

6. Select a **Read Type**.

7. Define the wavelength(s) at which the plate will be read.

8. Define other reading parameters as desired. Click the **Help** button for assistance.

9. When complete, click **OK** to return to the Procedure dialog.

- Click **Validate** if you would like Gen5™ to verify the defined parameters. If all parameters are valid, you will receive confirmation. If any parameters are invalid, Gen5 will provide information for correcting the problem. Refer also to the Troubleshooting section of the Help system.

10. Click **OK** again to save and close the Procedure dialog. The 'Plate Reading' dialog will open.

11. Enter any desired information, place the plate on the carrier, then click **READ** to begin the plate read. Click **OK** when the 'Load Plate' dialog appears. The plate will be read.

- To view the raw data results, use the **Data** drop-down arrow in the Plate screen to select one wavelength. The results will be displayed for the selected wavelength. Repeat, for other wavelengths.

- To analyze, manipulate, or print results, **Protocol** parameters should be defined. Refer to Gen5's Help system for instructions.

   Note: Gen5 Reader Control does not support data reduction.

Controlling the Reader With KCjunior™

ïïï	Before installing KCjunior™, verify that your computer meets the minimum system requirements specified in KCjunior's User's Guide or Help system.

Setting Up KCjunior

The ELx808™ can be operated via BioTek's KCjunior software installed on a host computer. The following instructions *briefly* show you how to set up KCjunior for operation of the reader. Refer to KCjunior's User's Guide or Help system for more detailed instructions.

1. Turn off the computer and the reader. Connect the appropriate serial cable (PN 75053) between the two machines.

2. Turn on both machines.

3. Install KCjunior on the computer's hard drive and register the software with BioTek.

4. Once installed, start KCjunior.

5. Select **Setup**|**Reader 1** (or **Reader 2)**.

6. To select the reader and define the communications parameters, choose the following setup parameters:

Reader:	**ELx808**, **EL808**, or
	ELx808I, **EL808I**, or
	ELx808U, **EL808U**, or
	ELx808UI, **EL808UI**
Com Port:	Select the COM port used for the RS-232 serial cable connection
Baud Rate:	**9600** (change the baud rate on the reader if necessary)
Data Bits:	**8**
Parity:	**None**
Stop Bits:	**2**
EOT Character:	Keep the default number.

7. Click the **Test Communications** button to attempt to establish communications with the reader, using the currently defined communication parameters. If a 'Serial Write Error' dialog is displayed, an incorrect COM port may have been selected. Select a different port and then repeat this step.

8. If the test passes, click **OK** to save the settings and close the dialog box. If the test fails, follow the directions provided by KCjunior, then click **Test Communications** again. See also the *Problems* section below.

Problems

If KCjunior™ fails to communicate with the reader, and displays a serial communications error, check the cable plug-in location to make sure it matches the setup choices and is not a Null cable. If this is suspected, add another Null and try again.

Getting Started With KCjunior

The following instructions *briefly* describe how to read a plate using KCjunior. Refer to KCjunior's Help system and User's Guide early and often to learn how to create protocols, assign well identifiers, read plates, print reports, and more.

To read a plate using KCjunior:

1. Click **Read Plate** from KCjunior's main screen. The 'Read Plate' dialog will appear.

2. If desired, enter a **Results ID** and a **Plate Description**, and then click **Read Plate**. The 'Protocol Definition' dialog will appear.

3. Select a **Read Method Type**.

4. Define the wavelength(s) at which the plate will be read.

5. Select a **Plate Geometry** from the drop-down list.

6. Define other reading parameters as necessary. Click the **Help** button for assistance.

7. When complete, click **OK** to return to the 'Read Plate' dialog.

8. If desired, enter a **Plate ID**.

9. Place the plate on the carrier, then click **OK** to start the plate read.

   • The plate will be read and then the raw data results will display in KCjunior. Print the raw data by selecting **Plate|Print Results**.

   • To analyze or manipulate results, a **Protocol** should be defined. Refer to KCjunior's Help system or User's Guide for instructions.

Controlling the Reader With KC4™

iii	Before installing KC4™, verify that your computer meets the minimum system requirements specified in KC4's User's Guide or Help system.

Setting Up KC4

The ELx808™ can be operated via BioTek's KC4 software installed on a host computer. The following instructions *briefly* show you how to set up KC4 for operation of the reader. Refer to KC4's User's Guide or Help system for more detailed instructions.

1. Turn off the computer and the reader. Connect the appropriate serial cable (PN 75053) between the two machines.

2. Turn on both machines.

3. Install KC4 on the computer's hard drive and register the software according to the instructions in the KC4 User's Guide.

4. Once installed, start KC4.

5. Select **System|Readers**.

6. Scroll through the list of **Available Readers** and select the appropriate ELx808 reader model. Click the **Port** button (and subsequent **Setup** button), to define the following communications parameters:

Port:	Select the COM port used for the RS-232 serial cable connection
Transmission Speed:	**9600** (default), **1200**, or **2400** (must match the baud rate on the reader)
Data Bits:	**8**
Parity:	**No**
Stop Bits:	**2**

7. Click **Current Reader** button to attempt to establish communication with the reader, using the currently defined communication parameters.

8. If the test passes, click **OK** to save the settings and close the dialog box. If the test fails, KC4 will provide appropriate instructions for resolving any problems. See also the ***Problems*** section on the next page.

Problems

If KC4™ fails to communicate with the reader and displays a serial communications error, check the cable plug-in location to ensure that it matches the setup choices and is not a Null cable. If this is suspected, add another Null and try again.

If an 'Incorrect Reader Model Connected' dialog is displayed, click **OK** to clear the screen and select **System|Readers|Available Readers**. Verify that the reader selected is correct.

Getting Started With KC4

The following instructions *briefly* describe how to read a plate using KC4. Refer to KC4's Help system and User's Guide early and often to learn how to create protocols, assign well identifiers, read plates, print reports, and more.

To read a plate using KC4:

1. Select **Data|New Plate**.

2. If prompted to select a protocol, select **Empty Protocol** and click **OK**. If not prompted, select **Protocol|New**, or use KC4's Protocol Wizard to step through protocol creation.

3. Select **Protocol|Reading**. The 'Reading' parameters dialog will appear.

4. Select a **Reading Type**.

5. Define the **Filters** (wavelengths) at which the plate will be read.

6. Define other reading parameters as necessary. Click the **Help** button for assistance.

7. When complete, click **OK**.

8. Select **Data|Read Plate**. The 'Plate Reading' dialog will appear.

9. Enter any comments, place the plate on the carrier, then click **START READING** to begin the plate read.

    - The plate will be read and then the raw data results will display in KC4.

    - To analyze, manipulate, or print results, **Protocol** parameters should be defined. Refer to KC4's Help system or User's Guide for instructions.

Controlling the Reader Using Serial Protocol

At baud rates of 1200, 2400, and 9600, the ELx808™ is capable of sending and receiving data through its serial port (RS-232C). The baud rate used for transmission is held in non-volatile memory and can be changed by the user. Other serial port parameters, Parity (None), Data Bits (8), and Stop Bits (2) are fixed and cannot be changed.

The reader's RS-232C serial port is configured as a DTE; that is, the unit is wired to resemble a modem. Data is received on Pin 3 (the RX Pin), and transmitted on Pin 2 (the TX pin).

See *Connect the Host PC* in *Chapter 2*, *Installation*.

Computer Control Command Set

A command from the computer to the reader consists of a single ASCII character, and in some cases, subsequent argument data. Upon receipt of a valid command character, the reader returns an <ACK> character. Some commands also return response data to the host computer. Upon completion of command processing, the ELx808 transmits a status string to the computer. When the reader sends data to the computer, the data is sent first, then the status string.

While awaiting a command, the ELx808 responds to nulls or other unexpected characters by clearing its input buffer and transmitting a <NAK>. Therefore, if valid commands are preceded by invalid characters, they may be missed.

Refer to *Table B-1* for the **ASCII Control Characters** used in the computer control protocol.

All ASCII character strings representing numbers or names are transmitted most significant digit or letter first. Data values not indicated as ASCII are treated as binary integers, and are transmitted least significant byte first.

Some commands described here may not be available with older reader basecode versions (prior to v3.00), indicated if the Check Feature command returns a <NAK>. These commands are marked with a '#'.

Table B-1
ASCII Control Characters Used in Computer Control Protocol

ASCII Code	Function	Hex Code	Decimal Code	Control Code	Reader <-- >
ACK	Acknowledge	06	06	^F	--->
NAK	Negative acknowledge	15	21	^U	--->
RS	Record separator	1E	30	^^	--->
ETX	End of text	03	03	^C	<-->
DLE	Data link escape	10	16	^P	--->
CR	Carriage return	0D	13	^M	<---
LF	Line feed	0A	10	^J	<---
CTRL-Z	Control Z	1A	26	^Z	<---

The following section describes the supported computer control command set.

STORE PLATE CARRIER ('A')

This command causes the plate carrier to move inside the instrument.

host:	valid limits:	response:
'A'	no arguments	<ACK>
		status string (5)

READ WELL SET ('d')

This command causes the reader to return data collected at the center of each of the indicated wells, in the order specified. The parameters for the actual data collection are taken from the currently loaded assay definition table (see Set Assay Definition).

host:	valid limits:	response:
'd'		<ACK/NAK>
data bytes:		
1 – 2	well #1 row ("01" – "16")	
3 – 4	well #1 column ("01" – "24")	
5 – 8	well #2	
9 – 12	well #3	
13 – 16	well #4	
17 – 20	well #5	
21 – 24	well #6	
25 – 28	well #7	
29 – 32	well #8	
33	position in read series	
	'0': single read	
	'1': first read in series	
	'2': next read in series	
	'3': final read in series	
<ETX>		<ACK> if data valid

reader response protocol:

initial status string (5)

for each kinetic interval (one if endpoint read):

    for each wavelength:

data start code (1)		\<CR\>
for each of eight wells:		
comma separator (1)	','	
sign (1)		'+' or '-'
data (4)		"1234"
row terminator (2)		\<CR\>, \<LF\>
temperature (3)	***(if specified)**	"000" – "999"
wavelength terminator (1)		\<^Z\>
checksum (1)		0 - 255

final terminator (1)   \<^Z\>

final status string (5)   **(ELx status mode only)**

*** Temperature response included only if requested. Format is three ASCII bytes scaled up by ten, i.e. "275"correponds to 27.5 degrees C. See the Include Temperature Response command.

Data is returned in the order wells are specified. If fewer than eight wells are to be read, each well's unused row and column must be replaced with "00". Data is always returned for eight wells, regardless of the actual number specified.

See the Read Plate command for details concerning results data format, and error handling and status string information. This command always returns data using the **ELx format**.

