

# AssayMax<sup>™</sup>

# Human Albumin ELISA Kit

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For any questions regarding troubleshooting or performing the assay, please contact our support team at <a href="mailto:support@assaypro.com">support@assaypro.com</a>.

Thank you for choosing Assaypro.

## **Assay Summary**

**Step 1**. Add 50  $\mu$ l of Standard or Sample per well. Incubate 1 hour.

**Step 2.** Wash, then add 50  $\mu$ l of Biotinylated Antibody per well. Incubate 30 minutes.

Step 3. Wash, then add 50  $\mu$ l of SP Conjugate per well. Incubate 30 minutes.

**Step 4.** Wash, then add 50  $\mu$ l of Chromogen Substrate per well. Incubate 20 minutes.

**Step 5.** Add 50  $\mu$ l of Stop Solution per well. Read at 450 nm immediately.

## **Symbol Key**



Consult instructions for use.

# **Assay Template**

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## **Human Albumin ELISA Kit**

Catalog No. EA3201-1

Sample insert for reference use only

#### Introduction

Albumin, a serum hepatic protein, is the most abundant protein in serum. It contributes to the maintenance of oncotic pressure as well as the transport of hydrophobic molecules (1).

#### Principle of the Assay

The AssayMax Human Albumin ELISA (Enzyme-Linked Immunosorbent Assay) kit is designed for detection of human albumin in **urine**, **saliva**, **milk**, **CSF**, **and cell culture samples**. This assay employs a quantitative **sandwich enzyme immunoassay** technique that measures human albumin in less than 3 hours. A polyclonal antibody specific for human albumin has been pre-coated onto a 96-well microplate with removable strips. Albumin in standards and samples is sandwiched by the immobilized polyclonal antibody and biotinylated polyclonal antibody specific for human albumin, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

## **Caution and Warning**

- This product is for Research Use Only and is Not For Use In Diagnostic Procedures.
- Prepare all reagents (working diluent buffer, wash buffer, standard, biotinylated antibody, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this insert. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated antibody vial before opening and using contents.
- The Stop Solution is an acidic solution.
- The kit should not be used beyond the expiration date.

#### Reagents

- Human Albumin Microplate: A 96-well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against human albumin.
- Sealing Tapes: Each kit contains 3 precut, pressure sensitive sealing tapes that can be cut to fit the format of the individual assay.
- Human Albumin Standard: Human albumin in a buffered protein base (360 ng, lyophilized).
- Biotinylated Human Albumin Antibody (60x): A 60-fold concentrated biotinylated polyclonal antibody against human albumin (100 µl).
- MIX Diluent Concentrate (10x): A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 μl).
- Chromogen Substrate: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- Stop Solution: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

#### **Storage Condition**

- Upon arrival, immediately store components of the kit at recommended temperatures up to the expiration date.
- Store SP Conjugate and Biotinylated Antibody at -20°C.
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C.
- Unused microplate wells may be returned to the foil pouch with the desiccant packs and resealed. May be stored for up to 30 days in a vacuum desiccator.
- Diluent (1x) may be stored for up to 30 days at 2-8°C.
- Store Standard at 2-8°C before reconstituting with Diluent and at -20°C after reconstituting with Diluent.

## **Other Supplies Required**

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 μl, 20-200 μl, 200-1000 μl, and multiple channel).
- Deionized or distilled reagent grade water.

#### Sample Collection, Preparation, and Storage

- Cell Culture Supernatants: Centrifuge cell culture media at 3000 x g for 10 minutes to remove debris. Collect supernatants and assay. The samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- Urine: Collect urine using sample pot. Centrifuge samples at 800 x g for 10 minutes. Dilute urine samples 1:200 into MIX Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- Saliva: Collect saliva using sample tube. Centrifuge samples at 800 x g for 10 minutes. Dilute saliva samples 1:200 into MIX Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- Milk: Collect milk using sample tube. Centrifuge samples at 800 x g for 10 minutes. Dilute milk samples 1:6000 into MIX Diluent and assay. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- **CSF:** Collect cerebrospinal fluid (CSF) using sample pot. Centrifuge samples at 3000 x g for 10 minutes. Dilute samples 1:6000 into MIX Diluent and assay. The undiluted samples can be stored at -80°C for up to 3 months. Avoid repeated freeze-thaw cycles.

