

Plasma-Serum HPV (High and Low Risk) PCR Detection Kit Product Insert Product # 31600

More than 70 types of human papillomavirus (HPV) have been identified, and are generally classified as high-risk or low-risk depending on their relationship or lack of relationship with cancer and high-grade cervical intraepithelial neoplasia (CIN 2-3). HPV viruses are predominantly sexually transmitted and high-risk HPV types are a major risk factor for development of cervical cancer. Low-risk HPV types 6 and 11 have been associated with the presence of genital warts. There are many other low-risk HPV types that are not associated with genital warts or cervical cancer. Until now, HPV cannot be cultured *in vitro*, and immunological tests are inadequate to determine the presence of HPV cervical infection. On the other hand, biopsies can be analyzed by nucleic acid hybridization to directly detect the presence of HPV DNA. HPV 16 and HPV 18 have been considered as high-risk cancer associated HPV types. HPV types 31, 33, and 35 have been shown to have an intermediate relationship with cancer. These five HPV types, including types 39, 45, 51, 52, 56, 58, 59 and 68, have been identified as the principal HPVs detectable in the remaining cancers.

Principle of the Test

Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit constituents a ready-to-use system for the isolation and detection of 18 HPV DNA groups low-risk HPV types 6/11/42/43/44; and high/intermediate-risk HPV types 16/18/31/33/35/39/45/51/52/56/58/59/68 using end-point PCR. The kit first allows for the isolation of total DNA, including viral DNA, from the plasma-serum samples using spin-column chromatography based on Norgen's proprietary resin. The viral DNA is isolated free from inhibitors, and can then be used as the template in a PCR reaction for HPV and HPV (Low Risk) detection using the provided HPV Master Mix. The HPV Master Mix contains reagents and enzymes for the specific amplification of a 280 bp region of HPV High and Low Risk. In addition, Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit contains a second heterologous amplification system to identify possible PCR inhibition and/or inadequate isolation. The amplification and detection of either the *HPV Isolation Control (IsoC)* or the *PCR control (PCRC)* does not reduce the detection limit of the analytical HPV (High and Low Risk) PCR. The kit is designed to allow for the testing of 24 samples.

Component	Contents
Lysis Solution	30 mL
Binding Solution 1	6 mL
Binding Solution 2	6 mL
Wash Solution	22 mL
Elution Buffer	3 mL
Mini Filter Spin Columns	24
Collection Tubes	24
Elution tubes (1.7 mL)	24
HPV 2X PCR Master Mix	0.35 mL
Control 2x PCR Master Mix	0.35 mL
Isolation Control (IsoC)* ^a	0.3 mL
HPV Positive Control (PosC)* ^b	0.1 mL
Nuclease-Free Water	1.25 mL
Norgen's DNA Marker	0.1 mL
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Kit Components:

*IsoC = Isolateion Control; PosC= Positive Control

^aThe isolation control is a cloned PCR product

^b The positive control is a fragment of HPV cloned in a plasmid

Customer-Supplied Reagents and Equipment

- Disposable powder-free gloves
- Benchtop microcentrifuge and 60°C incubator
- Micropipettors and Sterile pipette tips with filters
- PCR tubes
- 96 100% ethanol

Storage Conditions and Product Stability

All buffers should be kept tightly sealed and stored at room temperature (15-25°C) for up to 1 year without showing any reduction in performance.

The HPV 2X PCR Master Mix, Control 2X PCR Master Mix, Isolation Control (IsoC), and HPV Positive Control (PosC) should be kept tightly sealed and stored at -20° C for up to 1 year without showing any reduction in performance. Repeated thawing and freezing (> 2 x) should be avoided, as this may reduce the sensitivity. If the reagents are to be used only intermittently, they should be frozen in aliquots.

General Precautions

The user should exercise the following precautions while using the kit:

- Use sterile pipette tips with filters.
- Store and extract positive material (specimens, controls and amplicons) separately from all other reagents and add it to the reaction mix in a spatially separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Work quickly on ice

Quality Control

In accordance with Norgen's ISO 9001 and ISO 13485-certified Quality Management System, each lot of Norgen's HPV 2X PCR Master Mix, Control 2X PCR Master Mix, Isolation Control (IsoC) and HPV Positive Control (PosC) are tested against predetermined specifications to ensure consistent product quality

Product Use Limitations

Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit is designed for research purposes only. It is not intended for human or diagnostic use.

