



Model PA-201

Programmable Pulse Switch Electrode Array Driver

User Manual

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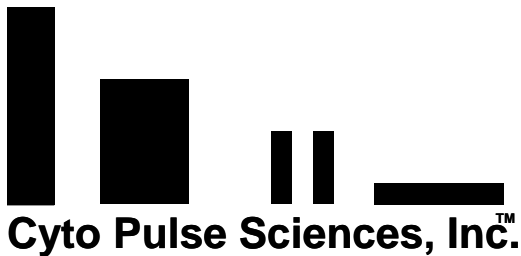
PA-201 Programmable Pulse Switch

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PA-201 Programmable Pulse Switch

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Table of Contents

| | Page |
|--|---|
| 1. Introduction | 1-1 |
| 2. Tutorial: Electroporation with PA-201 Switch | 2-1 |
| 2.1 Background | 2-1 |
| 2.2 Applications | 2-3 |
| 2.2.1 <i>In Vitro</i> Applications | 2-3 |
| 2.2.2 <i>In Vivo</i> Applications | 2-3 |
| References | 2-3 |
| 3. PA-4000/PA-201 Set-up and Operation | 3-1 |
| 3.1 Introduction | 3-1 |
| 3.2 Connecting the PA-201 to the PA-4000 | 3-1 |
| 3.3 High Voltage Outputs | 3-2 |
| 3.3.1 Custom Electrode Connector | 3-2 |
| 3.3.2 <i>In Vivo</i> / CE-24 Output Jacks | 3-3 |
| 3.3.3 Available <i>In Vivo</i> Electrodes | 3-3 |
| 3.4 PA-201 Front panel display | 3-4 |
| 4. Getting Started With the PA-201 | 4-1 |
| 4.1 Introduction | 4-1 |
| 4.2 Operational Concept | 4-1 |
| 4.3 Application Software | 4-2 |
| 4.3.1 Start Up Screen | 4-2 |
| 4.3.2 Select Application Type | 4-2 |
| 4.3.2.1 PPS Single Mode | 4-3 |
| 4.3.2.2 96-Well Model | 4-5 |
| 4.3.2.3 Custom PPS Mode | 4-5 |
| 5. Customer Service | 5-1 |
| 5.1 Limited Warranty | 5-1 |
| 5.2 Customer Service | 5-1 |
| Appendix A | Data Sheet |
| Appendix B | PA-201 Switch and Electrode Array Connections |

List of Figures

| | Page |
|---|-------------|
| 3-1 The Programmable Pulse Switch Electroporation System | 3-1 |
| 3-2 Back Panel Connections | 3-1 |
| 3-3 CS-201 Custom Output Cable | 3-2 |
| 3-4 Nine-Socket Output Jack | 3-2 |
| 3-5 Output Jacks for the optional CE-24 or in vivo electrodes | 3-3 |
| 3-6 Cyto Pulse Sciences <i>In Vivo</i> Electrodes | 3-3 |
| 3-7 PA-201 Front Panel Display | 3-3 |
| 3-8 Electrical Connections and Current Flow | 3-3 |
| | |
| 4-1 PA-201 Switches | 4-1 |
| 4-2 <i>PulseAgile</i> ® Opening Screen showing PA-201 PPS attached | 4-2 |
| 4-3 <i>Electroporation Pro Pulse Switch</i> Screen | 4-3 |
| 4-4 Available Electrodes for use with PA-201 | 4-3 |
| 4-5 PPS Single Mode Application Set-Up Screen | 4-4 |
| 4-6 Custom PPS Mode Application Set-Up Screen | 4-5 |
| 4-7 Screen after running Custom PPS protocol PPS-TEST1.pro | 4-6 |

Caution Notice

This instrument contains a high voltage power supply that can be adjusted to beyond 1,000 volts: such voltage can be lethal.

The user must read this manual carefully before the instrument is placed into operation.

Removing the cover may void the warranty.

Do not connect or disconnect the high voltage cable with the high voltage enabled. To connect or disconnect the cable, turn line/mains power off and unplug line cord.

Do not open the cuvette holder while the high voltage is on. If a problem occurs during a run, push the **STOP/RESET** button on the front panel.

If there are any questions about the operation of this instrument, call Cyto Pulse Customer service.

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1 Introduction

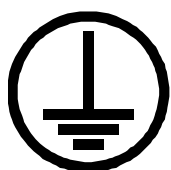
Electroporation has many uses in the fields of cell biology, medicine and microbiology. The PA-201 Programmable Pulse Switch add-on to the PA-4000 computer-controlled electroporator provides the ability to establish a number of electric field orientations *in vivo* or *in vitro*. The only limit to these field orientations is the configuration of the electrode used. All parameters are computer-controlled to give the operator maximum flexibility in performing optimization protocols.

This manual has been designed to help you realize the maximum benefit from using the PA-4000/PA-201 **PulseAgile®** system with programmable switching of pulsed electric fields. It contains information on how to operate the electroporator and switch, safety tips, and applications.

Note: The PA-4000 with the PA-201 contains a high voltage power supply and was designed with safety features to protect the user and the equipment. If used properly, the PA-4000 with the PA-201 is a safe and reliable instrument. Chapter 4 explains some important concepts related to operator safety in addition to concepts needed for accurate use of the instrument. Chapter 4 must be read before setting up this instrument. Our goal is the safe and productive use of the PA-4000 with the PA-201.

Back Panel Symbols

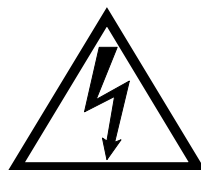
The PA-4000, with the optional PA-201, is rated for operation with line/mains voltage of



**Protective
Terminal
Conductor**



**Caution:
Refer to
documentation**



**Caution:
Risk of
electric shock**



**Chassis
Ground**

100-240 VAC, maximum current of 2 amps, at 50-60 Hz. The AC mains power supply cord is the disconnect device for this product. The power supply cord shall be an approved cord set Type SJT, rated 300 Volts AC, 18 AWG, 105° C, 3 conductor including ground.

This unit is rated for environmental conditions of 5-40° C, maximum relative humidity 80% for temperatures up to 31° C decreasing linearly to 50% relative humidity at 40° C, altitude to 2000 meters.

There are no operator replaceable parts inside the system; Cyto Pulse recommends that the user not remove the cabinet covers.

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2 Background and Applications

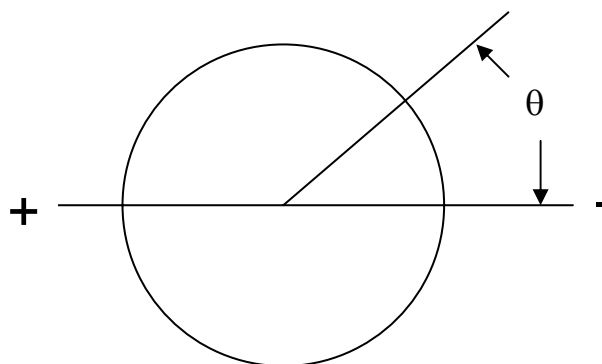
2.1 Background

The PA-201 is a user-programmable switch that is designed to direct the pulsed output of the PA-4000 electroporator to up to eight independent electrode elements. The **PulseAgile®** interface software included in the system allows the user to set each electrode to receive the pulse (anode), or to act as the return path (cathode), or to not be used at all. This ability provides flexibility in the delivery of electroporation pulses.