GET BASECODE VERSION ('e')

This command returns a string containing the part number and version number for the instrument downloadable basecode software.

host:	valid limits:	response:
'e'	no arguments	\<ACK/NAK\>

reader response protocol:

part number (7)	"0000000" – "9999999"
intermediate string (10)	" Version "
version code (7)	"x.xxyyy"
status string (5)	

The version number is indicated by the first four characters of the version code (x.xx). The remaining characters in the version code (yyy) may either be blank or represent a specific beta or prototype release code.

INCLUDE TEMPERATURE RESPONSE ('i')

This command causes the current temperature to be included in the results response for each plate read using the 'S', 'c', or 'd' commands.

host:	valid limits:	response:
'i'		<ACK/NAK>
data bytes:	inclusion	
	'0': no temperature with plate data	
	'1': include temperature with plate data	

By default the instrument is initialized to not include a temperature response, and automatically reverts to this default when a new assay definition table is loaded, either from the front panel or by computer control.

PRESENT PLATE CARRIER ('J')

This command causes the plate carrier to move back outside the instrument where it can be loaded with a microplate.

host:	valid limits:	response:
'J'	no arguments	<ACK>
		status string (5)

SET READ MODE ('k')

This command selects the read mode controlling the speed, accuracy, and precision of all subsequent plate reads.

host:	valid limits:	response:
'k'	no arguments	<ACK>
data bytes:	mode	
	'0': slow (enhanced) mode	
	'1': fast mode	
		status string (5)

SET STATUS FORMAT ('n')

This command selects the format for the status string (see Status String Format).

host:	valid limits:	response:
'n'	no arguments	<ACK>
data bytes:	format	
	'0': 312 status mode	
	'1': ELx status mode	
		status string (5)

When the status format is set to **ELx status mode**, any error detected during the execution of any command described in this document will be transmitted to the host PC in this format; in addition, plate read and scan commands ('d' and 'S') shall return an additional status string at the very end of the read sequence. See Status String Format for error descriptions.

GET CURRENT STATUS ('o')

This command returns the most recent error status set by the instrument. In the event of a reader timeout or <DLE> receipt, it can be used to determine the error code associated with the reader failure. It can also be used in place of pressing a key on the reader to stop the beeping after an error. The status is always returned in the **ELx status format**.

host:	valid limits:	response:
'o'	no arguments	<ACK>
		status string (5)

Requesting the current status through the use of this command will cause the status maintained on-board the instrument to be cleared except in the following situations:

1. A fatal error has been detected. The only way to clear this type of error is by recycling power on the instrument.

2. An error has been detected which requires a self-test to be executed before a read operation can be performed. In this case, only a successful self-test will cause the error to be cleared.

This command is most useful when invoked in pairs. The first try will yield the most recent error detected (or no error), which will then be cleared by the reader if possible. The next call will then yield either no error, or a previously-detected error that still needs to be cleared by running a self-test. Commands other than read commands may always be sent regardless of the current reader status; if there is a hardware issue preventing successful execution, a subsequent error status will be generated.

See Status String Format for error descriptions.

SET QUIET MODE ('q')

This command allows the user to select a "quiet" mode. When this mode is turned on, the reader bypasses the process of displaying an error code and beeping until a key is pressed. As long as the quiet mode is selected, errors detected while processing serial-control commands are available only if ELx Status Mode is selected, or in response to the Get Current Status ('o') command. Serial-only instruments are set to quiet mode automatically at powerup.

host:	valid limits:	response:
'q'	no arguments	<ACK>
data bytes:	mode	
	'0': off	
	'1': on (quiet)	
		status string (5)

READ PLATE ('S')

This command causes a microplate to be read according to the currently loaded assay definition table.

host:	valid limits:	response:
'S'	no arguments	<ACK/NAK>

reader response protocol:

status string (5)

for each kinetic interval (one if endpoint read):

    for each wavelength expected: **(312 format: one, ELx format: up to three)**

        wavelength start code (1)   <CR>

        for each row within specified row range:

            for each column (well) within specified column range:

                comma separator (1)   ','

                sign (1)   '+' or '-'

                data (4)   "1234"

            row terminator (2)   <CR>, <LF>

        temperature (3)   *****(if specified)**   "000" – "999"

    assay name (8)   **(312 format only)**   "ABCDEF", <CR>, <LF>

    unused extra characters (49)   **(312 format only)**

        wavelength terminator (1)   <^Z>

        checksum (1)   **(ELx format only)**   0 - 255

final terminator (1)   **(ELx format only)**   <^Z>

final status string (5)   **(ELx status mode only)**

*** Temperature response included only if requested. Format is three ASCII bytes scaled up by ten, i.e. "275"correponds to 27.5 degrees C. See the Include Temperature Response command.

Absorbance data is returned in the form of a 5-character ASCII string, with a sign character followed by four digits. A decimal point is not included, but should be inserted by the user after the first digit, i.e. "+1234" should be translated as +1.234 OD. Overrange readings are indicated by the 5-character string "*****" instead of a signed numerical value.

If dual-wavelength subtraction has been selected in the reader definition table, the subtraction is performed by the reader and the result is sent as a single plate representing the difference in ODs per well. Otherwise a complete plate of ODs is sent for each wavelength selected.

If an error is detected during a plate read, or the user aborts the process via serial 'X' or stop key press, the entire (remaining) data response is replaced by a single <DLE> character (0x10). In the ELx Status Mode, the final status string then indicates the error detected. In the case of reader error, data previously received during the current read sequence should be discarded.

312 Format:

This format is selected by default at reader powerup, and every time a new assay definition is loaded, either via serial control 'V' command, or by the user from the front panel. The reader always returns data as an 8 x 12 matrix, along with the assay name and extra characters. There are no checksums or final terminator.

ELx Format:

If any of the following occurs, the ELx format is selected and remains enabled until a new definition is loaded, either via serial control 'V' command, or by the user from the front panel:

1. Multi-wavelength mode is selected in the assay definition

2. A range of rows is set, even if for all rows (see Set Row Range).

3. A range of columns is set, even if for all columns (see Set Column Range).

The reader returns data in a matrix corresponding to the rows and columns selected, for up to three wavelengths. If dual-wavelength subtraction is selected, only one plate shall be returned (not available in the multi-wavelength mode).

Checksum calculation starts with the first byte AFTER the wavelength start code, up through and including the <^Z> data terminator code at the end of each wavelength's plate data. The checksum is transmitted as an integer data byte (not ASCII).

The final terminator indicates that the data stream is complete for the corresponding Read Plate command. No more data shall be transmitted until another command is sent.

312 Status Format:

If the middle character in the status string is not ASCII '0', an error has been detected and no results data is transmitted.

ELx Status Format:

If any character in the initial status string is not ASCII '0', an error has been detected and no results data is transmitted. Otherwise, if an error is detected during the read process, the final status string reflects that error.

See Status String Format for error descriptions.

SET ASSAY DEFINITION ('V')

This command downloads to the reader an assay definition table to be used in subsequent plate reads or wellset reads ('S' or 'd' commands) until a new definition replaces it, either via serial control or by the user from the front panel.

host:		valid limits:	response:
'V'			<ACK/NAK>
data bytes:			
1		don't care	
2 - 7		assay name (ASCII characters)	
8 - 29		don't care	
30		subtraction (0x00: none, 0x40: dual wavelength)	
31		encoded byte:	
	bits 0 - 3 (0x0F)	0	
	bit 4 (0x10)	1: shake, 0: no shake	
	bit 5 (0x20)	1: shake before every read, 0: before first read only	
	bit 6 (0x40)	1: continuous shake, 0: timed shake	
	bit 7 (0x80)	1: use total kinetic time, 0: use kinetic read count	
32		1D scanning points (1 – 31, must be odd)	
33 - 49		don't care	
50 – 52		measurement wavelength ("000", "340" - "900")	
53 - 55		reference wavelength ("000", "340" - "900")	
56		read type (0: endpoint, 2: kinetic, 3: 1D scan)	
57 - 59		don't care	
60 - 61		kinetic interval (seconds, 0 - 9999)	
62 - 63		kinetic read count (2 - 9999)	
64 - 66		don't care	
67 - 69		first wavelength ("000", "340" - "900")	
70 - 84		remaining wavelengths ("000", "340" - "900")	
85 - 162		don't care	

163 - 164	shake time (seconds, 0 - 999)	
165	shake speed (0: slow, 1: med, 2: fast, 3: variable)	
166 - 167	total kinetic read time (minutes, 1 - 9999)	
168	delay before first read (0: no, 1: yes)	
169 - 170	delay (seconds, 0 - 999)	
	status string (5)	

The multi-wavelength mode returns a plate of data for each wavelength specified. To select the multi-wavelength mode, the "measurement wavelength" must be set to "000", and wavelengths specified starting with "first wavelength". Up to three wavelengths may be specified in this mode. However, if dual-wavelength subtraction is desired, the measurement and reference wavelengths must be specified. A single-wavelength read may use either mode.

If a wavelength is not to be used, it must be replaced by "000".

If the multi-wavelength mode is selected, the **ELx format** for data response will be used until a new assay definition table is specified.

A kinetic interval of zero will cause the plate to be read repeatedly with no pauses between reads.

The assay definition table must be downloaded before other plate-specific commands are sent, such as Set Row Range and Set Column Range.

To minimize confusion, all "don't cares" should be set to 0.

GET WAVELENGTH TABLE ('W')

This command uploads the wavelength setup table from the reader.

host:	valid limits:	response:
'W'	no arguments	<ACK/NAK>

reader response protocol:

for each wavelength configured (six):

wavelength		("000 - "900")
comma terminator	','	

status string (5)

HALT ('X')

This command causes any read in progress to be halted.

host:	valid limits:	response:
'X'	no arguments	none (see below)

The following events occur when this command is invoked:

1. The read process is halted, and all axes are returned to their home positions (the plate is moved back outside where it can be accessed).

2. The reader transmits a <DLE> character to the computer when the above process is completed.

3. No more data is transmitted by the read or scan in progress. A final status string is transmitted in the ELx Status Mode.

 # SET ROW RANGE ('r')

This command selects a range of adjacent rows (lettered on plate or strip carrier) to be read.

host:	valid limits:	response:
'%'		<ACK/NAK>
data bytes:		
1	first row (1 - 16)	
2	last row (1 - 16)	
		status string (5)

By default the instrument is initialized to perform full plate reads, and automatically reverts to this default when a new assay definition table is loaded, either from the front panel or by computer control.

Once this command is invoked, the reader will use the **ELx format** for data response until a new assay definition table is specified.

This command has no effect on the read parameters for a wellset read.

SET COLUMN RANGE ('%')

This command selects a range of adjacent columns (numbered on plate or strip carrier) to be read.

host:	valid limits:	response:
'%'		<ACK/NAK>
data bytes:		
1	first column (1 - 24)	
2	last column (1 - 24)	
		status string (5)

By default the instrument is initialized to perform full plate reads, and automatically reverts to this default when a new assay definition table is loaded, either from the front panel or by computer control. The well range is also reset when the Set Geometry command is sent.

Once this command is invoked, the reader will use the **ELx format** for data response until a new assay definition table is specified.

