LabBase User Manual, Version 1.0

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Abstract

LabBase is a generic database management system for implementation of laboratory information systems. This manual constitutes both a tutorial introduction to and a reference manual for LabBase.

1 Data Model

The primary abstractions supplied by LabBase are *materials* and *steps*: materials are things one works on in the laboratory, such as genetic markers or expressed sequence tags, and steps record both actions taken on a material and the information they generate. For example, at the Whitehead/MIT Center for Genome Research (CGR), the discovery of a mouse genetic marker involves many steps:

- 1. sequencing a small DNA fragment,
- 2. determining if it contains a simple repeat (microsatellite),
- 3. checking to see that we have not already used this sequence,
- 4. selecting polymerase chain-reaction (PCR) primers,

and so on. In this example, the material is the potential genetic marker, and the steps correspond to the actions 1 through 4. Our experiences with MapBase [3, 4, 2], as well as

other literature on laboratory information systems [5, 6], suggest that the notions of material and step are ubiquitous in these systems.

Additional material describing LabBase appears in [8] (which discusses the user view of LabBase), and [7] (which focuses on LabBase's implementation). The CGR's approach to workflow management and CGR's workflow-management software are described in [9]. Documentation on the perl API to LabBase is found with the LabBase distribution.

In LabBase, almost all information about a material is stored as part of a step. In our work with MapBase we found this to be a robust organization, because we want almost all attributes of a material to be associated with information about the processing step that produced them. For example, when we record the sequence of a potential marker, we also record the file that contains the output of the automated DNA sequencer, as well as the user that entered the sequence and the time a which the sequence was entered.

In LabBase, each material is associated with

- a *material kind*, such as **marker** or **est** (expressed sequence tag), and
- a history (list) of steps that record the chronology of real-world operations performed on that material.

A step is associated with a *step kind*. For example, step 1 above might have the step kind **sequence_step**. In addition to its step kind, a step consists of a set of *tag*/value pairs, for example **sequence='ACCG...'** or **sequence_file='/usr/local/seq_output/3851'**. Tags serve as attribute names, and each tag has a unique type that defines the values that can be associated with that tag. Examples of types for a tag are 'STRING' or'SET'('INTEGER').

The current version of LabBase does not automatically support ordering constraints among steps, though this is a capability that would probably be useful.

In addition to materials and steps, LabBase provides *data-dictionary entries* that record what sorts of materials, steps, and tags are known to the system.

The remainder of this manual is organized so that the information most users will require appears first, followed by information that is of interest to smaller numbers of users. Therefore, the next section describes how to pose read-only LabBase queries. Subsequent sections describe how to pose queries that update the database and how to define new kinds of materials and steps. Sections 4 and 5 act as a reference manual: they describes all the predicates available in LabBase and define the grammar of the LabBase query language.

2 Queries

Queries (and updates) are posed in a non-recursive logic programming language: the syntax and semantics are essentially those of a subset of Prolog [1]. The essential idea is that Lab-Base automatically defines certain predicates over the materials, steps, and tags constituting the state of a LabBase database. For example, if t is a tag, then the predicate t(M,S) binds S to the most recent value associated with t in the step history associated with the material bound to M.

Each query is a non-empty sequence of comma-separated *terms*, terminated by a period. For example

```
marker(M),sequence(M,Seq).
```

This query prints, for each marker, M, that has a sequence, the most recent sequence recorded for M. The two terms in this query are marker(M) and sequence(M,Seq). To answer this query, each marker in the database is successively bound (assigned) to M, and then the sequence predicate binds S to the value associated with the most recent sequence tag in any step belonging to that particular marker.

Assuming there are four markers in the database, the output would look something like this:

```
M=B123,Seq=ACTTG...
M=Q443,Seq=GGATTG...
M=Z1N4,Seq=CCAAG...
M=L96,Seq=CTTTA...
YES
```

In this output, each variable is printed with a binding that made the query true. (A variable is an alphanumeric identifier beginning with a upper-case letter.) If the query has any bindings that make it true, LabBase prints YES after printing out the bindings. This is the default output. It is possible to tailor the form of the output by using the only predicate.