In the context of electroporation (also known as electropermeabilization), an electric field is used to induce a voltage across a cell membrane. If the induced voltage is above a critical threshold, pathways form in the membranes. These pathways permit material outside of the cell to enter the cytoplasm. These materials could be small molecule drugs or large molecules such as DNA. When the field is removed, the cell membrane seals with the material inside.

A critical feature of electroporation is that the pores (or areas of permeability) are formed in the ends of the cells in line with the electric field. This vectorial nature of electroporation is given by the formula for transmembrane voltage of a point in a cell membrane that is exposed to an electric field. The formula is $V = 1.5 r \times E \times \cos \theta$ (where V = transmembrane voltage, r = cell radius and E = applied electric field and θ = the angle shown below).

There are several implications to the vectorial nature of electroporation. One is that if poration is desired in areas of cells that are not in line with the electric field, then the angle of the electric field in relation to the cell needs to be changed. A principle use of the PA-201 is to assist in changing electric field vectors in cell suspensions and in tissue.



Another factor that affects the amount of permeability in cells in relation to the electric field vector is the resting transmembrane potential of cells before exposure to the electric field. This resting potential is approximately -70 mV. For electroporation, an average transmembrane voltage of approximately 1 volt is required. This higher transmembrane voltage is achieved by applying an electric field high enough to develop the desired transmembrane potential. However, because of the effect of the resting transmembrane potential there is actually a higher transmembrane at the pole of the cell facing the positive electrode than there is facing the negative electrode. This leads to an asymmetry in the permeabilized areas of the cell with a larger area of permeability at the pole of the cell facing the positive electrode. In fact this asymmetry has been observed by several investigators to include Sowers (1992) and Teissie (2002). Since DNA accumulates on the cell membrane with the smallest area of permeabilization, due to its negative charge, it may be more efficient if an electric field is first applied in one direction to induce a state of poration then applied in the opposite direction to move DNA to the cell and through the cell membrane.

2.2 Applications

There are many *in vitro* and *in vivo* applications for pulse-switching. **A number of these applications have been patented.** Cyto Pulse Sciences strongly cautions the user to not infringe on such patents. The most congested area in the intellectual property is the use of multi-element needle arrays *in vivo*. Several such patents are listed below. Some of these patents appear to have overlapping claims.

US 5,674,267, Mir et al, 07 October 1997 – *in vivo* use of an array of three or more needle electrodes where any electrode in the array may be connected to anode; and any other may be connected to cathode. There may also be a mode in which any number of pairs may be connected. A rectangular pulse sequence is used where all of the pulses are identical.

US 5,702,359, Hofmann, 30 December 1997 – *in vivo* use of an array of an array of six electrodes where opposing pairs, two anodes and two cathodes, may be connected in sequence.

US 6,041,252, Walker et al, 21 March 2000 - *in vivo* use of an array of three or more needle electrodes with at least one reference electrode and at least two satellite electrodes.

US 6,117,660 Walters et al, 12 September 2000 – an array of electrodes with at least one anode and one cathode connected and using a pulse sequences described in US 6,010,613, Walters et al.

2.2.1 *in vitro* and *ex vivo* Applications

One application of the PA-201 is reversing electric fields in a standard electroporation cuvette. This application permits the polarity of the electric field to be reversed from one pulse to the next. To perform this function, the PA-201 Programmable Pulse Switch and the CE-24 Cuvette Holder are required.

Sukharev, et. al. (1992) examined the movement of DNA into cells. His principle finding was that two phenomena are responsible for efficient DNA electrotransfection. One was electroporation induction of a permeability state in cells using high voltage pulsed electric fields. The other is movement of DNA into cells using electrophoresis. They speculated that the transfection efficiency might be increased by using the opposite polarity on the electrophoretic pulses. This suggestion supports the possible use of reversible electric fields to increase transfection efficiency.

2.2.3 *in vivo* Applications

As in the *in vitro* and *ex vivo* applications, the electric field between a cathode and anode may be reversed from pulse-to-pulse. To perform this function, the PA-201 Programmable Pulse Switch and one of the Cyto Pulse *in vivo* electrode arrays are required.

The first use of electroporation for *in vivo* gene delivery was by Titomirov (1991). This was done in mouse skin using parallel plate electrodes. Another tissue, muscle, was transfected by Lee (1992) using electroporation. Since then, many other tissues have been transfected using electroporation *in vivo*.

The next improvement in therapeutic efficiency was the use of acupuncture needles with electrochemotherapy (Mir 1993). The needles were used to penetrate the tumors to improve the depth of treatment of the tumor.

Arrays of electrodes were used to improve treatment coverage area. For this, switching of electric field vectors was done by Mir (1994) to independently address pairs of electrodes in the array imbedded in tissue. This process was described in an international patent application that included a needle array of three or more needles. Electrical pulses from a pulse generator were switched from pulse to pulse to address any single pair of electrodes for any one pulse (one anode and one cathode).

Another application of switching of *in vivo* implanted electrodes used opposing pairs of electrodes. Gilbert (1997) published a paper describing a pulse generator connected to a switching device connected two opposing pairs of electrodes in a six needle electrode array. The electric field was rotated 120 degrees among pulses, allowing delivery of electric fields at multiple angles during treatment.

Cyto Pulse has a number of electrodes available for *in vivo* transfection of tissues for use in animal research.

References

Currently, several hundred papers are being published yearly regarding *in vivo* and *in vitro* electroporation. It has a large potential for many uses.

In vivo expression of a nonselected gene transferred into murine hematopoietic stem cells by electroporation. Narayanan R, Jastreboff MM, Chiu CF, Bertino JR. *Biochem Biophys Res Commun.* 1986 Dec 30;141(3):1018-1024.

In vivo electroporation and stable transformation of skin cells of newborn mice by plasmid DNA. Titomirov AV, Sukharev S, Kistanova E. *Biochim Biophys Acta.* 1991 Jan 17;1088(1):131-134.

Electrochemotherapy: Potential of anti-tumor effect of bleomycin by local electrical pulses. Mir, LM, Orslowski, J, Belehradek Jr, J, Paoletti, C. *European J. Cancer.* 1991; 27: 68-72.

Electrochemotherapy, a novel anti-tumor treatment: first clinical trial. Mir. LM, Belehradek, M, Domenge, C, Orloski, S, Poddevin, B, Schwaab, G, Luboinski, B, Paoletti, Claude. *C.R. Acad. Sci. Paris* 1991; 313(3): 613-618.

Theoretical study of rectangular pulse electrical stimulation (RPES) on skin cells (in vivo) under conforming electrodes. Cheng K, Tarjan PP, Mertz PM. *Biomed Sci Instrum.* 1993;29:349-354.

A new brain tumor therapy combining bleomycin with in vivo electroporomeabilization. Salford, LG, Persson, BRR, Brun, A, Ceberg, CP, Kongstad, PC, Mir, LM. *Biochem Biophys Res. Comm.* 1993; 194(2): 938-943.

Electroporation of mammalian skin: a mechanism to enhance transdermal drug delivery.

Prausnitz MR, Bose VG, Langer R, Weaver JC. *Proc Natl Acad Sci U S A*. 1993 Nov 15;90(22):10504-10508.

Electrochemotherapy--a novel method of cancer treatment. Dev SB, Hofmann GA. *Cancer Treat Rev*. 1994 Jan;20(1):105-115.

Enhanced boron uptake in RG 2 rat gliomas by electroporpermabilization in vivo- a new possibility in boron. Ceberb, CP, Brun, A, Mir, LM, Persson, BRR, Salford, LG. *Anti-Cancer Drugs* 1994; 5:463-466

Treatment of cutaneous nodules using electrochemotherapy. Heller R. *J Fla Med Assoc*. 1995 Feb;82(2):147-150.