This command has no effect on the read parameters for a wellset read.

GET MINIMUM KINETIC INTERVAL ('$')

This command provides a means for the user to determine the minimum kinetic read interval, as dictated by the current assay definition table, well range, and even baud rate selected.

host:	valid limits	response:
'$'	no arguments	<ACK/NAK>
		kinetic interval (2)
		status string (5)

The reader returns the minimum kinetic interval as a 2-byte integer value (not ASCII, so low byte first).

Minimum kinetic intervals are not available for wellset reads.

SELF-TEST ('*')

This command causes the reader to perform a system self-test and calibration. This should be performed any time the wavelength configuration is changed ('M' command).

host:	valid limits:	response:
'*'	no arguments	<ACK/NAK>
		data stream
		status string (5)

The reader responds by sending a variable-sized stream of ASCII character data representing various calibration and test results. This data stream is followed by the standard status response string.

SET TEMPERATURE ('[')

This command sets the incubation chamber temperature setpoint.

host:	valid limits:	response:
'['		<ACK/NAK>
data bytes:		
1 - 2	temperature setpoint (0, 22 - 50)	
		status string (5)

If the instrument is not equipped with a working incubator, or if the indicated temperature setpoint is out of range, an incubator setpoint error will be returned with the standard status response string.   A setpoint of zero will turn the incubator heaters off.

GET TEMPERATURE (']')

This command returns the current temperature in the incubation chamber.

host:	valid limits	response:
']'	no arguments	<ACK/NAK>
		temperature (2)
		status string (5)

The reader returns the current temperature as a 2-byte integer value (not ASCII, so low byte first).

Temperature is returned scaled up by 10, i.e., 370 indicates a temperature of 37.0 degrees Celsius. Temperatures below the instrument minimum of 18 degrees are returned as 111 (0x006F); temperatures above the instrument maximum of 55 degrees are returned as 999 (0x03E7).

If the instrument does not have incubation, or an incubation error has been detected, 0x0000 will be returned as temperature data. An incubator temperature error will then be flagged and returned with the standard status response string. Otherwise, the current temperature (averaged over four thermal zones) is returned as defined above.

The reader will accept and process the Get Temperature command at any time, including during a read cycle. The temperature response will NOT interrupt a plate data response stream, however. If a Get Temperature command is received during a data transmission, it will not be processed until the transmission has completed, i.e. after the <^Z> checksum combination has been sent.

GET PLATE BARCODE ('s')

This command returns the barcode for the microplate currently in the instrument carrier.

host:	valid limits	response:
's'	no arguments	<ACK/NAK>
		barcode (32)
		status string (5)

The barcode is returned as a stream of 32 characters. If the barcode itself is less than 32 characters it is padded with nulls (0x00) up through the 32nd character. Otherwise the barcode is truncated to 32 characters and sent with no null terminator.

If no barcode is detected after two passes of the microplate carrier in front of the scanner, the null-terminated string "NR" is returned, followed by the remaining 29 nulls. A barcode error is flagged and returned with the standard status response string.

OPEN PLATE ACCESS DOOR ('(')

This command opens the robotic plate access door

host:	valid limits	response:
'('	no arguments	<ACK/NAK>
		status string (5)

CLOSE PLATE ACCESS DOOR (')')

This command closes the robotic plate access door

host:	valid limits	response:
')'	no arguments	<ACK/NAK>
		status string (5)

GET INSTRUMENT CONFIGURATION ('}')

This command returns a 16-bit binary-encoded word defining the instrument configuration.

host:	valid limits	response:
'}'	no arguments	<ACK/NAK>
		configuration (2)
		no status string

Instrument configuration data is returned low byte first. Encoding is defined as follows:

code:	description:
0x0000	ELx808-UV
0x0001	ELx808
0x0008	ELx808-UV w/ Incubator
0x0009	ELx808 w/ Incubator
0x0080	ELx808-UV w/ Robot Access Door
0x0081	ELx808 w/ Robot Access Door
0x0088	ELx808-UV w/ Incubator and Robotic Access Door
0x0089	ELx808 w/ Incubator and Robotic Access Door

CHECK FEATURE ('z')

This command answers queries regarding features that may have been added after the initial release of an instrument.

host:	valid limits:	response:
'z'	no arguments	<ACK/NAK>

data bytes:

1 - 4	updated feature:	associated commands:
	"1001": well set reads	'd'
	"1002": reads using two wavelength sets	(legacy - 'V')
	"1003": area scanning	(not available)
	"1004": serial wavelength configuration	(legacy – 'M')
	"1005": shaking	(legacy – 'V')
	"1006": absorbance reads	(legacy)
	"1007": high-speed sweep reads	(not available)
	"1008": partial plate reads	'r', '%', '$'
	"1009": incubation	(legacy, 'I' readers only - '[', ']', 'i')
	"1010": sensitivity response	(not available)
	"1011": multi-wavelength read format	'V'
	"1012": serial read mode selection	'k'
	"1013": ELx format status response	'n', 'o'
	"1014": serial-only instrument	
	"1015": basecode checksum	'p'
	"1016": barcode scanning	's'

<ETX>

reader response protocol:

feature available (1)   '0' / '1'

miscellaneous additional data (16)

status string (5)

If the 'z' is acknowledged (ACK), the instrument shall then return '1' if the feature is available, '0' if not.

If the 'z' command is not recognized (indicated by a NAK response to the 'z'), it means the instrument basecode version was released prior to implementation of this command (along with the other commands marked as such with a '#'). In this case, if the feature is marked in the table above as "legacy", the indicated commands and functions are and always have been available anyway.

Using the Stop Key to Halt Plate Reads

Pressing the **STOP** key on the reader while a computer-control-initiated plate read is in progress causes the following to occur.

1. The read process is halted, and all axes are returned to their home positions (the plate is moved back outside where it can be accessed).

2. The reader transmits a <DLE> character to the computer when the above process is completed.

3. No more data is transmitted by the read in progress. A final status string is transmitted in the ELx Status Mode.

Status String Format

Following execution of each command (except as noted), the *ELx808* sends a status string back to the computer.

312 Status Mode Format:

This status string consists of 5 successive ASCII characters -- RS, S1, S2, S3, and ETX:

    ⇒ **RS**   A record separator that marks the beginning of the status string.

    ⇒ **S1**   Always ASCII zero ('0')

    ⇒ **S2**   A single digit, used as a reader fault or error code number.

        ERROR CODES

        Error codes indicate the following:

        '0': no fault or error

        '8': instrument failure - perform self-test

        '9': error in assay, scan, or table definition

        'A': error in well range selection

        'B': incubator setpoint error

        'C': incubator temperature error

        'E': barcode error

    ⇒ **S3**   Always ASCII zero ('0')

    ⇒ **ETX**   End Of Text -- marks the end of the status block.

Instrument failures are usually accompanied by an error code on the reader display

ELx Status Mode Format:

This status string consists of 5 successive ASCII characters – a four-byte string representing a hexadecimal status code, and then ETX.

Items described in angle brackets (<>) are indicated by an ASCII digit replacing the last '0' character in the status code.

Items described in curly braces ({}) are indicated by an ASCII digit replacing the next-to-last '0' character in the status code.

Fatal errors indicate a hardware failure, also shown on the instrument display screen, and require recycling of instrument power.

Note – Errors listed below are common to all reader instruments, and may not all be applicable to any single given reader.

<u>*Fatal Errors*</u>

TCB NOT AVAIL ERR	"A100"	// task control block not available
READ NOT AVAIL ERR	"A200"	// read already in progress
NOT AVAIL ERR	"A300"	// <device> not available
CHECKSUM ERR	"A400"	// failed code checksum test on powerup
DR ALLOC ERR	"A500"	// DR steps alloc/free error <assay num)
DFLASH TIMEOUT ERR	"A600"	// data flash write timed out
DFLASH ERR	"A700"	// data flash readback didn't match write {test}<chip>
CFLASH TIMEOUT ERR	"A800"	// code flash write timed out
HEAP CORRUPTION ERR	"A900"	// memory allocation heap corrupted

<u>*Non-Fatal Errors*</u>

NO ERR	"0000"	// no errors detected
ABORT ERR	"0100"	// read function aborted
NO SENSOR ERR	"0200"	// <motor> didn't find opto-sensor transition
NO BEAM ERR	"0300"	// <motor> didn't find saturation transition
MOTOR VERIFY ERR	"0400"	// <motor> failed positional verify
SATURATION ERR	"0500"	// A/D signal saturated <test type>
FILTER GAIN ERR	"0600"	// <filter> gain out of range
NOISE TEST ERR	"0700"	// reader {channel} failed noise test
OFFSET TEST ERR	"0800"	// reader {channel} failed offset test
DARK RANGE ERR	"0900"	// read-time {channel}<filter> dark out of range
AIR RANGE ERR	"0A00"	// read-time {channel}<filter> air/blank out
ASSAY NUM ERR	"0B00"	// invalid <assay number>
PRINT TIMEOUT ERR	"0C00"	// printer timed out
CAL CHECKSUM ERR	"0D00"	// failed calibration checksum test
WAVE NOT FOUND ERR	"0E00"	// wavelength not found in table <read filter>
FILTER SIGNAL ERR	"0F00"	// {channel}<filter> signal out of range
CNFG DATA ERR	"1000"	// necessary configuration data missing
CNFG CHECKSUM ERR	"1100"	// failed configuration checksum test
CAL DATA ERR	"1200"	// necessary calibration data missing
MOTOR NOT HOMED ERR	"1300"	// <motor> not homed successfully
INCUBATOR FAILURE	"1500"	// incubator failure {error code}<zone(s)>
SC ASSAY DEF ERR	"1600"	// computer control assay definition error
KIN INTERVAL ERR	"1700"	// interval too short for selected options
KIN COUNT ERR	"1800"	// too many kinetic intervals
MALLOC ERR	"1900"	// malloc failed

STORE CURVE ERR	"1A00"	// store curve failure
GET CURVE ERR	"1B00"	// get curve failure
ATOD INIT ERR	"1C00"	// A/D calib STBY transition not detected
RESULTS DATA ERR	"1D00"	// results data error
CLOCK ERR	"1E00"	// error in clock communications
OVERLAP ERR	"1F00"	// bandpass overlap in filterset
BARCODE ERR	"2000"	// no valid barcode detected
INVALID PARAM ERR	"2100"	// invalid parameter value selected
PMT ERR	"2200"	// PMT test signal too high <test type>
LAMP ERR	"2300"	// lamp control failure <test type>
SENSOR POS ERR	"2400"	// test sensor position incorrect <motor>
FLASH MISS ERR	"2500"	// motor went by flash location too soon
XY LIMIT ERR	"2600"	// physical limit exceeded for area scan request
PANEL METHOD ERR	"2700"	// <assay> method doesn't match first panel assay
MOTOR TIMER ERR	"2800"	// <motor> timer not available

Test Type Codes *(lowest digit in returned error code)*

FAIL POWER 5V	'1'	// 5V power failed
FAIL POWER 24V	'2'	// 24V power failed

Motor Codes *(lowest digit in returned error code)*

Carrier X Axis	'0'	
Filter Wheel	'1'	
Robotic Door	'2'	('R' instruments only)

Incubator Codes *(second lowest digit in returned error code)*

Range Error	'0'
Thermistor Error	'1'
A/D Error	'2'

Affected zones are encoded in the lowest digit returned – one bit per zone.