In the query above, marker is a material kind, and sequence is a tag. For any material kind k, the predicate k(X) binds X to each material of kind k. For any tag, t, the predicate t(M,V) requires M to be bound to a material. Then the predicate t binds V to each value associated with t in the most recent step of M that has an instance of t. Because the first argument to the predicate t must be bound, and the second argument can be free, we say that its *adornment* is $[bf]^1$

Tags can be set- (or list-) valued. For example, in the mouse genetic mapping schema at CGR, the tag duplicate_id_set is associated with a set of (string, integer) pairs, where each of the strings is the id of a marker, and each of the integers is a measure similarity between two sequences.

Sometimes many steps have the same tag; for example, every step has a when and a who tag. To restrict LabBase to the most recent step of a particular step kind, one can pose a query such as the following:

```
marker(M), sequence_step(material=M, sequence=Seq, when=W).
```

In this query, sequence_step is a step kind, and Seq and W are bound to the values of the sequence and when tag (respectively) of the most recent sequence_step. If, for example, there were no sequence tag associated with marker m's most recent sequence_step, m would not appear as a binding for M in the output of from this query.

Error Reporting When LabBase detects a syntax or evaluation error it prints out a string of the form

¹Some Prolog manuals label an argument with '-' to indicate that it can be free (corresponding to LabBase's 'f'), and with '+' to indicate that it must be bound (corresponding to LabBase's 'b').

ERROR=message near line n, column m.

where *message* tries to explain what went wrong, n is a line number near the error, and m is the column position near the error. If you keep each term in a query to single line, n should be accurate. For evaluation errors, m is often at the end of the term which could not be evaluated. For a single query, it is possible, for some valid bindings to be printed before LabBase detects an evaluation error.

Queries can be interrupted with control-C (SIGINT), in which case an error message is printed. However, the SIGINT must directed to either the lbserv or lbback process. For more details see the man pages for these programs and also [7].

It is often desirable to have a query print out an error message when no bindings can be found to make a term true. The **insist** predicate can do this. For example

```
marker(M), sequence_step(material=M),
insist(sequence_step(material=M,sequence=Seq,when=W)).
```

which prints an error for those markers that have no sequence or when tag on in their most recent sequence_step. The insist predicate succeeds each time its argument succeeds, put prints out an error message if its argument does not succeed at least once.

Example Queries We close the section on queries with a few more example queries:

• Print all markers and selected typing_step information for those markers that have been typed on more than 2 typing panels.

```
marker(M),
count( typing_step(material=M,the_typing_panel=P), C),
C > 2,typing_step(material=M,the_typing_panel=Q).
```

• Print the number of markers that have been typed on more than 3 typing panels.

```
count( marker(M),
            count(typing_panel(P), typing_string(M,P,S), C),
            C > 3, D ).
```

• Find all markers with no step with a sequence tag.

```
marker(M), not(sequence(M,D)).
```

• Find all markers whose most recent typing_step *follows* a mapmaker_step (two versions).

```
marker(M),next(M,typing_step,mapmaker_step),
    mapmaker_step(material=M).
```

```
marker(M), mapmaker_step(material=M,when=W1),
    typing_step(material=M,when=W2), W2 > W1.
```

• Print all markers that have *any* typing step following a mapmaker step.

```
marker(M),all_steps(M,S1),all_steps(M,S2),
mapmaker_step(S1),typing_step(S2),when(S1,W1),
when(S2,W2),W2 > W1.
```

3 Writes

Tag associations are a key concept in the expression of writes to LabBase. A tag associations is a syntactic representation of a tag/value pair to be inserted into the database, and is of the form

t=v

where t is a tag (as determined by the set of tags in data dictionary), and v is a single value.

3.1 Inserts

Inserts of a new instance of a material have the form

```
insert(x(args))
```

where x is a material kind, and *args* is a comma-separated list of tag associations. The *args* must contain tag associations with tags x_{id} (whose value becomes the id of the newly created material) and who. The result is the creation of a new material with a first step of kind create with the specified tag/value pairs, and, in addition the a created_material tag associated with the newly created material itself.