Direct visualization at the single-cell level of electrically mediated gene delivery. Golzio, M., J. Teissie, and M. P. Rols, *Proc.Natl.Acad.Sci. U.S.A*, 2002. 99, 1292-1297.

Novel electrode designs for electrochemotherapy. Gilbert, R. A., M. J. Jaroszeski, and R. Heller, *Biochim.Biophys.Acta*, 1997. 1334, 9-14.

3. PA-4000/PA201 Set-Up

3.1 Introduction

The PA-201 Programmable Pulse Switch is an optional attachment to the PA-4000 **PulseAgile®** Electroporator. It provides the ability to connect up to eight individual electrodes in various user-programmed configurations. The system consists of:



Figure 3-1: PA-4000/PA-201 Programmable Pulse Switch Electroporation System

| | |
|------------|-----------------------|
| PA-4000S | Electroporator System |
| PA-201 | Pulse Switch |
| CS-OPT | Interface Cable Set |
| CS-201 | Custom HV Cable |
| PA201-UMAN | User Manual |

Laptop shown is not included.

Optional accessories include:

| | |
|-------|--|
| CE-24 | Cuvette Holder for reversing electric fields |
|-------|--|

A selection of *in vivo* electrodes are available. These include parallel-row needle arrays(with limited customization), and tweezer electrodes.

3.2 Connecting the PA-201 to the PA-4000

Follow the instructions in Chapter 4 of the PA-4000 manual to set-up the PA-4000. The PA-201 should be placed on top of the PA-4000. Figure 3-2 shows the connections to be made. The Mains/Line Power Switch must be off and the Mains/Line power cord must be unplugged from the PA-4000.

- Connect the Option Interconnect cable between the DB25 connectors on the back of the PA-201 and the PA-4000.

**DO NOT ATTACH
TO A COMPUTER PARALLEL PORT
OR A PRINTER!**

- Connect the high voltage cable between the MHV connectors on the back of the PA-201 and PA-4000. This cord delivers the pulses generated by the PA-4000 to the PA-201

DO NOT USE A CABLE WITH BNC PLUGS!

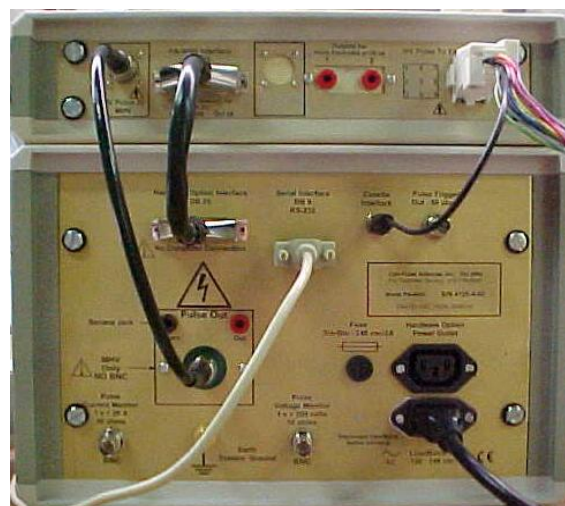


Figure 3-2: Back Panel Connections

3.3 High Voltage Outputs

There are two sets of output contacts on the back of the PA-201: the Custom Electrode connector and the In Vivo/CE-24 output jacks. It is important to note the following:

! ONLY ONE SET OF CONTACTS SHOULD BE USED AT ONE TIME !

3.3.1 Custom Electrode Connector

The Custom Electrode connector is shown in Figure 3-3. It is a nine-position jack that contains eight socket-contacts for use with the CS-201 Custom cable, shown in Figure 3-4. This cable is supplied with the PA-201 and is used to connect the user's custom electrode to the PA-201.

The CS-201 Custom cable has eight-wires with a nine-pin plug on one end and nothing on the other. The nine-pin plug is inserted into the on the back of the PA-201. The end of the high voltage cable without the plug is connected to the user's custom electrode. The user is responsible for using good practices in connecting the electrode to insure there is no safety hazard.

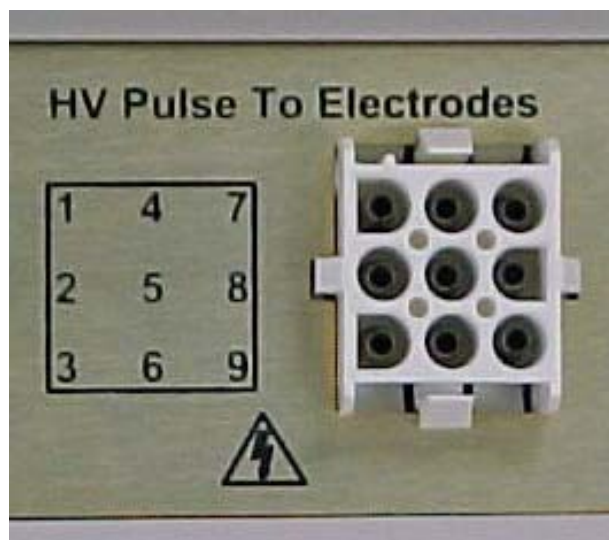


Figure 3-3: Nine-Position HV Output Jack

!CAUTION – DO NOT OPERATE THE UNIT WITH THE BARE WIRES AT THE END OF THE CS-201 CUSTOM CABLE UNDRESSED!

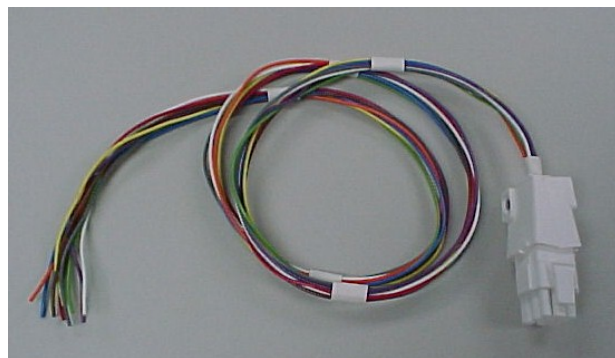


Figure 3-4: CS-201 Custom Output Cable for attaching custom electrodes

| CS-201 Cable Color to Pin Number Table | | |
|--|--------|---------------|
| Position | Color | Connection |
| 1 | Brown | Electrode 1 |
| 2 | Red | Electrode 2 |
| 3 | Orange | Electrode 3 |
| 4 | Yellow | Electrode 4 |
| 5 | Green | Electrode 5 |
| 6 | Blue | Electrode 6 |
| 7 | Violet | Electrode 7 |
| 8 | Grey | Electrode 8 |
| 9 | n/a | Not connected |

It is not necessary to use all of the wires provided. For example, if the custom electrode assembly consists of just five independent electrodes, then only connect them to wires 1 through 5 (brown -.green). However, take care to properly insulate the ends of the unused wires from unintended user contact. For additional safety, the protocol programming should only address electrodes that are actually connected to the wires.

3.3.2 In Vivo / CE-24 Output Jacks

These banana jacks are used to connect the optional CE-24 Cuvette Holder or *in vivo* electrodes to the PA-201. They are internally connected to the #1 and #2 sockets contacts of the Custom Electrode connector. They are particularly useful for electric field reversal protocols. If these output jacks are in use, the CS-201 Custom cable assembly must be removed.

The CE-24 is a standard cuvette holder with a dual banana-plug cable attached. The CE-24 is an optional accessory that must be ordered separately. To use a Cyto Pulse Sciences *in vivo* electrode with the PA-201, a special cable (CS-INVIVO-B) is required.