Data Flash Codes *(2nd lowest digit in returned error code)*

Readback Error	'0'	// data read back didn't match data written
Copy Error	'1'	// final data readback didn't match original passed in

Error Codes

This appendix provides a list of Error Codes that may appear on the ELx808™, and their probable causes. If an error code appears on the display (or in Gen5™, KC4™, or KCjunior™), look up the code in the Troubleshooting tables in this appendix and attempt to solve the problem.

If you are unable to solve the problem, contact BioTek's Technical Assistance Center. See *Chapter 1* for contact information.

Glossary of Terms

➢ **Air Blank:** A full light reading through a filter with no plate in the light path.

➢ **Dark Current:** A reading taken with the light blocked to measure background light levels in the reading chamber. Also used as a measure of background electronic noise within the measurement circuit.

➢ **Gain:** An automatic electronic adjustment to the measurement circuit. The gain adjustment compensates for changing light levels or filter variations. For example, if the lamp output decreases slightly, the gain will increase to make up the difference.

➢ **Axis:** Refers to a motor that positions the filter wheel or plate carrier.

➢ **Offset:** A numerical limit, usually a range. For example, if the gain fails an offset test, it may be too high or too low.

Error Codes

Displayed Error	Potential Cause
ERROR 0200	Plate carrier did not find the home sensor
ERROR 0201	Filter wheel motor did not find the home sensor; filter wheel not installed or detached from hub.

Errors 0200 and 0201 indicate that a motor was not able to move to its "home" position as registered by feedback from an optical sensor.

PROBABLE CAUSE:

- Carrier movement may be limited so that the optical sensor cannot be interrupted. Check the carrier path for obstructions.

- Filter wheel not installed. The wheel is shipped in a separate envelope and may not be installed or may have broken free of the magnetic hub.

- Filter wheel movement may be impeded. Remove the wheel to confirm if a loose filter is impeding the movement of the wheel.

Displayed Error	Potential Cause
ERROR 0300	Carrier failed to find light beam
ERROR 0301	Filter wheel did not find home

Errors 0300 and 0301 indicate that a particular axis has moved to a point where the light beam from the optics is no longer detectable by the measurement electronics. This error is usually only seen during the service Auto Cal sequence.

PROBABLE CAUSE:

- Carrier: Lamp may be out or blocked by a faulty filter wheel. A loose belt or a loose motor pulley may cause the carrier to ignore movement instructions.

- Filter Wheel Motor: The filter wheel is missing or loose or a loose filter may be impeding filter wheel movement.

Displayed Error	Potential Cause
ERROR 0400	Carrier axis failed position verify
ERROR 0401	Filter Wheel motor failed position verify

Errors 0400 and 0401 indicate that an axis failed its Position Verify test. After moving a predefined number of steps from a home position, the motor should return to the home position in the proper amount of time and steps. If the axis moves back to its home position in the wrong amount of time or in too few or too many steps, the test fails.

PROBABLE CAUSE:

- Mechanical obstruction

- Carrier belt or pulley slipping

- Loose filter wheel hub

- Loose filter may be impeding filter wheel movement

- Defective motor drive circuit or 12-volt power problem

Displayed Error	Potential Cause
ERROR 0500, 0502…0506	Filter is not installed

Error 0500 indicates an empty filter location on the filter wheel, or that the filter wheel is not installed.

PROBABLE CAUSE:

- All filter locations must have either a filter or a filter blank (BTI PN 3122037) installed. The reader may also display this error if the filter wheel is not installed.

Displayed Error	Potential Cause
ERROR 0601 to 0606	FILTER #1 (to #6) Gain out of range

Errors 0601-0606 indicate that the gain for a specific filter is out of range. Gain out of range indicates that the lamp is out, or that a particular filter absorbs or transmits entirely incorrect light levels.

PROBABLE CAUSE :

- Lamp out. Check that the lamp is lit when the unit is ON. Replace lamp if necessary.

- A missing, defective, or misplaced filter or filter blank will cause this error. Observe that the filters in the wheel match the filters programmed in the software. Look for obvious physical defects in the filters.

Displayed Error	Potential Cause
ERROR 0700	Reader failed noise test

Error 0700 indicates significant variations in background electronic noise were detected when blocking the light and increasing the Gain to maximum.

PROBABLE CAUSE:

- Electrical noise penetrating the measurement chamber. The bottom and top shrouds are part of the electrical shielding. Check that the shrouds are installed and properly fastened.

- Ambient light leak. Check that the plate chamber door is closed to keep out ambient light.

- Missing or loose filter allowing unintended lamp light into the measurement chamber. Check filter wheel for empty positions.

- Internal electronic noise caused by a faulty Analog PCB or faulty internal grounding.

Displayed Error	Potential Cause
ERROR 0800	Reader failed offset test

Error 0800 indicates background electronic signal detected is outside of acceptable limits at maximum gain when blocking the light.

PROBABLE CAUSE:

- Electrical noise penetrating the measurement chamber. The bottom and top shrouds are part of the electrical shielding. Check that the shrouds are installed and properly fastened.

- Ambient light leak. Check that the plate chamber door is closed to keep out ambient light.

- Missing or loose filter allowing unintended lamp light into the measurement chamber. Check filter wheel for empty positions.

- Internal electronic noise caused by a faulty Analog PCB or faulty internal grounding.

Displayed Error	Potential Cause
ERROR 0900	Read time dark value out of range

Error 0900 indicates that the dark current value taken during the current read is significantly different than the same reading taken during the power-up self-check.

PROBABLE CAUSE:

- Read chamber door has opened or closed relative to when the unit was powered ON. Close door and reboot reader to correct.

- Electrical noise penetrating the measurement chamber. The bottom and top shrouds are part of the electrical shielding. Check that the shrouds are installed and properly fastened.

- Internal electronic noise caused by a faulty Analog PCB or faulty internal grounding.

Displayed Error	Potential Cause
ERROR 0A00, 0A01…0A05	Read time air blank out of range

Error 0A00 indicates that the blank (full signal) reading taken during the current read has changed significantly from the same reading taken during the power-up self-check.

PROBABLE CAUSE:

- Lamp has failed since power-up. Check that the lamp is lit and replace if necessary.

- Analog PCB failure.

Displayed Error	Potential Cause
ERROR 0C00	Printer time out error

Error 0C00 indicates that the time allotted for the instrument to make a valid connection to a printer has expired.

PROBABLE CAUSE:

- Printer out of paper, not connected, or not on-line.

Displayed Error	Potential Cause
ERROR A900	Memory Manager corruption detected. Power instrument off, then back on.
ERROR 0D00	Reader failed calibration checksum test. Internal software corruption.
ERROR 0E00, 0E01…0E05	Wavelength not detected in reader's filter table. Compare filters wavelengths to the filter table.
ERROR 0F00…0F86	Reader's filter or channel signal is out of specified range. Check for lamp failure, and/or replace lamp.
ERROR 1000	Required reader configuration data missing. Re-download software basecode and assays.
ERROR 1100	Failed configuration checksum test. Re-download software basecode and assays.
ERROR 1200	Calibration data missing.
ERROR 1300	Motor not homed. Autocal error, see error 0200.
ERROR 1400	Assay incubation Error. Assay requires incubation but the instrument does not appear to have an incubator.
ERROR 1500	Incubator failed to hold temperature within tolerances during the assay.
ERROR 1600	Computer control assay definition error. This error is used only for software development.
ERROR 1700	Kinetic interval too short for selected options.
ERROR 1800	Too many kinetic intervals selected.
ERROR 1900	Memory allocation failure. This error is used only for software development.
ERROR 2311…2386	Lamp unstable. Replace lamp or clean lamp contacts (page 5-8).

Report Format

The following are examples of reports that can be generated and/or printed from the ELx808™. For details on how to print these reports, refer to ***OUTPUT Options*** in ***Chapter 2***, ***Installation***.

In addition, an Assay List, Assay Definition, Map, and Result can be printed by choosing **Report** from the Main Menu screen.

Bio-Tek Instruments

Assay:ASSAY 01   Date:07/23/96   Lot: _____
                 Time:03:05:35AM Operator:
Wavelength:405   Temp: Plate ID:SONTA

COMMENTS
    OUT indicates an out of range value was read.

	1	2	3	4	5	6	7	8	9	10	11	12
A CALL		POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS
CalcOD	0.182	0.072	0.073	0.077	0.071	0.075	0.073	0.074	0.073	0.075	0.072	0.076
Well	STD1	SMP5	SMP13	SMP21	SMP29	SMP37	SMP45	SMP53	SMP61	SMP69	SMP77	SMP85
RSLT												
B CALL		POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS
CalcOD	0.529	0.076	0.084	0.085	0.096	0.073	0.073	0.069	0.072	0.070	0.070	0.076
Well	STD2	SMP6	SMP14	SMP22	SMP30	SMP38	SMP46	SMP54	SMP62	SMP70	SMP78	SMP86
RSLT	0.483											
C CALL		POS	NEG	NEG	NEG	POS	POS	POS	POS	POS	POS	POS
CalcOD	0.721	0.070	0.351	0.443	0.376	0.076	0.074	0.077	0.071	0.076	0.081	0.066
Well	STD3	SMP7	SMP15	SMP23	SMP31	SMP39	SMP47	SMP55	SMP63	SMP71	SMP79	SMP87
RSLT	0.686		0.294	0.391	0.321							
D CALL		POS	NEG	NEG	NEG	POS	POS	POS	POS	POS	POS	NEG
CalcOD	0.749	0.081	OUT	OUT	OUT	0.078	0.074	0.079	0.074	0.072	0.076	0.683
Well	STD4	SMP8	SMP16	SMP24	SMP32	SMP40	SMP48	SMP56	SMP64	SMP72	SMP80	SMP88
RSLT	0.715											0.646
E CALL	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS
CalcOD	0.068	0.070	0.093	0.101	0.081	0.078	0.078	0.074	0.074	0.071	0.074	0.072
Well	SMP1	SMP9	SMP17	SMP25	SMP33	SMP41	SMP49	SMP57	SMP65	SMP73	SMP81	SMP89
RSLT												
F CALL	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	NEG
CalcOD	0.083	0.069	0.075	0.070	0.072	0.079	0.082	0.085	0.074	0.069	0.078	0.671
Well	SMP2	SMP10	SMP18	SMP26	SMP34	SMP42	SMP50	SMP58	SMP66	SMP74	SMP82	SMP90
RSLT												0.632
G CALL	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS	POS
CalcOD	0.073	0.070	0.073	0.072	0.076	0.072	0.074	0.080	0.076	0.072	0.072	0.073
Well	SMP3	SMP11	SMP19	SMP27	SMP35	SMP43	SMP51	SMP59	SMP67	SMP75	SMP83	SMP91
RSLT												
H CALL	POS	POS	POS	POS	POS	POS	POS	POS	NEG	POS	POS	POS
CalcOD	0.072	0.075	0.073	0.074	0.081	0.069	0.076	0.069	0.738	0.076	0.075	0.072
Well	SMP4	SMP12	SMP20	SMP28	SMP36	SMP44	SMP52	SMP60	SMP68	SMP76	SMP84	SMP92
RSLT									0.704			