Product Warranty and Satisfaction Guarantee

NORGEN BIOTEK CORPORATION guarantees the performance of all products in the manner described in our product manual. The customer must determine the suitability of the product for its particular use.

Safety Information

Ensure that a suitable lab coat, disposable gloves and protective goggles are worn when working with chemicals. For more information, please consult the appropriate Material Safety Data Sheets (MSDSs). These are available as convenient PDF files online at <u>www.norgenbiotek.com</u>.

CAUTION: DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

The Lysis Solution, Binding Solution 1 and Binding Solution 2 contain guanidine hydrochloride, and should be handled with care. Guanidine hydrochloride forms highly reactive compounds when combined with bleach, thus care must be taken to properly dispose of any of these solutions.

If liquid containing these buffers is spilt, clean with suitable laboratory detergent and water. If the spilt liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

Plasma or Serum of all human and animal subjects is considered potentially infectious. All necessary precautions recommended by the appropriate authorities in the country of use should be taken when working with plasma or serum.

1. Protocol

A. Specimen Collection, Storage and Transport

Precaution: All samples have to be treated as potentially infectious material.

1. Specimen Collection and Sample Storage

- Blood withdrawal causes injury of blood vessels (arteries, veins and capillaries).
- Only safe and sterile material should be used.
- For blood withdrawal appropriate disposables are available. For the vein puncture, too fine capillary needles should not be employed.
- Venous blood withdrawal should be carried out on the appropriate parts of the elbow bend, the forearm or the back of the hand.
- Blood has to be withdrawn with standard specimen collection tubes (red cap, Sarstedt or equivalent tube of another manufacturer). 5 10 ml EDTA blood should be withdrawn.

Precaution: Samples of heparinised humans must not be used

2. Sample Storage

- Whole blood should be separated into plasma and cellular components by centrifugation for 20 minutes at 800 1,600 x g within six hours. The isolated plasma has to be transferred into sterile polypropylene tubes.
- The sensitivity of the assay can be reduced if you freeze the samples as a matter of routine or store them for a longer period of time.
- Virus encapsulated DNA is stable for days if stored at +4°C, for weeks if stored at -20°C and even for months and years when stored at -70°C.

3. Sample Transport

- Sample material should be transported in a shatterproof, leak-proof transport container as a matter of principle. Thus, a potential danger of infection due to a leakage of sample can be avoided.
- The samples should be transported following the local and national instructions for the transport of pathogen material.
- We recommend sample transport with a courier. The blood samples should be shipped cooled (+2°C to +8°C) and the separated plasma deep frozen (-20°C).

4. Interfering substances

- Elevated levels of bilirubin (15 mg/dl) and lipids (800 mg/dl) and haemolytic samples do not influence the system.
- Heparin (10 IU/ml) affects the PCR. Samples which have been collected in tubes containing heparin as an anticoagulant should not to be used. Also, samples of heparinised patients must not be used.

B. Isolation of DNA from Plasma-Serum

Notes:

• Ensure that all solutions are at room temperature prior to use, and that no precipitates have formed. If necessary, warm the solutions and mix well until the solutions become clear again.