Figure 3-5: Output Jacks for the optional CE-24 or *in vivo* electrodes

3.3.3 Available In Vivo Electrodes

| | | |
|---------------------------------|-------|------------------------------|
| Gehl Parallel Row Needle Arrays | NE4-4 | 4mm row space, 4 needles/row |
| | NE4-6 | 4mm row space, 6 needles/row |
| | NE6-4 | 6mm row space, 4 needles/row |
| | NE6-6 | 6mm row space, 6 needles/row |

Needle lengths available from 2mm to 10mm

| | | |
|--------------------|---------|------------------------------|
| Tweezer Electrodes | TE-5-10 | Rectangular Pads, 5mm x 10mm |
| | TE-5R | Round Pad, 5mm diameter |

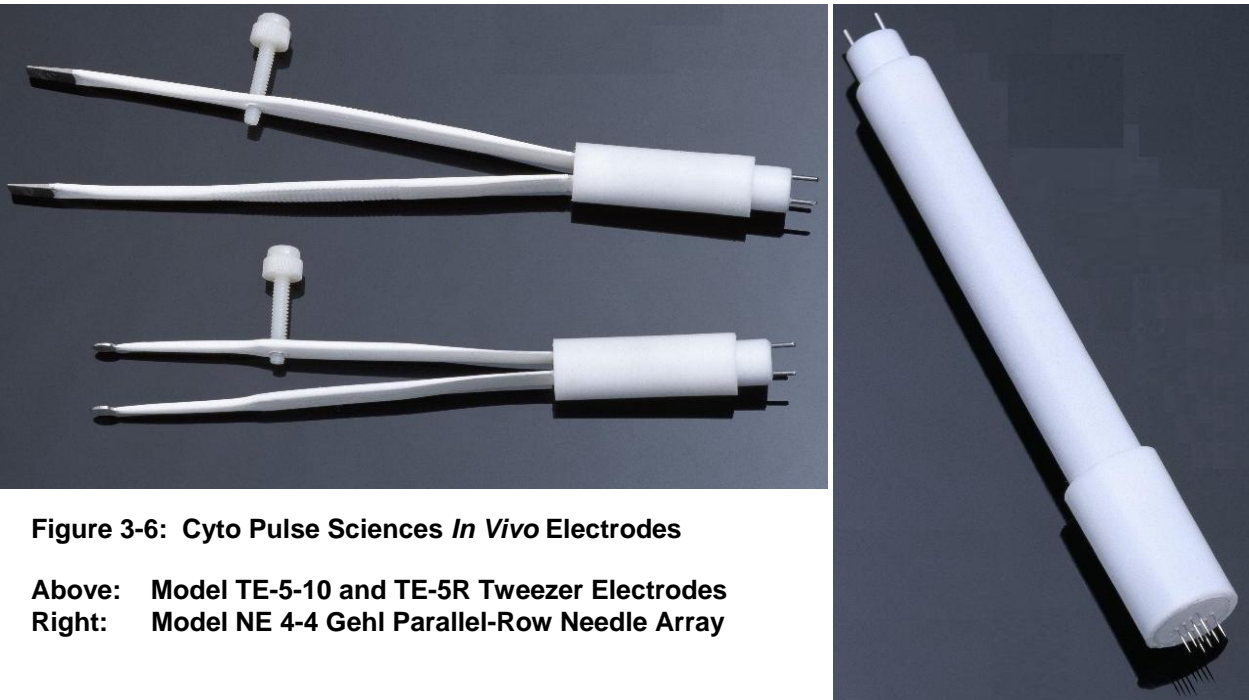


Figure 3-6: Cyto Pulse Sciences *In Vivo* Electrodes

Above: Model TE-5-10 and TE-5R Tweezer Electrodes
Right: Model NE 4-4 Gehl Parallel-Row Needle Array

3.4 PA-201 Front Panel Display

The PA-201 front panel consists of a series of light-emitting diode (LED) pairs that indicate the state of the internal switches for each output channel. Additionally, there is one LED labeled *Power* that will light when the PA-201 is properly connected to the PA-4000 and the line/mains power is on. For each output channel there is one red LED and one green LED. If a red LED is lit above an output number, then that output is internally connected to the anode of the PA-4000. In other words, that output channel will be the source of the pulse to the electrode attached to it. Conversely, if a green LED is lit below an output number, then the electrode attached to that channel will be connected to the cathode of the PA-4000. For example, if Channel 1 is lit red and Channel 2 is lit green (as shown in Figure 3-7), then the pulse current flows out of the PA-201 from pin 1 of the nine-wire jack, and flows back in to the PA-201 through pin 2. Figure 3-8 shows the internal connections made and the direction of current flow for the status shown in Figure 3-7.



Figure 3-7: PA-201 Front Panel Display

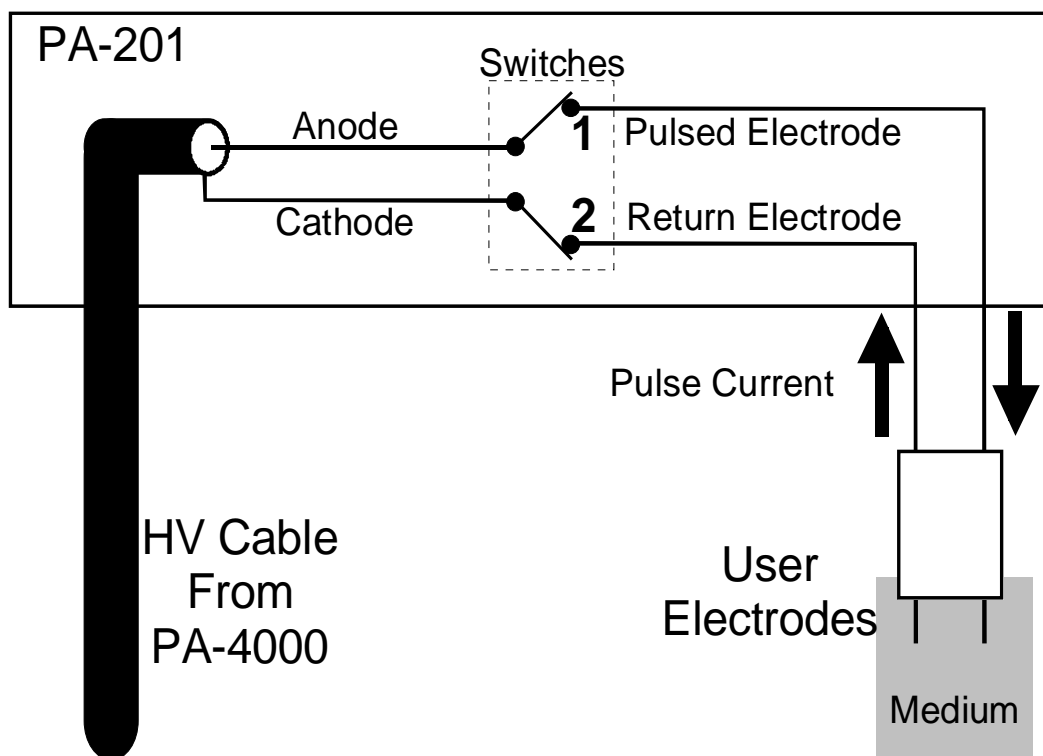


Figure 3-8: Electrical Connections and Current Flow with an electrode attached

4. Getting Started with the PA-201

4.1 Introduction

This Chapter will provide details regarding the software programming of protocols for the PA-201 using the PA-4000 *PulseAgile*® interface software.