Figure D-1: Samples with Calls on Matrix Report

Bio-Tek Instruments

Assay:ASSAY 01   Date:07/23/96 Lot: _____
            Time:03:05:35AM Operator:
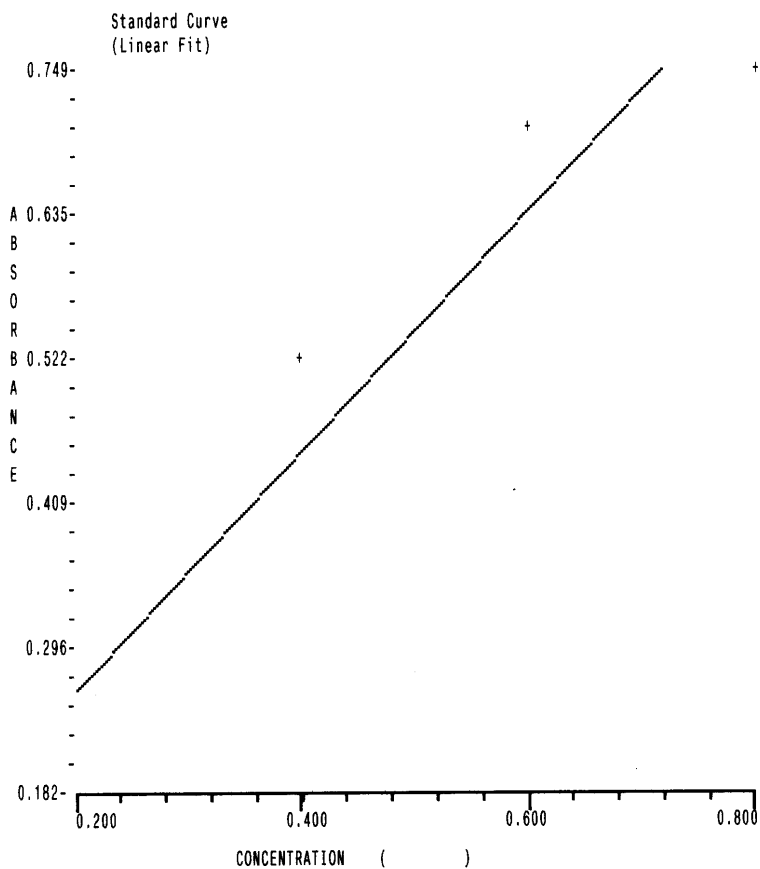Wavelength:405   Temp: Plate ID:SONTA

COMMENTS
  OUT indicates an out of range value was read.

Figure D-2: Curve Fit Report

Assay:ASSAY 01   Date:07/23/96   Lot: _____
                               Time:03:05:35AM   Operator:
Wavelength:405   Temp: Plate ID:SONTA

COMMENTS
      OUT indicates an out of range value was read.

Interpretation of Results

      GREYZONE=0.00   =0.000
      CUTOFF=.2   =0.200

      SMP : SMP<=(CUTOFF+(CUTOFF*GREYZONE))AND(SMP>=(CUTOFF-(CUTOFF*GREYZONE))) : EQUIV
      SMP : SMP>(CUTOFF+(CUTOFF*GREYZONE)) : NEG
      SMP : SMP<(CUTOFF-(CUTOFF*GREYZONE)) : POS

Well	ID	BlkdOD	CalcOD	Call	Pred Conc	RSLT	Std Dev	CV%	Notes:
A01	STD1	0.182	0.182		0.200				
B01	STD2	0.529	0.529		0.400	0.483			
C01	STD3	0.721	0.721		0.600	0.686			
D01	STD4	0.749	0.749		0.800	0.715			
E01	SMP0001	0.068	0.068	POS					
F01	SMP0002	0.083	0.083	POS					
G01	SMP0003	0.073	0.073	POS					
H01	SMP0004	0.072	0.072	POS					
A02	SMP0005	0.072	0.072	POS					
B02	SMP0006	0.076	0.076	POS					
C02	SMP0007	0.070	0.070	POS					
D02	SMP0008	0.081	0.081	POS					
E02	SMP0009	0.070	0.070	POS					
F02	SMP0010	0.069	0.069	POS					
G02	SMP0011	0.070	0.070	POS					
H02	SMP0012	0.075	0.075	POS					
A03	SMP0013	0.073	0.073	POS					

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Figure D-3: Samples with Calls on Column Report

Bio-Tek Instruments

Assay:ASSAY 01   Date:07/29/96   Lot: _____
                                   Time:09:55:19AM   Operator:
Wavelength:405   Temp: Plate ID:

COMMENTS
        OUT indicates an out of range value was read.

Interpretation of Results

        GREYZONE=0.25   =0.250
        CUTOFF=.1   =0.100

        SMP : SMP<=(CUTOFF+(CUTOFF*GREYZONE))AND(SMP>=(CUTOFF-(CUTOFF*GREYZONE))) : EQUIV
        SMP : SMP>(CUTOFF+(CUTOFF*GREYZONE)) : NEG
        SMP : SMP<(CUTOFF-(CUTOFF*GREYZONE)) : POS

Well ID	BlkdOD	CalcOD	Call	Pred Conc	RSLT	Std Dev	CV%	Notes:
A01 STD1	0.184	0.184						
B01 STD2	0.529	0.529						
C01 PC	0.723	0.723						
D01 NC	0.746	0.746						

End of Report

Figure D-4: Column Report without Samples

Bio-Tek Instruments

Panel

Date:07/31/96
Time:02:24:01PM
Temp:

Lot: _____
Operator:
Plate ID:

COMMENTS

Specimen	Assay	Well	BlkdOD	CalcOD	Call	RSLT	Std Dev	CV%	Notes:
SMP0001	ASSAY 01	F01	-.006	-.006					
	ASSAY 02	B05	0.000	0.000					
SMP0002	ASSAY 01	G01	0.002	0.002					
	ASSAY 02	C05	0.007	0.007					
SMP0003	ASSAY 01	H01	0.005	0.005					
	ASSAY 02	D05	0.004	0.004					
SMP0004	ASSAY 01	A02	0.002	0.002					
	ASSAY 02	E05	0.012	0.012					
SMP0005	ASSAY 01	B02	-.002	-.002					
	ASSAY 02	F05	0.010	0.010					
SMP0006	ASSAY 01	C02	0.005	0.005					
	ASSAY 02	G05	-.001	-.001					
SMP0007	ASSAY 01	D02	-.001	-.001					
	ASSAY 02	H05	0.002	0.002					
SMP0008	ASSAY 01	E02	0.009	0.009					
	ASSAY 02	A06	0.004	0.004					
SMP0009	ASSAY 01	F02	0.006	0.006					
	ASSAY 02	B06	0.007	0.007					
SMP0010	ASSAY 01	G02	-.006	-.006					
	ASSAY 02	C06	0.010	0.010					
SMP0011	ASSAY 01	H02	-.005	-.005					
	ASSAY 02	D06	0.004	0.004					
SMP0012	ASSAY 01	A03	-.00	-.00					
	ASSAY 02	E06	0.003	0.003					
SMP0013	ASSAY 01	B03	0.002	0.002					
	ASSAY 02	F06	-.001	-.001					
SMP0014	ASSAY 01	C03	-.005	-.005					
	ASSAY 02	G06	0.001	0.001					
SMP0015	ASSAY 01	D03	-.002	-.002					

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Figure D-5: Panel Report

```
NAME:                       ASSAY 55
READ TYPE:                  ENDPOINT
WAVELENGTH:                 450
BLANK MAP:                  AIR
AVERAGE STANDARDS?          NO
CONCN:                      STD1          6.2
                            STD2         12.5

CURVE-FIT TYPE:             LINEAR
EDIT STD OUTLIERS:          NONE
X/Y AXIS TYPE:              LIN
EXTRAPOLATE UNKNOWNS?       NO
```

```
    1      2     3     4     5     6     7     8     9    10    11    12
A  STD1   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
B  STD1   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
C  STD2   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
D  STD2   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
E  PC     SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
F  PC     SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
G  NC     SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
H  NC     SMP   SMP   SMP   SMP   SMP   SMP   SMP   SMP
```

Figure D-6: Assay Detail Report

```
Assay List
Version: 2.82
_____

 1    _Quick Read
 2    ASSAY 02
 3    ASSAY 03
 4    ASSAY 04
 5    ASSAY 05
 6    ASSAY 06
 7    ASSAY 07
 8    ASSAY 08
 9    ASSAY 09
10    ASSAY 10
11    ASSAY 11
12    ASSAY 12
13    ASSAY 13
14    ASSAY 14
15    ASSAY 15
16    ASSAY 16
17    ASSAY 17
18    ASSAY 18
19    ASSAY 19
20    ASSAY 20
21    ASSAY 21
22    ASSAY 22
23    ASSAY 23
24    ASSAY 24
25    ASSAY 25
26    ASSAY 26
27    ASSAY 27
28    ASSAY 28
29    ASSAY 29
30    ASSAY 30
31    ASSAY 31
32    ASSAY 32
33    ASSAY 33
34    ASSAY 34
35    ASSAY 35
36    ASSAY 36
37    ASSAY 37
38    ASSAY 38
39    ASSAY 39
40    ASSAY 40
41    ASSAY 41
42    ASSAY 42
43    ASSAY 43
44    ASSAY 44
45    ASSAY 45
46    ASSAY 46
47    ASSAY 47
48    ASSAY 48
49    ASSAY 49
50    ASSAY 50
51    ASSAY 51
52    ASSAY 52
53    ASSAY 53
54    ASSAY 54
55    ASSAY 55
```

Figure D-7: *Assay List*

Programming a New Assay

This appendix provides two examples of assay kit instructions and step-by-step directions for programming each assay. The appendix includes two sample assays: one with a ratio transformation calculation and a POS/NEG cutoff determination, and another with a standard curve.

(i) **Important!** The kit instructions are provided so that users can see how it is possible to translate the kit wording into an ELx808™ assay program. We are providing the following programming examples using fictitious *In-Vitro Diagnostic* (IVD) assays. These instructions are not intended to promote the use of the ELx808 with these or any other specific IVD assays, and they do not constitute a claim of IVD use in jurisdictions where there are additional regulatory requirements. For clarity, only the user menu choices from the reader screens are shown. Refer to *Chapter 3* for details.