- Clean, disposable gloves should be worn at all times when handling reagents, samples, pipettes, disposable tubes, etc. It is recommended that gloves are changed frequently to avoid contamination
- All centrifugation steps are performed at room temperature.
- A variable speed centrifuge should be used for maximum kit performance. If a variable speed centrifuge is not available a fixed speed centrifuge can be used, however reduced yields may be observed.
- Preheat an incubator or heating block to 60°C.
- Prepare a working concentration of the **Wash Solution** by adding 50 mL of 96-100% ethanol (provided by the user) to the supplied bottles containing the concentrated Wash Solution. This will give a final volume of 72 mL. The labels on the bottles have a box that may be checked to indicate that the ethanol has been added.
- Ensure that samples have not undergone more than one freeze-thaw cycle, as this may lead to DNA degradation.
- Binding Solution 1 contains resin and must be mixed well before every pipetting.
- This kit is suitable for the isolation of DNA from fresh or frozen serum or plasma prepared from blood collected on either Heparin, EDTA or citrate.
- Isolation Control (IsoC)
 - An Isolation Control (IsoC) is supplied. This allows the user to control the DNA isolation procedure. For this assay, add the Isolation Control (IsoC) as indicated during the isolation procedure
 - The Isolation Control (IsoC) must not be added to the sample material directly.
 - Do not freeze and thaw the Isolation Control (IsoC) more than 2 times.
 - The Isolation Control (IsoC) must be kept on ice at all times during the isolation procedure.
 - The PCR components of the Plasma-Serum HPV (High and Low Risk) PCR Detection Kit should remain at -20°C until DNA is extracted and ready for PCR amplification.
 - It is important to work quickly during this procedure.
 - 1. In a 2mL tube add 400 μ L Plasma/Serum sample.
 - 2. To each 400 μ L Plasma/Serum sample add 1.2 mL of Lysis Solution. Mix well by vortexing for 15 seconds.
 - 3. Incubate the mixture from **Step 2** for 10 minutes at 60°C.
 - 4. After incubation add 200 μL of **Binding Solution 1** and mix well by vortexing for 15 seconds. (Note: Binding Solution 1 contains resin and must be mixed well before every pipeting)
 - 5. Centrifuge the mixture from Step 4 for 1 minute at 2,000 RPM, then carefully decant the supernatant in order to ensure that the slurry pellet is not dislodged.
 - To the slurry pellet from Step 5 add 180 μL from Binding Solution 2 followed by the addition of 420 μL of 96-100% Ethanol (provided by the user) and 10 μL Isolation Control (IsoC). Mix well by vortexing for 15 seconds.
 - 7. Centrifuge the mixture from **Step 6** for **1 minute at 2,000 RPM**, then carefully decant the supernatant in order to ensure that the slurry pellet is not dislodged.
 - 8. To the slurry pellet from **Step 7** add 1 mL **Wash Solution**, mix well by vortexing for 15 seconds and then centrifuge for **1 minute at 2,000 RPM**. Carefully decant the supernatant in order to ensure that the slurry pellet is not dislodged.
 - 9. Repeat **Step 8** to wash the slurry pellet for a second time.
 - 10. To the slurry pellet from Step 9 add 500 µL Wash Solution, Mix well by vortexing for 15 seconds.
 - 11. Transfer the entire mixture from **Step 10** into a Mini Filter Spin column. Centrifuge for **1 minute at 14,000 RPM**. Discard the flowthrough and reassemble the spin column with its collection tube.
 - 12. Spin the column, empty, for 2 minute at 14,000 RPM. Discard the collection tube.
 - Transfer the spin column to a fresh 1.7 mL Elution tube. Apply 100 μL of Elution Buffer to the column and centrifuge for 2 minutes at 200 x g (~2,000 RPM), followed by 2 minute at 10,000 x g (~14,000 RPM).

C. HPV (High and Low Risk) PCR Assay Preparation

Notes:

- Before use, suitable amounts of all PCR components should be completely thawed at room temperature, vortexed and centrifuged briefly.
- The amount of HPV 2X Detection PCR Master Mix and Control 2X PCR Master Mix provided is enough for up to 32 PCR reactions (24 sample PCR, 4 positive control PCR and 4 no template control PCR).
- For each sample, one PCR reaction using the HPV 2X PCR Master Mix and one PCR reaction using Control 2X PCR Master Mix should be set up in order to have a proper interpretation of the results.
- For every PCR run, one reaction containing HPV Positive Control and one reaction as no template control must be included for proper interpretation of results.
- The recommended minimum number of DNA samples tested per PCR run is 6.
- Using a lower volume from the sample than recommended may affect the sensitivity of the HPV (High and Low Risk) Limit of Detection.
 - Prepare the PCR reaction for sample detection (Set #1, using HPV 2X PCR Mastermix) and the PCR reaction for control detection (Set #2, using Control 2X PCR Mastermix) as shown in Table 1 below. The recommended amount of sample DNA to be used is 2.5 μL. However, a volume between 1 and 5 μL of sample DNA may be used as template. Ensure that one HPV detection reaction and one control reaction is prepared for each DNA sample. Adjust the final volume of the PCR reaction to 20 μL using the Nuclease-Free Water provided.