4.2 Operational Concept

The PA-201 Programmable Pulse Switch contains eight three-state high-voltage switches that can connect each electrode in an array of up to eight electrodes in one of three configurations:

1. Connect the electrode to the anode (+ Out) of the PA-4000 Pulse Generator.
2. Connect the electrode to the cathode (- Return) of the PA-4000 Pulse Generator.
3. Make no connection to the electrode.

Important Note: In order for current to flow (and an electric field to be generated), **at least two** electrodes must be connected, **and** at least one of the electrodes must be an **anode** and one must be a **cathode**.

The PA-4000 *PulseAgile*® interface software is used to set the state of each switch for each pulse. In Figure 3-1, the PA-201 is configured to set the polarity for an array of two electrodes. Electrode 1 has been connected to the anode (+) and Electrode 2 has been connected to the cathode (-). So if the two electrodes were inserted in some conductive medium, the pulsed current would flow from Electrode 1 to Electrode 2.

The PA-4000/PA-201 can be programmed to change the state of the switches from one pulse to the next. For example, the polarity of the two electrodes connected in Figure 3.1 can be reversed after delivering a pulse and before delivering the next. In doing so, the electric field produced by the second pulse will be opposite that of the first pulse. The *PulseAgile*® software makes it easy with a graphical interface and “one-click” electrode connection setting.

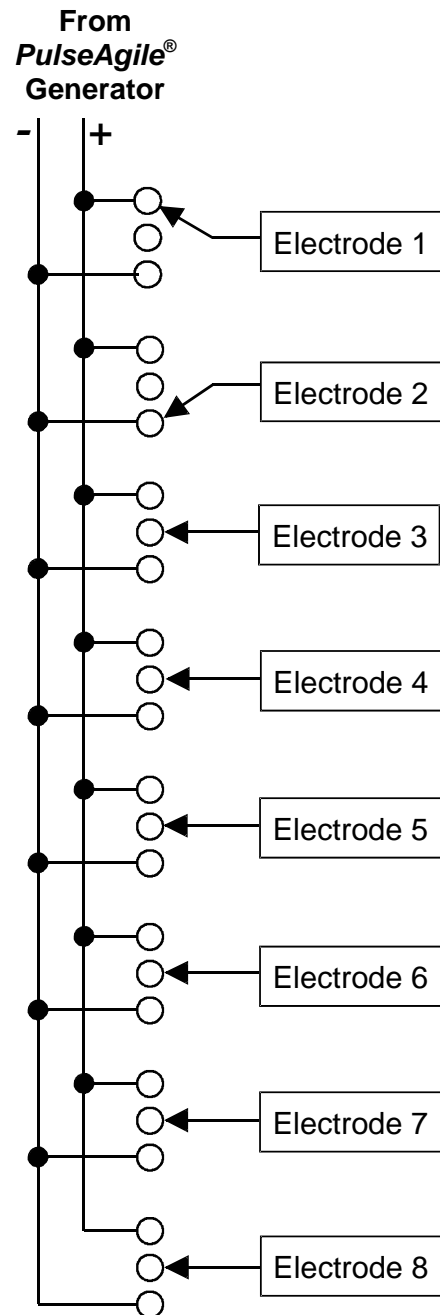


Figure 4-1: PA-201 Switches

4.3 Application Software

The PA-201 application software is a part of the PA-4000 **PulseAgile®** interface software. It is accessible only when a PA-201 is connected to the PA-4000 using the DB25 interface cable as described in Chapter 3.

4.3.1 Power Up

After installing the PA-201, turn on the PA-4000, and start the **PulseAgile®** software. The screen should appear as shown in Figure 4-2. In the *Options Connected* area, look for a check mark in Box 1 of the PPS section. If there is no check, then the DB25 interface cable is not connected properly. Power down the system and re-check the cable installation. If the problem persists, then call Cyto Pulse Sciences Customer Service.

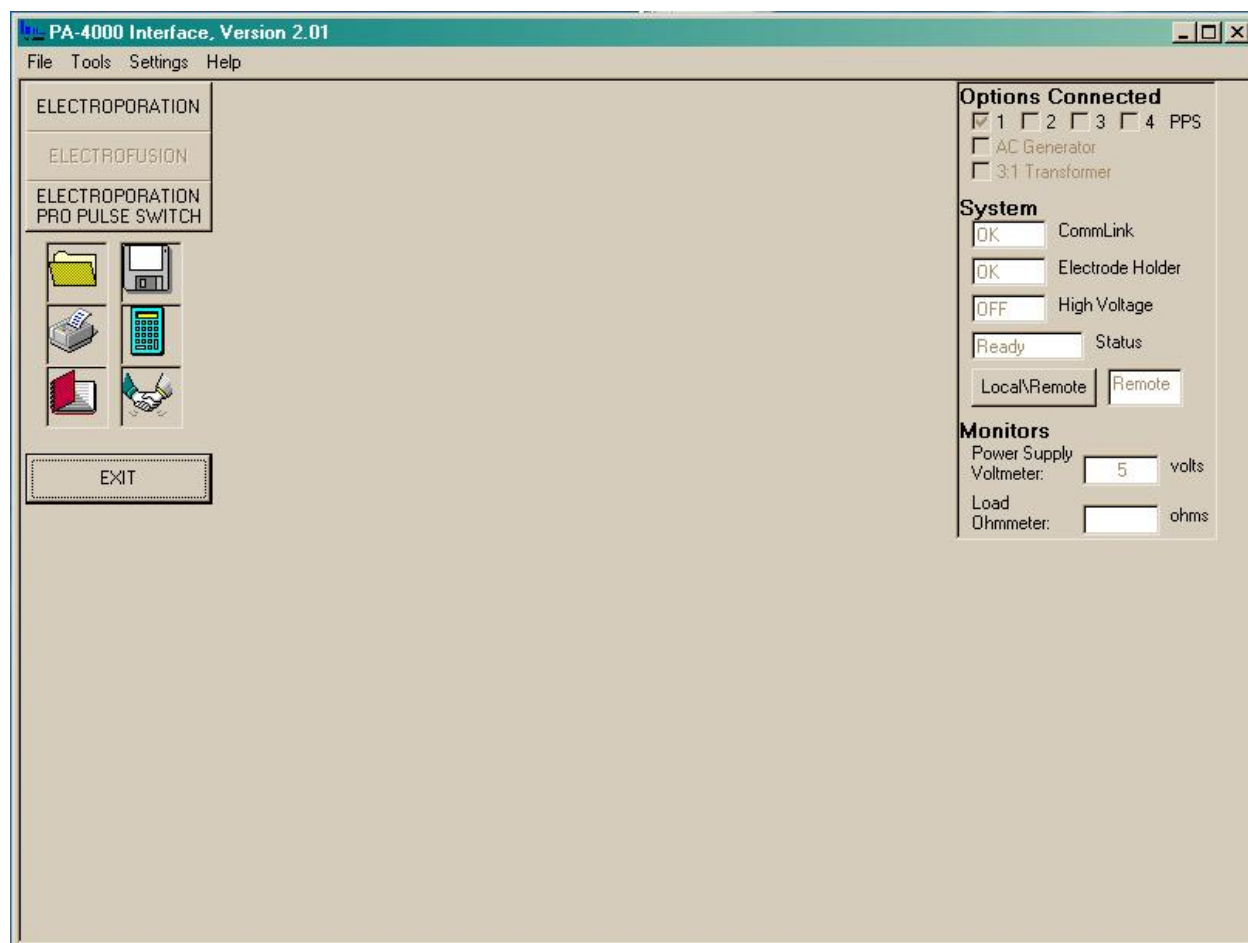


Figure 4-2: **PulseAgile®** Opening Screen showing PA-201 PPS attached

4.3.2 Select Application Type

Click *ELECTROPORATION PRO PULSE SWITCH*. The screen should now appear as shown in Figure 4-3. All of the regular panels and buttons appear with an additional set of buttons to the left of the *GROUP LIST* titled *Pro Pulse*. These are the various Programmable Pulse Switch application types available.