#### Refer to Sample Dilution Guidelines below for further instruction.

|       | Guidelines for Dilutions of 1:100 or Greater                                   |    |   |  |  |
|-------|--|----|---|--|--|
|       | (for reference only; please follow the insert for specific dilution suggested) |    |   |  |  |
| 1:100 |  |    | 1:10000                                 |  |  |
|       |  |    |   |  |  |
| A)    | 4 ul sample: 396 μl buffer(100x)   | A) | 4 μl sample : 396 μl buffer (100x)      |  |  |
|       | = 100 fold dilution  | B) | 4 μl of A : 396 μl buffer (100x)        |  |  |
|       |  |    | = 10000 fold dilution                   |  |  |
|       | Assuming the needed volume is less than  |    | Assuming the needed volume is less than |  |  |
|       | or equal to 400 μl.  |    | or equal to 400 μl.                     |  |  |
|       | 1:1000   |    | 1:100000                                |  |  |
|       |  |    |   |  |  |
| A)    | 4 μl sample : 396 μl buffer (100x)   | A) | 4 μl sample : 396 μl buffer (100x)      |  |  |
| B)    | 24 μl of A : 216 μl buffer (10x)   | B) | 4 μl of A : 396 μl buffer (100x)        |  |  |
|       | = 1000 fold dilution   | C) | 24 μl of B : 216 μl buffer (10x)        |  |  |
|       |  | ,  | = 100000 fold dilution                  |  |  |
|       | Assuming the needed volume is less than  |    | Assuming the needed volume is less than |  |  |
|       | or equal to 240 μl.  |    | or equal to 240 μl.                     |  |  |

#### **Reagent Preparation**

 Freshly dilute all reagents and bring all reagents to room temperature before use.

- MIX Diluent Concentrate (10x): If crystals have formed in the
  concentrate, mix gently until the crystals have completely dissolved.
   Dilute the MIX Diluent Concentrate 1:10 with reagent grade water. Store
  for up to 30 days at 2-8°C.
- Standard Curve: Reconstitute the 360 ng of Human Albumin Standard with 1.8 ml of MIX Diluent to generate a 200 ng/ml standard stock solution. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the standard stock solution (200 ng/ml) 1:2 with MIX Diluent to produce 100, 50, 25, 12.5, 6.25, and 3.125 ng/ml solutions. MIX Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C and used within 30 days.

| Standard<br>Point | Dilution                       | [Human Albumin]<br>(ng/ml) |
|-------------------|--------------------------------|----------------------------|
| P1                | 1 part Standard (200 ng/ml)    | 200.0                      |
| P2                | 1 part P1 + 1 part MIX Diluent | 100.0                      |
| Р3                | 1 part P2 + 1 part MIX Diluent | 50.00                      |
| P4                | 1 part P3 + 1 part MIX Diluent | 25.00                      |
| P5                | 1 part P4 + 1 part MIX Diluent | 12.50                      |
| P6                | 1 part P5 + 1 part MIX Diluent | 6.250                      |
| P7                | 1 part P6 + 1 part MIX Diluent | 3.125                      |
| P8                | MIX Diluent                    | 0.000                      |

- Biotinylated Human Albumin Antibody (60x): Spin down the antibody briefly and dilute the desired amount of the antibody 1:60 with MIX Diluent. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
   Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- SP Conjugate (100x): Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with MIX Diluent. Any remaining solution should be frozen at -20°C.

## **Assay Procedure**

- Prepare all reagents, standard solutions, and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-25°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccants inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.

- Add 50 µl of Human Albumin Standard or sample per well. Cover wells with a sealing tape and incubate for 1 hour. Start the timer after the last addition.
- Wash five times with 200 µl of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 µl of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
- Add 50 µl of Biotinylated Human Albumin Antibody to each well and incubate for 30 minutes.
- Wash the microplate as described above.
- Add 50 µl of Streptavidin-Peroxidase Conjugate to each well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.
- Add 50 µl of Chromogen Substrate per well and incubate for 20 minutes or till the optimal blue color density develops. Gently tap plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50  $\mu$ l of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm immediately. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections.
   Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

## **Data Analysis**

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance (OD) on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

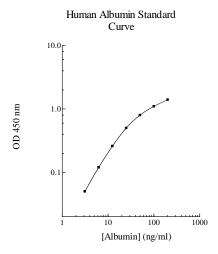
#### **Typical Data**

The typical data is provided for reference only. Individual laboratory
means may vary from the values listed. Variations between laboratories
may be caused by technique differences.