PCR Components	Volume Per PCR Reaction
HPV 2X PCR Master Mix OR Control 2X PCR Master Mix	10 μL
Sample DNA	2.5 μL
Nuclease-Free Water	7.5 μL
Total Volume	20 µL

Table 1. PCR Assay Preparation

2. For each PCR set, prepare **one** positive control PCR as shown in Table 2 below:

Table 2	PCR	Positive	Control	Preparation
	I UN	I USILIVE	CONTROL	

PCR Components	Volume Per PCR Reaction
HPV 2X PCR Master Mix OR Control 2X PCR Master Mix	10 µL
HPV Positive Control (PosC)	10 µL
Total Volume	20 µL

3. For each PCR set, prepare **one** no template control PCR as shown in Table 3 below:

PCR Components	Volume Per PCR Reaction
HPV (2X PCR Master Mix OR Control 2X PCR Master Mix	10 μL
Nuclease-Free Water	10 µL
Total Volume	20 µL

Table 3. PCR Negative Control Preparation

E. HPV (High and Low Risk) PCR Assay Programming

1. Program the thermocylcer according to the program shown in Table 4 below. 2. Run one step PCR.

PCR Cycle	Step	Temperature	Duration
Cycle 1	Step 1	95°C	3 min
	Step 1	94°C	15 sec
Cycle 2 (40x)	Step 2	60°C	30 sec
	Step 3	72°C	45 sec
Cycle 3	Step 1	72°C	5 min
Cycle 4	Step 1	4°C	œ

Table 4: HPV (High Risk) PCR Assay Program

E. HPV PCR Assay Interpretation

- 1. For the analysis of the PCR data, the entire 20 μL PCR reaction should be loaded on a 1X TAE 2% Agarose DNA gel along with 10 μL of Norgen's DNA Marker (provided).
- 2. The PCR products should be resolved on the 1X TAE, 2% Agarose gel at 150V for 30 minutes (Gel running time will vary depending on an electrophoresis apparatus).

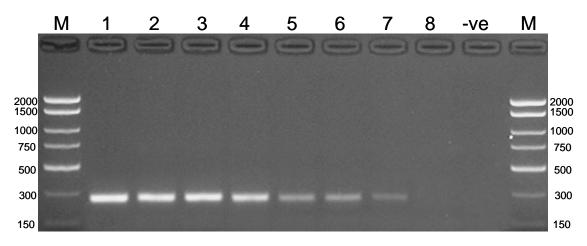


Figure 1: A representative 1X TAE 2% agarose gel showing the amplification of HPV at different concentrations. The size of the HPV target amplicon corresponds to the 280 bp band represented by the provided DNA Marker (M). No amplification of the target is observed in with the Negative Control

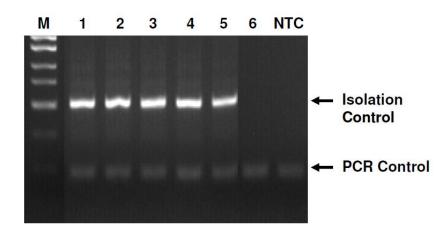


Figure 2: A representative 1X TAE 1.7% agarose gel showing the amplification of Isolation Control and PCR Control under different conditions using the Control 2X PCR Master Mix. The size of the Isolation Control amplicon and PCR Control amplicon correspond to 499 bp and 150 bp, respectively, as represented by the provided DNA Marker (M). Lanes 1 to 5 showed detection of both Isolation Control and PCR Control, suggesting that the DNA isolation as well as the PCR reaction was successful. Lane 6 showed only the detection of PCR Control suggesting that while the PCR was successful, the isolation failed to recover even the spiked-in Isolation control. NTC=Negative Control.

Input Type	Target Reaction	Control Reaction		Interpretation
	HPV (High and Low Risk) Target Band (280 bp)	lsoC Band (499bp)	PCRC Band (150 bp)	
Positive Control	x	х	Х	Valid
Negative Control			Х	Valid
Sample	х	Х	Х	Positive
Sample		Х	Х	Negative
Sample		Х		Negative
Sample			Х	Re-Test
Sample				Re-Test
Sample	Х	Х		Positive
Sample	Х		Х	Positive
Sample	Х			Re-Test

Table 5.	Interpretation of PC	R Assay Results
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** For results obtained that are not covered in Table 5 above, please refer to the Troubleshooting Section.

** Ignore any bands that appear between the Isolation Control band and the PCR Control band

F. Specificity

 The specificity of Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit is first and foremost ensured by the selection of the HPV-specific primers, as well as the selection of stringent reaction conditions. The primers were checked for possible homologies in GenBank published sequences by sequence comparison analyses. Furthermore, the specificity of the HPV-specific primers were tested against most of the known sexually-transmitted pathogens.