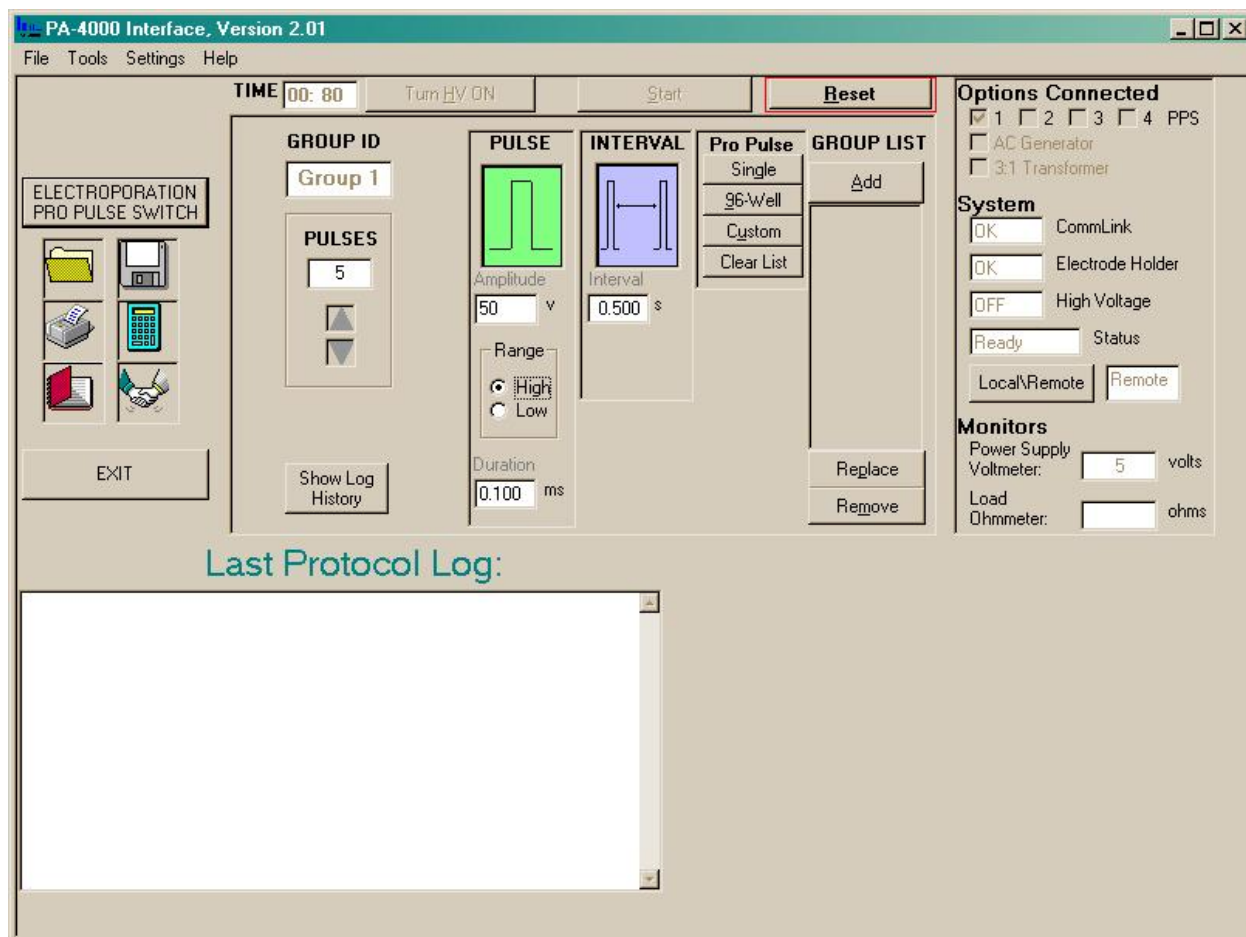


Figure 4-3: Electroporation Pro Pulse Switch Screen showing Pro Pulse Application Types

There are currently three Programmable Pulse Switch application modes available:

- Single
- 96-Well (for use with a PA-96W only)
- Custom

4.3.2.1 PPS Single Mode

This application permits reversing the electric field from pulse-to-pulse between a pair of electrodes. The two electrodes could be the plates in a standard electroporation cuvette, the needle rows in a Cyto Pulse Sciences Parallel-Row Needle Array, or the pads of a Cyto Pulse Sciences Tweezer Electrode. The Model CE-24 Cuvette Holder is required for use with electroporation cuvettes. The Models NE4-4 (or -6) and NE6-6 (or -4) Gehl parallel-row needle electrode; and the TE-5R and TE-5-10 are required for *in vivo* work using this application. All of the above are purchased separately as they are not included in the PA-201S system. Alternatively, the user may provide a custom-designed electrode and attach it to the system using the Custom cable assembly provided.