(i) **Important!** The ELx808 user is responsible for programming the reader properly according to their specific kit instructions, and for verifying that the calculations are performed correctly. If you are not familiar with programming formulas onboard the ELx808, refer to "Formula Entry" in *Chapter 3* for further information.

This appendix contains the following sections:

Sample ANA Screen Enzyme Immunoassay Kit
(Transformation and Cutoff)

> **(i)** **Important!** The kit instructions are provided so that users can see how it is possible to translate the kit wording into an ELx808™ assay program. We are providing the following programming example using a fictitious *In-Vitro Diagnostic* (IVD) assay. These instructions are not intended to promote the use of the ELx808 with this or any other specific IVD assays, and they do not constitute a claim of IVD use in jurisdictions where there are additional regulatory requirements. For clarity, only the user menu choices from the reader screens are shown. Refer to ***Chapter 3*** for details.

Intended Use

This assay is designed for the in vitro screening of serum for the presence of specific IgG antinuclear antibodies (ANAs), to aid in diagnosing certain systemic rheumatic diseases. Sufficient materials are supplied to allow a maximum of 93 samples to be screened in single, with a positive, cutoff, and negative controls.

Background

Antinuclear antibodies occur in a large number of patients with systemic rheumatic diseases. These diseases are characterized by the presence of one or more ANAs.

Sera positive on this ANA kit should be tested for the specific autoantibodies indicative of the various systemic rheumatic diseases.

Principle of the Assay

Microwells are pre-coated with purified antigens. The prediluted controls and diluted patient samples are added to the wells, and autoantibodies recognizing one or a combination of antigens bind during the first incubation. After washing the wells to remove all unbound proteins, IgG conjugate is added. The conjugate binds to the captured autoantibody, and the excess unbound conjugate is removed by a further wash step. Substrate is added that causes a blue reaction, thereby exposing the bound conjugate, and producing an intensity proportional to the concentration of autoantibody in the sample. Phosphoric acid is added to each well to stop the reaction. This produces a yellow end-point color, which is read at 450 nm.

Materials

Materials Supplied

- **Instruction Leaflet:** Giving full assay details.

- **QC Certificate:** Indicating the expected performance of the batch.

- **ANA Coated Wells:** 12 x 8 well strips coated with purified antigens.

- **Type III Wash Buffer 20x Concentrate:** 1 bottle containing 50 ml of a concentrated buffer for washing the wells.

- **Type III Sample Diluent**: 2 bottles containing 50 ml of buffer for sample dilution. Ready to use.

- **ANA Positive Control:** 1 bottle containing 1.8 ml of diluted stabilized serum. Ready to use.

- **ANA Cutoff Control:** 1 bottle containing 1.8 ml of diluted stabilized serum. Ready to use.

- **ANA Negative Control:** 1 bottle containing 1.8 ml of diluted stabilized serum. Ready to use.

- **ANA Conjugate:** 1 bottle containing 12 ml of peroxidase labeled antibody to IgG. Colored red, ready to use.

- **TMB Substrate:** 1 bottle containing 14 ml TMB substrate. Ready to use.

- **Stop Solution:** 1 bottle containing 14 ml of 3M Phosphoric acid. Ready to use.

Additional Materials and Equipment – Not Supplied

- **Automatic Microplate Plate Washer:** This is recommended; however, plate washing can be performed manually.

- **Plate Reader:** Capable of measuring optical densities at 450 nm referenced on air.

- **Distilled or Deionized Water:** This should be of the highest quality available.

- **Calibrated Micropipettes:** For dispensing 1000, 100, and 10 µl.

- **Multichannel Pipette:** Recommended for dispensing 100 µl volumes of conjugate, substrate, and stop solution.

- **Glass/Plastic Tubes:** For sample dilution.

Quality Control and Results

Quality Control

For an assay to be valid, all the following criteria must be met:

- Cutoff as well as Positive and Negative controls must be included in each run.

- The OD of the cutoff and the ANA result of the Negative and Positive Controls should be in the ranges specified on the QC Certificate.

  For example: The absorbance of the Positive Control must be greater than 1.200 OD.

  The absorbance of the Negative Control must be less than 0.300 OD.

  Note: If the above criteria are not met, the assay is invalid and the test should be repeated.

Calculation of the Sample Results

Use the following formula to calculate the ANA result for each sample:

$$\frac{\text{Control or sample OD}}{\text{Cutoff control OD}} \quad x \quad 10 \quad = \quad \text{Control or sample value (U/ml)}$$

Expected Values

The normal range was determined on serum from 200 normal adult blood donors. The cutoff control has been set at a point equivalent to the upper normal limit, or a cutoff level of 10 U/ml.

The ranges are provided as a guide only. ELISA assays are very sensitive and capable of detecting small differences in sample populations. It is recommended that each laboratory determine its own normal range, based on the population techniques and equipment employed.

ANA Result	Interpretation
< 10.0	Negative
> 10.0	Positive

Programming the ANA Screen Enzyme Immunoassay Kit (Transformation and Cutoff)

(i) **Important!** The ELx808™ user is responsible for programming the reader properly according to their specific kit instructions, and for verifying that the calculations are performed correctly. If you are not familiar with programming formulas onboard the ELx808, refer to "Formula Entry" in *Chapter 3*.

From the Main Menu, press DEFINE. Enter the assay number and edit the name if desired. At the DEFINE menu:

STEP	COMMENTS
1. To program the reading method, press:	
METHOD:	
READ TYPE (Endpoint, Kinetic, or Scan): Endpoint	
DELAY FIRST READ:	
IF INCUBATOR MODEL:	
- INCUBATION TEMP (Ambient or Temperature):	
WAVELENGTH (Single or Dual): Single	
MEASURE (Wavelength[s] to use): 450	
SHAKE (First, Every, or None):	
2. To program the plate map, press:	
MAP	
AUTO	"Auto" mapping is normally preferred, because it fills in the well IDs logically and automatically after determining which direction to map and how many wells to fill.
DOWN	Maps the wells down the column
DOWN	Locates replicates in a vertical orientation down the column
A01	Begins mapping at well location A01
BLANK MAP: AIR	Choose to blank on "AIR" if no blank wells are required

STEP	COMMENTS
NUMBER STDS: 00	
REUSE STANDARD CURVE?	Can only be programmed in assay positions 31-55, and cannot be reused on panels.
NUMBER CTLS: 03	
CONTROL 1: PC	
CONTROL 2: CTL1	Suggested choice for the cutoff control
CONTROL 3: NC	
NUMBER OF REPLICATES	
PC: 01	
CTL1: 03	
NC: 01	
SAMPLES: 91	User-defined; recommendation is to fill the entire plate
SAMPLE REPLICATES: 01	
Use the MAP and MATH keys to create the control validation formula:	
FORMULA	
VAL	
CONTROL:	When defining control validation formulas, use "PC" to indicate the criterion for each of the PC replicates, and "PC;x" to indicate the average of the Positive Control replicates.
PC;x > 1.200	
NO. OF REPLICATES: 01	
NC;x < 0.300	
NO. OF REPLICATES: 01	

STEP	COMMENTS
3. To create the plate transformation for CALC, divide the ANA result by a cutoff standard.	
FORMULA	
*MORE	
TRANS-VAR	
SCOPE VARIABLE (SMP or OD)	Select OD to advance to the formula definition screen and define the transformation variable (Tvar)
FORMULA	
CTL1;X	Defining TRANS VAR = CTL1;X isolates the OD value for CTL1;X for use in transformation
TRANS:	
FORMULA: (OD/TVAR)*10	Converts all OD values on plate to "ANA Result" per kit insert instructions
4. To define a cutoff formula for Positive and Negative calls:	
FORMULA	
CUTOFF: 10.0	
GREYZONE: 00%	
SAMPLE > CUTOFF: POS	

Sample Anticardiolipin IgG Enzyme Immunoassay Kit (Standard Curve and Cutoff)

(i) **Important!** The kit instructions are provided so that users can see how it is possible to translate the kit wording into an ELx808™ assay program. We are providing the following programming example using a fictitious *In-Vitro Diagnostic* (IVD) assay. These instructions are not intended to promote the use of the ELx808 with this or any other specific IVD assays, and they do not constitute a claim of IVD use in jurisdictions where there are additional regulatory requirements. For clarity, only the user menu choices from the reader screens are shown. Refer to *Chapter 3* for details.

Intended Use

This assay is intended for the in vitro measurement of IgG anticardiolipin antibodies in serum, as an aid in the diagnosis of antiphospholipid syndrome (APS).

Sufficient materials are supplied to allow a maximum of 41 samples to be tested in duplicate or 89 in single, with a standard curve and positive and negative controls.

Background

Anticardiolipin antibodies are found in a wide range of conditions either transiently, in some infectious diseases, or more persistently in autoimmune diseases such as systemic lupus erythematosus (SLE) and antiphospholipid syndrome (APS). Anticardiolipin antibodies have also been associated with a range of clinical conditions including fetal loss, endocarditis, stroke, heart attack and autoimmune haemolytic.

Principle of the Assay

Microwells are pre-coated with cardiolipin and cofactor. Standards, controls, and patient samples are added to the wells, and autoantibodies recognizing cardiolipin bind during the first incubation. After washing the wells to remove all unbound proteins, conjugate is added. The conjugate binds to the captured antibody, and the excess unbound conjugate is removed by a further wash step. Substrate is added that causes a blue reaction, thereby exposing the bound conjugate and producing an intensity proportional to the concentration of autoantibody in the sample. Phosphoric acid is added to each well to stop the reaction. This produces a yellow end-point color, which is read at 450 nm.

Materials

Materials Supplied

- *Instruction Leaflet:* Giving full assay details.

- *QC Certificate:* Indicating the expected performance of the batch.

- *Cardiolipin Coated Wells:* 12 break-apart 8-well strips coated with bovine cardiolipin antigen. The plate is packaged in a re-sealable foil bag containing two desiccant pouches.

- *Type II Sample Diluent:* 2 bottles containing 50 ml of buffer for sample dilution. Colored yellow, ready to use.

- *Type II Wash Buffer (20x Concentrate):* 1 bottle containing 50 ml of a 20-fold concentrated buffer for washing the wells.

- *Cardiolipin IgG Standards:* 5 bottles each containing 1.2 ml of diluted serum, with the following concentrations of anticardiolipin autoantibody: 100, 50, 25, 12.5, 6.25 GPL U/ml. Ready to use.

- The standard set is calibrated against the Louisville APL reference preparation (see *Assay Calibration,* page E-11).

- *Cardiolipin IgG Positive Control:* 1 bottle containing 1.2 ml of diluted serum. The expected value is given on the QC certificate. Ready to use.

- *Cardiolipin Negative Control:* 1 bottle containing 1.2 ml of diluted serum. The expected value is given on the QC certificate. Ready to use.

- *Cardiolipin IgG Conjugate:* 1 bottle containing 12 ml of purified peroxidase labeled antibody. Colored red, ready to use.