G. Linear Range

- The linear range (analytical measurement) of Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit was determined by analyzing a dilution series of an HPV quantitative standard ranging from 8.46 x 10⁹ VP/µl to 1 x 10⁻¹ IU/µl.
- Each dilution has been tested in replicates (n = 4) using Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit on 1X TAE, 1.7% Agarose gels.
- The linear range of Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit has been determined to cover concentrations from 2.7 VP/µI to at least 8 x 10⁶ VP/µI
- Under the conditions of Norgen's Plasma-Serum DNA Isolation procedure, Norgen's Plasma-Serum HPV PCR detection Kit covers a linear range from 200VP/mL Plasma-Serum to at least 8 x 10⁹ VP/mL Plasma-Serum.

G. Frequently Asked Questions

1. How many samples should be included per PCR run?

- Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit is designed to test 24 samples. For every 6 samples, a Negative Control and a Positive Control must be included. It is preferable to pool and test 6 samples at a time. If not, the provided Negative Control and Positive Control are enough to run 3 samples at a time.
- 2. How can I interpret my results for a sample if neither the HPV PCR Control nor the HPV Isolation Control (IsoC) amplifies?
 - If neither the HPV PCR control nor the HPV Isolation Control (IsoC) amplifies, the sample must be retested. If the positive control showed amplification, then the problem occurred during the isolation, where as if the Positive control did not amplify the problem has occurred during the setup of the PCR assay reaction.

3. How should it be interpreted if only the HPV PCR control showed amplification but neither the HPV target nor the HPV Isolation Control (IsoC) amplified for a sample?

- This indicates a poor isolation. The isolation procedure must be repeated.
- 4. How should it be interpreted if only the HPV Isolation Control (IsoC) was amplified in a sample?
 The sample tested can be considered as HPV negative.

5. How should it be interpreted if only the HPV target and the HPV PCR control were amplified in a sample?

• The sample tested can be considered as HPV positive.

6. How should it be interpreted if only the HPV target was amplified in a sample?

• The sample tested can be considered positive. At high HPV viral load, the HPV amplicon will be predominant and the HPV PCR control as well as the HPV Isolation control may not amplify.

7. How should it be interpreted if only the HPV PCR control and the HPV Isolation Control (IsoC) showed amplification?

• The sample tested can be considered negative

8. Can I process a different Plasma-Serum volume?

• The reagents provided with the isolation kit are only sufficient to process 24 Plasma-serum samples of 0.5mL each.

9. What If I added more or less of the specified reagents' volume?

 Adding less volume may reduce your DNA yields. Adding more may not affect the DNA yields EXCEPT if more Elution Buffer was added. Eluting DNA in higher volumes of Elution Buffer will result in diluting your DNA.

10. What If my incubation temperature varied from the specified 60°C?

• The incubation temperature can be in the range of 55°C - 65°C. At other temperatures the activity of both the Proteinase K and the Pronase will be reduced. This will result in a reduction in your DNA yields.

11. What If my incubation varied from the 20 minutes specified in the product manual?

• Less than 20 minutes will result in lower DNA yields. More than 20 minutes may not affect your DNA yields.

12. What If I forgot to do a dry spin after my second wash?

• Your first DNA elution will be contaminated with the Wash Solution. This may dilute the DNA yield in your first elution and it may interfere with your down stream applications.

13. What If I forgot to add the HPV Isolation Control (IsoC) during the isolation?

• The isolation must be repeated.

Technical Assistance

NORGEN's Technical Service Department is staffed by experienced scientists with extensive practical and theoretical expertise in sample and assay technologies and the use of NORGEN products. If you have any questions or experience any difficulties regarding Norgen's Plasma-Serum HPV (High and Low Risk) PCR Detection Kit or NORGEN products in general, please do not hesitate to contact us.

NORGEN customers are a valuable source of information regarding advanced or specialized uses of our products. This information is helpful to other scientists as well as to the researchers at NORGEN. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques.

For technical assistance and more information, please contact our Technical Support Team between the hours of 8:30 and 5:30 (Eastern Standard Time) at (905) 227-8848 or Toll Free at 1-866-667-4362 or call one of the NORGEN local distributors (<u>www.norgenbiotek.com</u>) or through email at techsupport@norgenbiotek.com.

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