| Standard Point | ng/ml            | OD    | Average OD |
|----------------|------------------|-------|------------|
| P1             | 200.0            | 2.109 | 2.115      |
| LI             | 200.0            | 2.121 | 2.113      |
| P2             | 100.0            | 1.716 | 1.678      |
| ΓZ             | 100.0            | 1.641 | 1.076      |
| P3             | 50.00            | 1.231 | 1.261      |
| FJ             | 30.00            | 1.290 | 1.201      |
| P4             | 25.00            | 0.811 | 0.797      |
| 1 7            | 25.00            | 0.784 | 0.757      |
| DE             | 12.50            | 0.444 | 0.453      |
| P5             | 12.50            | 0.461 | 0.453      |
| P6             | P6 6.250         |       | 0.269      |
| PO             | 6.230            | 0.264 | 0.209      |
| P7             | 3.125            | 0.195 | 0.189      |
| F/             | 3.123            | 0.183 | 0.103      |
| P8             | 0.000            | 0.119 | 0.121      |
| FO             | 0.000            | 0.123 | 0.121      |
| Cample: Huma   | n Milk (6000v)   | 0.776 | 0.757      |
| Sample: Huma   | II WIIIK (OUUUX) | 0.739 | 0.757      |

#### **Standard Curve**

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



#### **Performance Characteristics**

- The minimum detectable dose of albumin as calculated by 2SD from the mean of a zero standard was established to be 1.5 ng/ml.
- Intra-assay precision was determined by testing replicates of three milk samples in one assay.
- Inter-assay precision was determined by testing three milk samples in twenty assays.

|                   | Intra-Assay Precision |      |      | Inter | -Assay Prec | ision |
|-------------------|-----------------------|------|------|-------|-------------|-------|
| Sample            | 1                     | 2    | 3    | 1     | 2           | 3     |
| n                 | 20                    | 20   | 20   | 20    | 20          | 20    |
| CV (%)            | 3.5%                  | 4.2% | 4.0% | 8.9%  | 9.5%        | 8.8%  |
| Average<br>CV (%) | 3.9%                  |      |      |       | 9.0%        |       |

## Recovery

| Standard Added Value | 6.25 – 50 ng/ml |  |
|----------------------|-----------------|--|
| Recovery %           | 91 – 110%       |  |
| Average Recovery %   | 96.5%           |  |

## Linearity

• Milk samples were serially-diluted to test for linearity.

|                 | Average Percentage of Expected Value (%) |
|-----------------|--|
| Sample Dilution | Human Milk                               |
| 1:3000          | 96%                                      |
| 1:6000          | 98%                                      |
| 1:12000         | 105%                                     |

## **Cross-Reactivity**

| Species | Cross Reactivity (%) |  |  |
|---------|----------------------|--|--|
| Beagle  | None                 |  |  |
| Bovine  | None                 |  |  |
| Monkey  | None                 |  |  |
| Mouse   | None                 |  |  |
| Rat     | None                 |  |  |
| Swine   | None                 |  |  |
| Rabbit  | None                 |  |  |
| Human   | 100%                 |  |  |