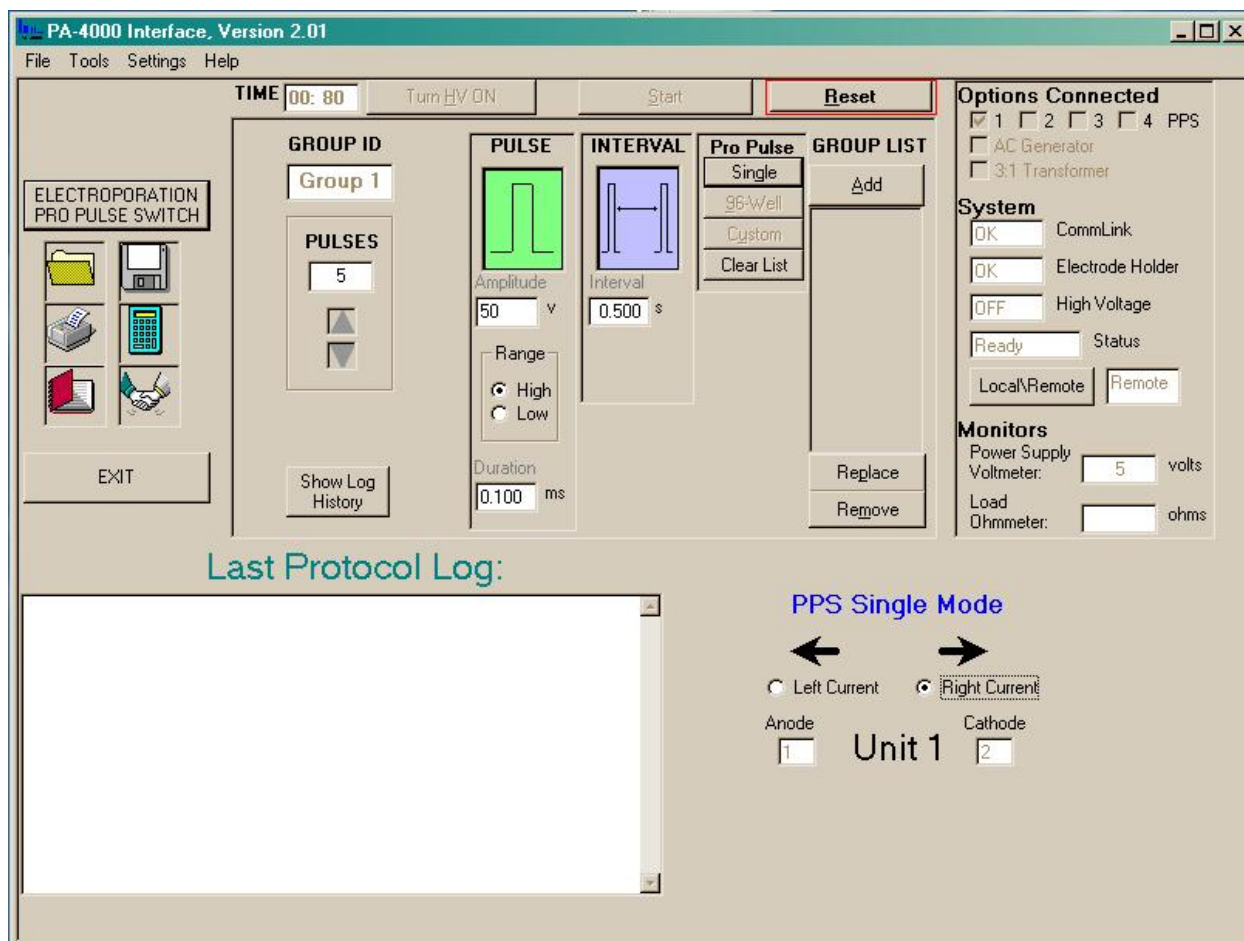


Figure 4-4: PPS Single Mode Application Set-Up Screen

Click *Single* and the **PPS Single Mode** screen appears as shown in Figure 4-4. In the lower right corner of the screen are two arrows labeled Left Current and Right Current. Imagine two electrodes side-by-side. “Right” current means that the current will flow from the left electrode (anode) to the right electrode (cathode). “Left” current is the opposite. Clicking *Right Current* sets Output 1 to the anode (+) and Output 2 to the cathode (-) inside the PA-201. The state of the two outputs is shown by the two status boxes below the arrows. If this protocol were run, current would flow from the electrode connected to Output 1, through the medium between the electrodes, and to the electrode connected to Output 2.

The polarity of the electric field between two electrodes corresponds with the direction of current flow. To program a simple electric field polarity reversing protocol, follow these steps:

- Click *Add* and Group 1 appears in the GROUP LIST.
- Set the pulse parameters, including the direction (Right or Left) of the current.
- Click *Add* and Group 2 appears in the GROUP LIST.
- Set the pulse parameters for Group 2, this time selecting the opposite direction.
- Click *Replace*.

If this protocol were run, the electric field generated by the Group 2 pulse(s) would be the opposite of that generated by the Group 1 pulse(s).

4.3.2.2 96-Well Mode

The 96-Well Application is only available when a Cyto Pulse Sciences Model PA-96W Programmable 96-Well Driver is installed. The PA-96W is an attachment to the PA-4000 **PulseAgile®** electroporator that provides the capability of directing the pulses from the PA-4000 to user-selected well(s) in a 96 well microplate. This User Manual does not cover the use of the PA-96W. Clicking *96-Well* will result in display of a message stating that a PA-96W is required.

4.3.2.3 Custom PPS Mode

This application gives the user the ability to utilize the full potential of the PA-201. Using the PA-201 with the included Custom HV cable assembly, up to eight electrodes can be connected in one of three states:

- Anode (+) Electrode is connected to the Pulse Out of the PA-4000.
- Cathode (-) Electrode is connected to the Return of the PA-4000.
- Not Connected Electrode is not in the current path at all.

To enter Custom PPS Mode, return to the Application Select screen by clicking *Clear List*, and then *OK* when prompted. Then click *Custom* and the **Custom PPS** screen appears as shown in Figure 4-5.

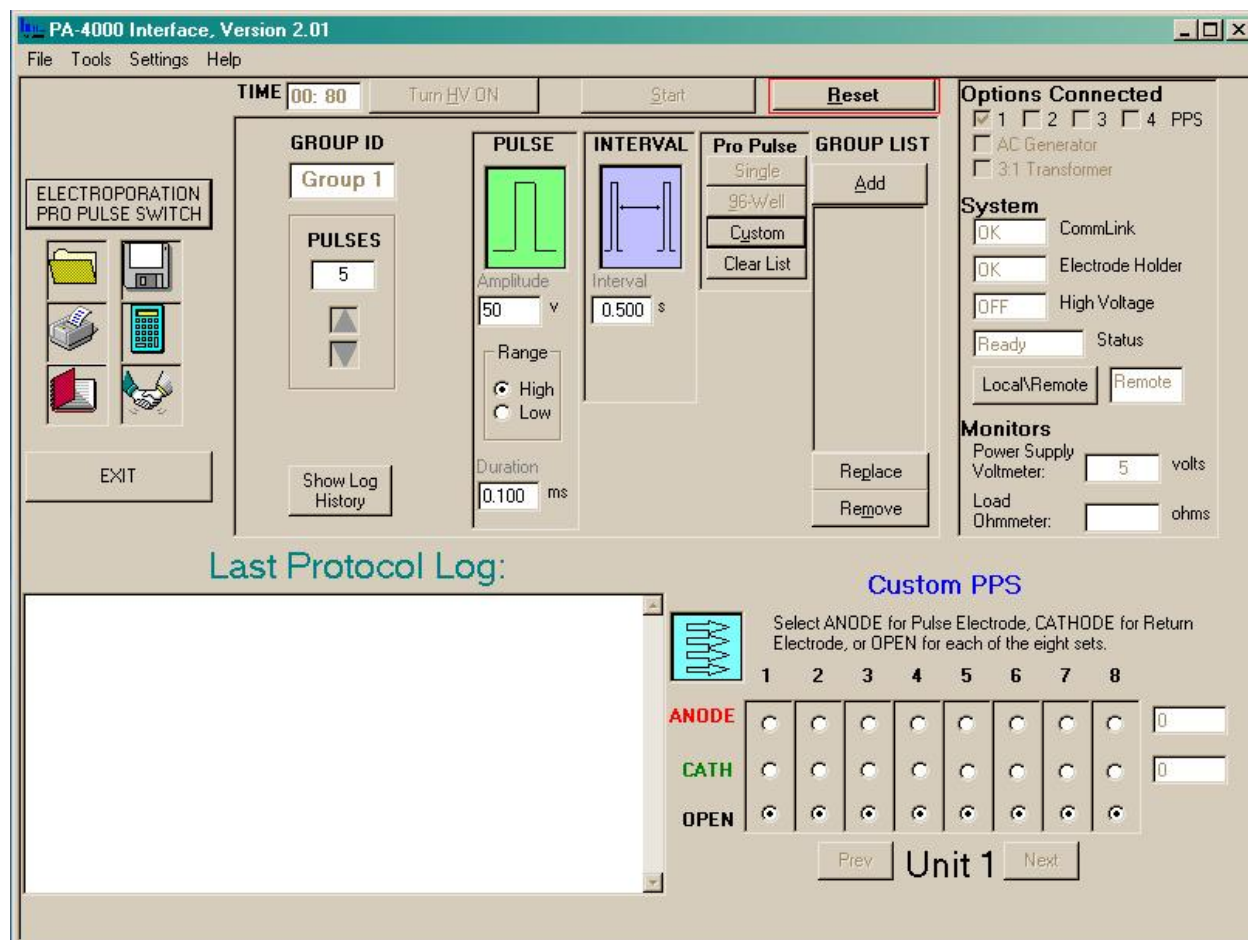


Figure 4-5: Custom PPS Mode Application Set-Up Screen

To program a Custom Mode protocol, use the lower right section of the screen to set the state of each electrode connected to the Custom HV cable. An example is provided here:

- Click *Add*. Group 1 appears in the GROUP LIST.
- Set parameters for Group 1 as shown below, Click *Replace*.
- Click *Add*, Set Group 2 as shown in below, Click *Replace*.
- Click *Add*, Set Group 3, as shown in below, Click *Replace*.
- Save the protocol as PPS-Test1.pro.
- Run the protocol by pushing F12.
- After the protocol is run the screen should appear as shown in Figure 4-6.