- *TMB Substrate:* 1 bottle containing 14 ml TMB substrate. Ready to use.

- *Stop Solution:* 1 bottle containing 14 ml of 3M phosphoric acid. Ready to use.

Additional Materials and Equipment – Not Supplied

- *Automatic Microplate Plate Washer:* This is recommended; however, plate washing can be performed manually.

- *Plate Reader:* Capable of measuring optical densities at 450 nm referenced on air.

- *Distilled or Deionized Water:* This should be of the highest quality available.

- *Calibrated Micropipettes:* For **dispensing** 1000, 100, and 10 µl.

- *Multichannel Pipette:* Recommended for dispensing 100 µl volumes of conjugate, substrate, and stop solution.

- *Glass/Plastic Tubes:* For sample dilution.

Quality Control and Results

Quality Control

For an assay to be valid, all the following criteria must be met:

- Standards and the positive and negative controls must be included in each run.

- The values obtained for all the controls should be in the ranges specified on the QC Certificate.

- The curve shape should be similar to the standard curve, shown on the QC Certificate.

  If the above criteria are not met, the assay is invalid and the test should be repeated.

Calculate Mean Optical Densities (for assays run in duplicate only)

For each standard, control and sample calculate the mean OD of the duplicate readings. The user must verify that the percentage coefficient of variation (%CV) for each duplicate OD is less than 15.0%.

Plot Calibration Curve

The calibration curve can be plotted either automatically or manually as follows by plotting the anticardiolipin autoantibody concentration on the log scale against the OD on the linear scale for each calibrator:

- Automatic - Use appropriately validated software, and the curve fit that best fits the data.

- Manual - Using log/linear graph paper, draw a smooth curve through the points (not a straight line or point to point).

Treatment of Anomalous Points

If any one point does not lie on the curve, it can be removed. If the absence of this point means that the curve has a shape dissimilar to that of the sample calibration curve, or more than one point appears to be anomalous, then the assay should be repeated.

Calculation of Autoantibody Levels in Controls and Samples

Read the level of the anticardiolipin autoantibody in the controls and diluted samples directly from the calibration curve. The control values should fall within the range given on the QC Certificate.

Note: The standard values have been adjusted by a factor of 100 to account for a 1:100 sample dilution. No further correction is required.

Assay Calibration

The assays are calibrated against the Louisville reference LAPL-GM-100. One GPL unit is defined as the cardiolipin binding activity of 1 µg/ml of an affinity purified IgG anticardiolipin preparation from a standard serum.

The Louisville reference center recommends the following positive discrimination criteria according to the recommendation of the 2nd International Anticardiolipin Workshop.

Criteria Range	(GPLU/ml)
High Positive	> 80
Medium Positive	≥ 20-80
Low Positive	≥ 10, < 20

Results Interpretation

The association between low positive levels of anticardiolipin antibodies and clinical findings is unclear.

Normal population studies indicate that there is a higher prevalence of IgM positives in the normal population than IgG, 9.4% and 6.5%, respectively. In normal pregnancy, the levels are higher still at 17.0% (IgM) and 10.6% (IgG).

Expected Values

The normal range was determined on serum from 102 normal adult blood donors. The ranges below are provided as a guide only. ELISA assays are very sensitive and capable of detecting small differences in sample populations. It is recommended that each laboratory determine its own normal range, based on the population techniques and equipment employed.

IgG Anticardiolipin	
< 11 GPL U/ml	Negative result
> 11 GPL U/ml	Positive result

Programming the Anticardiolipin IgG Enzyme Immunoassay Kit (Standard Curve and Cutoff)

Important! The ELx808™ user is responsible for programming the reader properly according to their specific kit instructions, and for verifying that the calculations are performed correctly. If you are not familiar with programming formulas onboard the ELx808, refer to "Formula Entry" in *Chapter 3*.

From the Main Menu, press DEFINE. Enter the assay number and edit the name if desired. At the define menu, follow the steps below:

STEP	COMMENTS
1. To program the reading method, press:	

METHOD:

READ TYPE: Endpoint

DELAY FIRST READ:

IF INCUBATOR MODEL:

- INCUBATION TEMP (Ambient or Temperature):

DELAY FIRST READ:

WAVELENGTH (Single or Dual): Single

MEASURE (Wavelength[s] to use): 450

SHAKE (First, Every, or None):

2. To program the plate map, press:

MAP

AUTO

DOWN

DOWN

A01

BLANK MAP: AIR

NUMBER STDS: 05

NUMBER STD REPLICATES: 01

CONCENTRATIONS:
      STD1: 6.25
      STD2: 12.5
      STD3: 25
      STD4: 50
      STD5:100

STEP	COMMENTS
REUSE STANDARD CURVE?	
NUMBER CTLS: 02	
CONTROL 1: PC	
CONTROL 2: NC	
NUMBER OF REPLICATES	
PC: 01	
NC: 01	
SAMPLES: 89	
SAMPLE REPLICATES: 01	

3. To define a cutoff formula for Positive and Negative calls:

   Kit instructions specify that samples with concentration values greater than 11 should appear as positive. The ELx808 software calculates the cutoff based on absorbance value or transformed value (see the previous example) and cannot calculate based on concentration. The technician must make the positive or negative determination visually, based on the calculated concentration results.

4. Curve: 4P

   As a general guideline, choose "linear" if you expect a straight line result. Choose "4P" for all others, unless otherwise specified by the kit instructions.

   The reader will automatically calculate the concentrations of the samples when the assay is run.

Adjusting the Power Input Voltage Setting

This appendix provides instructions for adjusting the power input voltage setting in ELx808™ instruments that are equipped with the internal four-voltage range power input module.

This appendix contains the following sections:

Overview

Previous versions of the ELx808™ (models manufactured before December 2005) have an internal, four-voltage range power input module, instead of an external power supply. Located on the right side of the ELx808, the input module can be adjusted for a 100, 120, 230, or 240 V~ voltage setting.

The voltage setting that is currently selected can be determined by observing which of the four possible voltage indicator holes has a white peg in it. Check the peg now to make sure the setting is appropriate for your location.

If you need to adjust the setting, first determine which type of power input module is installed in the reader:

- The **newer style module** has a cover that *can swing back toward the reader*, when opened with small needle-nose pliers. If your reader has this module, follow the instructions on pages F-3 through F-6 for adjusting the power input voltage setting and reconfiguring/replacing the fuses.

- The **older style module** has a cover that *can be detached from the reader* when opened with a small flat-blade screwdriver. If your reader has this module, follow the instructions on pages F-7 through F-9 for adjusting the power input voltage setting and reconfiguring/replacing the fuses.

Adjust the Power Input Voltage Setting *(Newer-Style Module)*

(i)   **Important!** Turn off and unplug the reader before adjusting the voltage setting.

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1. Unplug the reader and remove the power cord.

2. Insert a small pair of **needle-nose pliers** into the two holes on the cover of the power input module. Lift the cover open and swing it back toward the power cord socket (see *Figure F-1* below).

3. As shown below, a small voltage selector card is located to the right of the fuse holder. Using the pliers, carefully pull this card straight out of the compartment.
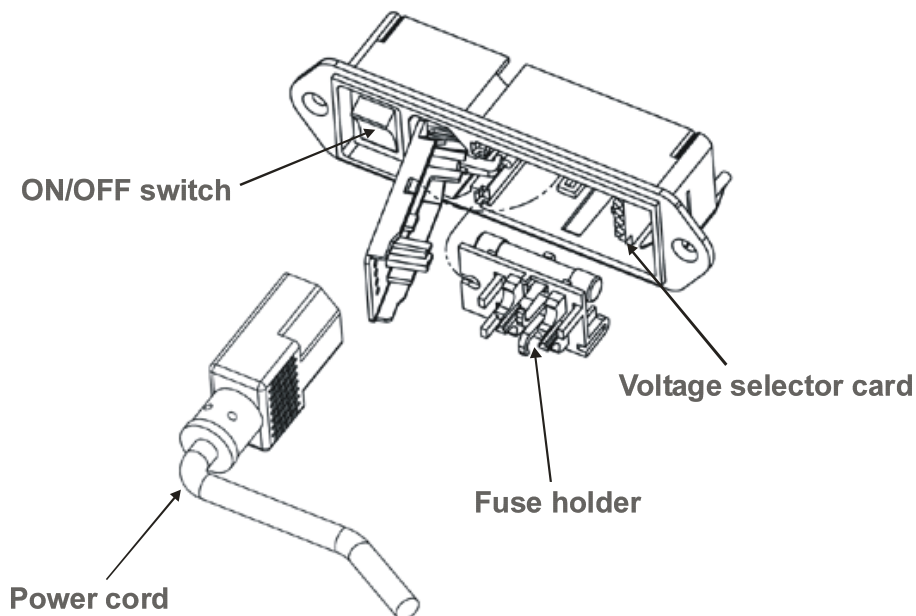

ON/OFF switch

Voltage selector card

Fuse holder

Power cord

Figure F-1: Power input module; accessing the voltage selector card and fuse holder

4. As shown in *Figure F-2* below, there are four different voltage range positions. Rotate the white plastic indicator peg around the card so that it fits into the correct groove for the desired voltage.

    Note: Point the peg upward so that the text matching the desired voltage range is readable at the **bottom** of the card, as shown below.

White plastic indicator peg should point upward.

This side (showing text) should face the ON/OFF switch (voltage range should be on the bottom).
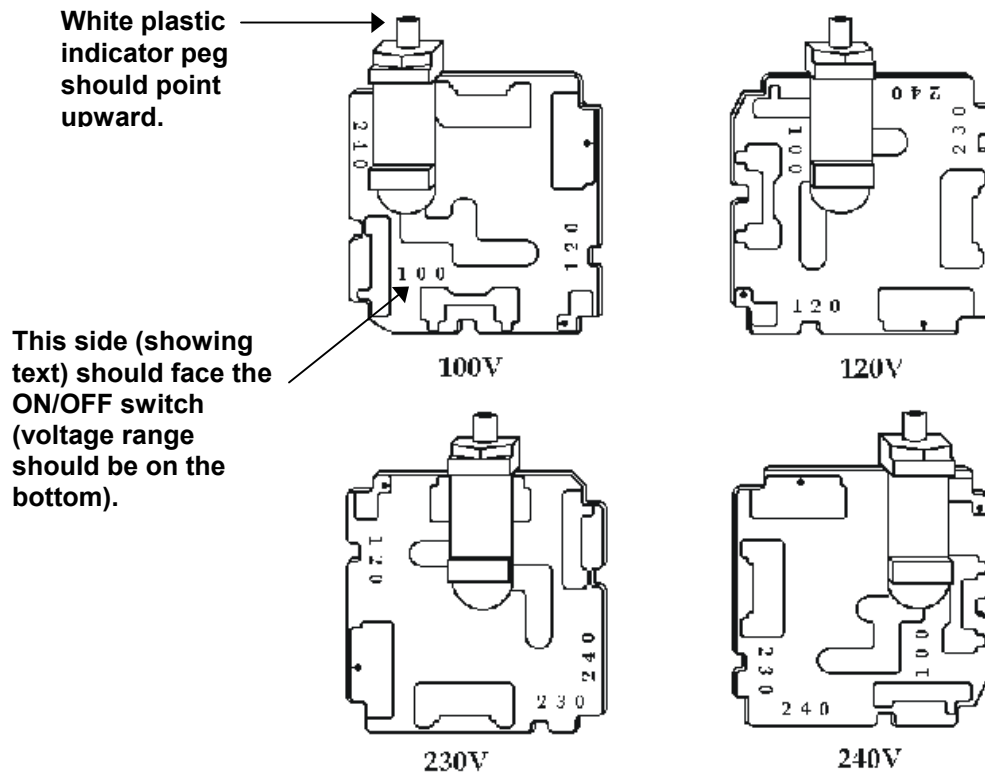
100V 120V

230V 240V

Figure F-2: Voltage selector card input range positions

5. Reinsert the voltage selector card into the power input module so that the text side faces the ON/OFF switch, and the desired voltage range is on the bottom of the card (see above).