# **Troubleshooting**

| Issue  | Causes                                      | Course of Action   |  |  |
|--|---|--|--|--|
|  | Use of expired                              | Check the expiration date listed before use.                                       |  |  |
|  | components                                  | Do not interchange components from different lots.                                 |  |  |
|  |   | <ul> <li>Check that the correct wash buffer is being used.</li> </ul>              |  |  |
|  |   | Check that all wells are empty after aspiration.                                   |  |  |
|  | Improper wash step                          | Check that the microplate washer is dispensing properly.                           |  |  |
|  |   | If washing by pipette, check for proper pipetting                                  |  |  |
| Low Precision                                | Splashing of reagents while loading wells   | technique.  • Pipette properly in a controlled and careful manner.                 |  |  |
| Je C   |   | Pipette properly in a controlled and careful manner.                               |  |  |
| P  | Inconsistent volumes                        | Check pipette calibration.   |  |  |
| 8  | loaded into wells                           | Check pipette for proper performance.  |  |  |
| _  | Insufficient miving of                      | Thoroughly agitate the lyophilized components after                                |  |  |
|  | Insufficient mixing of<br>reagent dilutions | reconstitution.  |  |  |
|  | reagent unutions                            | Thoroughly mix dilutions.  |  |  |
|  |   | <ul> <li>Check the microplate pouch for proper sealing.</li> </ul>                 |  |  |
|  | Improperly sealed                           | Check that the microplate pouch has no punctures.                                  |  |  |
|  | microplate                                  | Check that three desiccants are inside the microplate                              |  |  |
|  | A4: 1: 16:                                  | pouch prior to sealing.  |  |  |
| a a  | Microplate was left<br>unattended between   | Each step of the procedure should be performed                                     |  |  |
| gu   | steps                                       | uninterrupted.   |  |  |
| i Si   | Omission of step                            | Consult the provided procedure for complete list of steps.                         |  |  |
| igh  | Steps performed in                          | Consult the provided procedure for the correct order.                              |  |  |
| Ξ.   | incorrect order                             | consult the provided procedure for the correct order.                              |  |  |
| Ē  | Insufficient amount of                      | Check pipette calibration.   |  |  |
| ow   | reagents added to                           | Check pipette for proper performance.  |  |  |
| ly Low o<br>Intensity                        | wells                                       |  |  |  |
| Unexpectedly Low or High Signal<br>Intensity | Wash step was skipped                       | Consult the provided procedure for all wash steps.                                 |  |  |
| ţ  | Improper wash buffer                        | Check that the correct wash buffer is being used.                                  |  |  |
| tbe  | Improper reagent                            | Consult reagent preparation section for the correct     dilutions of all reagents. |  |  |
| ) G  | preparation                                 | dilutions of all reagents.   |  |  |
| j  | Insufficient or<br>prolonged incubation     | Consult the provided procedure for correct incubation time.                        |  |  |
|  | periods                                     | unie.  |  |  |
|  | p   | Sandwich ELISA: If samples generate OD values higher                               |  |  |
| ا با   |   | than the highest standard point (P1), dilute samples                               |  |  |
| 证  |   | further and repeat the assay.  |  |  |
| Ş  | Non-optimal sample                          | Competitive ELISA: If samples generate OD values lower                             |  |  |
| , in   | dilution                                    | than the highest standard point (P1), dilute samples                               |  |  |
| Þ  |   | further and repeat the assay.  |  |  |
| dar  |   | User should determine the optimal dilution factor for                              |  |  |
| Deficient Standard Curve Fit                 | Contamination of                            | samples.      A new tip must be used for each addition of different                |  |  |
| St   | reagents                                    | samples or reagents during the assay procedure.                                    |  |  |
| , i  | Contents of wells                           | Verify that the sealing film is firmly in place before placing                     |  |  |
| cie  | evaporated                                  | the assay in the incubator or at room temperature.                                 |  |  |
| efi  |   | Pipette properly in a controlled and careful manner.                               |  |  |
|  | Improper pipetting                          | Check pipette calibration.   |  |  |
|  |   | Check pipette for proper performance.  |  |  |

| Insufficient mixing of reagent dilutions | Thoroughly agitate the lyophilized components after reconstitution. Thoroughly mix dilutions. |
|--|---|
|--|---|

#### References

(1) Gekle M. (2004) Ann. Rev. Physiol.

Version 8.3R

#### **Related Product**

- EA2201-1 AssayMax Human Albumin ELISA Kit (Plasma and Serum samples)
- EKA2201-1 AssayMax Monkey Albumin ELISA Kit (Plasma and Serum samples)
- EKA3201-1 AssayMax Monkey Albumin ELISA Kit (Urine and Cell Culture samples)
- EMA2201-1 AssayMax Mouse Albumin ELISA Kit (Plasma and Serum samples)
- EMA3201-1 AssayMax Mouse Albumin ELISA Kit (Urine and Cell Culture samples)
- ERA2201-1 AssayMax Rat Albumin ELISA Kit (Plasma and Serum samples)
- ERA3201-1 AssayMax Rat Albumin ELISA Kit (Urine and Cell Culture samples)
- ETA2202-1 AssayMax Rabbit Albumin ELISA Kit (Plasma, Serum, Urine, and Cell Culture samples)
- EPA3201-1 AssayMax Swine Albumin ELISA Kit (Urine and Cell Culture samples)
- EPA2201-1 AssayMax Swine Albumin ELISA Kit (Plasma and Serum samples)