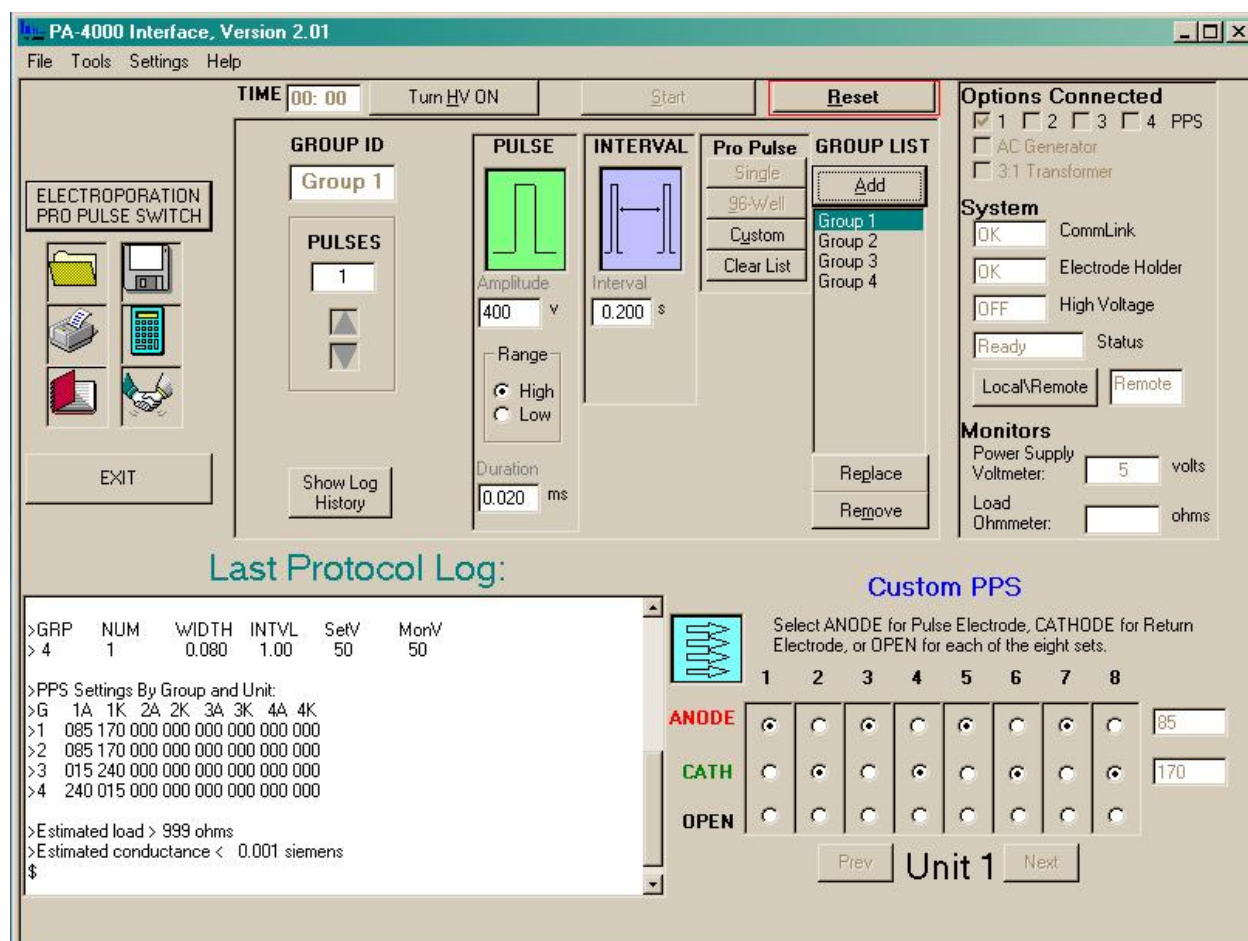
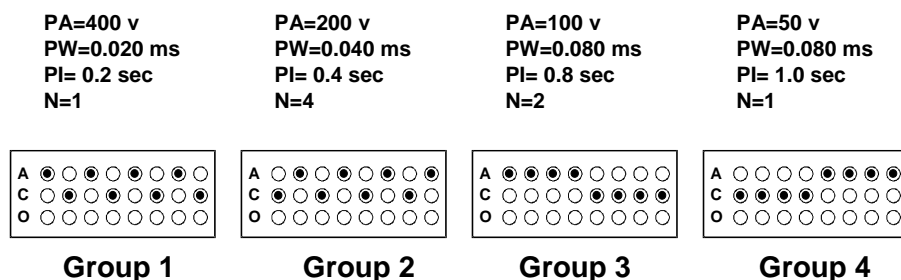


Figure 4-6: Screen after running Custom PPS protocol PPS-TEST1.pro

5. Customer Service

5.1 Limited Warranty

CYTO PULSE products are warranted against defect in materials and workmanship. If the customer provides notice of such a defect during warranty period, CYTO PULSE, at its option, will either repair or replace the products, which were found to be defective. The limited warranty set forth above is exclusive and no other warranty whether written or oral, is expressed or implied. CYTO PULSE specifically disclaims implied warranties of merchantability and fitness for a particular purpose.

EXCEPT AS SET FORTH ABOVE, CYTO PULSE MAKES NO WARRANTY WITH RESPECT TO THE PRODUCT, AND IN NO EVENT, REGARDLESS OF CAUSE, SHALL CYTO PULSE BE LIABLE FOR INDIRECT, SPECIAL, OR CONSEQUENTIAL DAMAGES OR OTHER LOSSES OF ANY KIND ARISING FROM BREACH OR WARRANTY OR OTHER USES OF THIS PRODUCT. CYTO PULSE'S OBLIGATION TO REPAIR OR TO REPLACE TO THE EXTENT SET FORTH ABOVE CONSTITUTES THE EXCLUSIVE REMEDIES OF THE CUSTOMER FOR ANY BREACH OF WARRANTY.

This warranty shall not apply to products, which after inspection by CYTO PULSE, were found to be improperly used or to have been modified in any manner. CYTO PULSE recommends that the user not open the product cabinet. This limited warranty is valid for one year from the date of shipment.

5.2 Customer Service

If the user believes that there is a defect in the CYTO PULSE product, the customer should contact CYTO PULSE Customer Service through our website at **www.cytopulse.com** or phone 410-787-1890, or contact the local CYTO PULSE representative. A determination if the product is still in warranty will be made. If the warranty period is still in effect, the user will be given an authorization number (RMA) to return the product. If after receipt and inspection the product is found to be defective, it will be replaced or repaired and returned to the customer. If the product is found to have been modified or misused, the user will be given a quote for repair. If the warranty period has expired and the user requests repair, CYTO PULSE will inspect the product and provide a written quote for repair. The user must provide a purchase order number before the product will be repaired. If the unit is damaged in shipment, the user must recover the insured value to replace or repair from the carrier.

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