6. Reconfigure or replace fuses as needed (see next page).

7. Replace the cover, and verify that the white peg lines up with the correct voltage indicator hole.

ⓘ   **Important!** The peg indicating the intended voltage should protrude through the cover. Do not power up the instrument until the voltage range to be used is indicated correctly by the peg.

Reconfigure or Replace Fuses *(Newer-Style Module)*

(i)   **Important!** Turn off and unplug the reader before reconfiguring or replacing the fuses.

Either USA *or* International style fuses are installed in the fuse holder that is located inside the power input module. The reader's fuses are configured at the factory prior to shipping. Use the procedure described below if you need to reconfigure or replace fuses.

Note: A failed fuse is usually an indication of another problem that a new fuse is not likely to remedy. Contact BioTek's Technical Assistance Center if the fuse replacement fails to resolve the problem. See *Chapter 1* for contact information.

The fuse holder can have either one of two fuse configurations:

- 100/120 V (USA): A fused hot AG 1.5 A, 250 V Slo Blow (PN 46024), or

- 230/240 V (International): Both hot and neutral fused with a 5 x 20 mm, 0.63 A, 250 V Slo Blow (PN 46038).

To reconfigure fuses:

1. Turn off and unplug the reader.
2. Insert the needle-nose pliers into the two holes on the cover of the power input module. Lift the cover open and swing it back toward the power cord socket.
3. Using the pliers, gently pull the fuse holder out of the power input module and turn it over.

Changing to the 100/120 V Configuration:	• Remove the two 0.63 A, 250 V fuses.
	• Install one AG 1.5 A, 250 V fuse (PN 46024) on the fuse holder on the side opposite the original 0.63 A fuse location.
	• Reinstall the fuse holder with the fuse toward the back of the power input module.
	• Replace the cover, and verify that the white plastic indicator peg lines up with the correct voltage indicator hole.
Changing to the 230/240 V Configuration:	• Remove the single AG 1.5 A, 250 V fuse.
	• Install two 0.63 A, 250 V fuses (PN 46038) on the fuse holder on the side opposite the original AG 1.5 A fuse location.
	• Reinstall the fuse holder with the fuses toward the back of the power input module.
	• Replace the cover, and verify that the white plastic indicator peg lines up with the correct voltage indicator hole.

To replace defective fuses:

1. Follow steps 1-3 on the previous page to access the fuse holder.

2. Remove the defective fuse and replace it with the correct new one.

3. Reinstall the fuse holder with the fuse toward the back of the power input module.

4. Replace the cover, and verify that the white plastic indicator peg lines up with the correct voltage indicator hole.

Adjust the Power Input Voltage Setting *(Older-Style Module)*

(i) **Important!** Turn off and unplug the reader before adjusting the voltage setting.

(i) **Important!** Be careful when removing the cover from the power input module: the fuse holder is attached to the inside of the cover.

Refer to *Figure F-3* on page F-8 for the following instructions:

1. Unplug the reader and remove the power cord.

2. Insert a small **flat-blade screwdriver** into the access hole between the power cord inlet and the power input module cover. Carefully pry the cover off the module. Note that the fuse holder is attached to the inside of the cover.

3. A small voltage selector card is located on the right side of the power input module. Using a small pair of **needle-nose pliers**, carefully pull this card straight out of the module.

4. Text printed on the card identifies the four different voltage positions. Each voltage number has an arrow printed next to it. Rotate the white plastic indicator peg so that it fits into the groove on the card that represents the desired voltage; the peg should point in the *opposite direction* from the arrow printed on the board.

5. Orient the card so that the peg is facing out and the text on the card is facing the ON/OFF switch. The arrow should be pointing into the power input module. Slide the card all the way back into the input module.

6. Reconfigure or replace fuses in the fuse holder, as needed (see the instructions on page F-9).

7. Replace the cover on the power input module and verify that the white peg lines up with the correct voltage indicator hole.

(i) **Important!** The peg indicating the intended voltage should protrude through the cover. Do not power up the instrument until the voltage range to be used is indicated correctly by the peg.
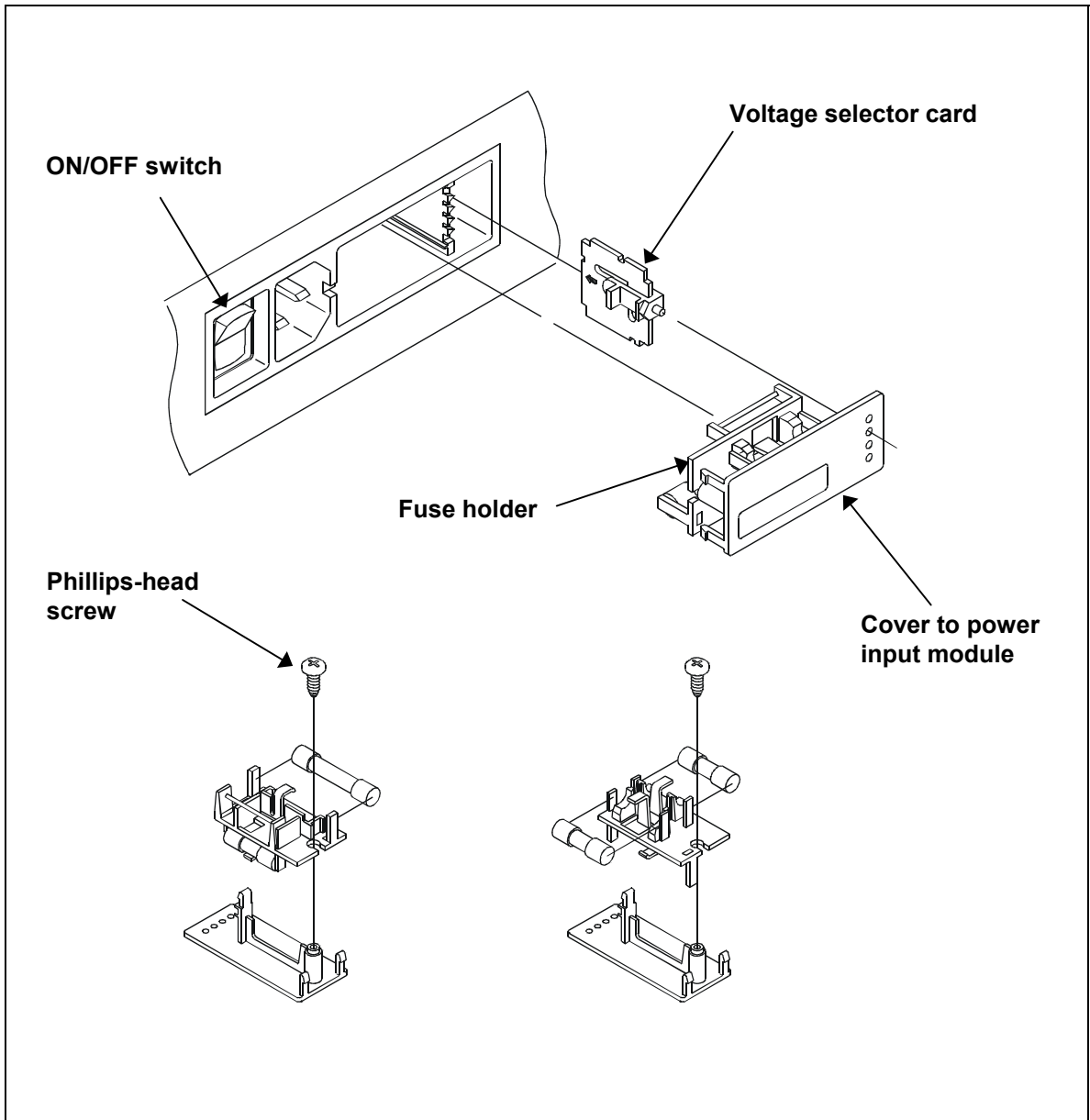
Voltage selector card

ON/OFF switch

Fuse holder

Phillips-head screw

Cover to power input module

Figure F-3: *Power input module; accessing the voltage selector card and fuse holder*

Reconfigure or Replace the Fuses *(Older-Style Module)*

(i) **Important!** Turn off and unplug the reader before reconfiguring or replacing the fuses.

Both USA *and* International style fuses are installed in the fuse holder that is attached to the inside of the power input module's cover. The reader's fuses are configured at the factory prior to shipping. Use the procedure below if you need to reconfigure or replace fuses.

Note: A failed fuse is usually an indication of another problem that a new fuse is not likely to remedy. Contact BioTek's Technical Assistance Center if the fuse replacement fails to resolve the problem. See *Chapter 1* for contact information.

The fuse holder has two fuse configurations:

- 100/120 V (USA): A fused hot AG 1.5 A, 250 V Slo Blow (PN 46024), and

- 230/240 V (International): Both hot and neutral fused with a 5 x 20 mm, 0.63 A, 250 V Slo Blow (PN 46038).

The fuse configuration (U.S. or International) that is currently being used by the reader is determined by which fusing network is facing the inside of the power input module.

To reconfigure fuses:

1. Turn off and unplug the reader.
2. Insert a **small flat-blade screwdriver** into the access hole between the power cord inlet and the power input module cover. Carefully pry the cover off the module.
3. Use the screwdriver to remove the Phillips-head screw that anchors the fuse holder to the inside of the cover. See *Figure F-3*.
4. Turn the fuse holder over.
5. Reattach the fuse holder to the cover with the screw.
6. Replace the cover, and verify that the white plastic indicator peg lines up with the correct voltage indicator hole.

To replace defective fuses:

1. Repeat steps 1 through 3 above to access the fuse holder.
2. Remove the defective fuse and replace it with the correct new one. Do *not* turn the fuse holder over, or you will change the configuration.
3. Reattach the fuse holder to the cover with the screw.
4. Replace the cover, and verify that the white plastic indicator peg lines up with the correct voltage indicator hole.