Here is code to insert a new marker:

```
insert(marker(marker_id='A1', who='', when=1991:06:12:09:23:47)).
```

Here is code to create a screening panel:

```
insert(
  screening_panel(
    screening_panel_id='standard mouse screening panel',
    who=steve, when=1991:01:01:00:00:00
    screening_panel_abbreviations
    =[ob, cast, spr, a, b6, c3h, dba, balb, akr, non, nod, lp],
    screening_panel_members
    =['C57/6J-0b/0b', 'Castaneus', 'Spretus', 'A/J',
        'C57bl/6J', 'C3H/HEJ', 'DBA/2J', 'BALB/CJ', 'AKR/J',
        'NON', 'NOD', 'LP/J'] )).
```

Inserts of a new step have the same form as insert of a new material instance, except that x must a step kind; args must contain a who tag, and must contain at least one tag of type 'MATERIAL_POINTER'.

Here is an example of inserting a step of kind external_choice_step.

```
insist(marker_id(M,'L59')),
insert(external_choice_step(
  material=M,who='',when=1991:07:09:10:45:08,
  left_primer='ATGGGTACCACCCTATCATACCTA',
  right_primer='TTATACACTGATATCTTGATAGCC',
  product_length=48,
  external_choice_source='First WIBR Bluescript Library')).
```

3.2 Value Sets

Value sets (also called "material sets" for historical reasons) are sets of values that occur "at the top level"—not as part of a step. They are often used to hold partial results of a multi-statement query, or to represent the state of materials in a laboratory production line.

The predicates value_set and temp_value_set allow one to manipulate permanent and temporary value sets. Currently value_set's are completely persistent: they survive the shutdown and restart of the database server, while temp_value_set's disappear when the database server is shut down. The predicates material_set and temp_material_set are retained for backward compatibility with earlier releases of LabBase; they are *synonyms* for value_set and temp_value_set, respectively. In addition lbserv (see the lbserv man page) provides temp_material_set's that are visible only within a single database session.

Section 4.2 describes how to use value_set and temp_value_set to query value sets. To insert a value into a value set one would write something like

```
marker_id(M,'D1118'),insert(value_set('My Set',M)).
```

which inserts the marker with id D1118 into value set 'My Set'. To create a temporary value set containing all markers with no sequence write

marker(M),not(sequence(M,S)),insert(temp_value_set(no_sequence,M)).

Here is an example of deleting a *particular* value from a material set

```
hybrid_screening_panel(P,panel_X),delete(value_set(good_panels,P)).
```

This query deletes the panel with panel_X from the set good_panels.

It will soon be impossible to delete an entire value set (or temporary value set) with name X using the form

```
delete(value_set(X))
```

Instead, use

```
delete_entire_value_set(X).
```

and

```
delete_entire_temp_value_set(X).
```

It is safe to insert to or delete from a value set while iterating over it. LabBase makes a copy of the value set before finding bindings for value_set [bf] or temp_value_set [bf].

3.3 Deletes and Updates

We expect deletes and updates to be rare, and used only to make corrections to data when no workflow step is needed to record the correction. Since MapBase currently offers no facilities for updates or deletes, their implementation in LabBase (except for value sets) has been deferred. Current practice is to edit the ASCII roll-forward logs to perform the updates and deletes when they are absolutely required.

4 Reference Manual

4.1 Types

The legal atomic types are: 'STRING', 'INTEGER', 'FLOAT', 'DATE', 'DNA_SEQUENCE', 'BOOLEAN', 'MATERIAL_POINTER', 'STEP_POINTER', and 'TERM'. Every atomic type is a legal type. The legal type constructors are: 'LIST', 'SLIST' and 'SET'. 'SLIST' is a space-efficient representation for lists that have mostly zero elements. (The zero elements are the empty string for 'STRING', 0 for 'INTEGER' and BOOLEAN, 0.0 for 'FLOAT', the date corresponding to the Unix time_t 0 for 'DATE', the empty sequence for 'DNA_SEQUENCE', the empty set for 'SET', and the empty list for 'LIST' and 'SLIST'. An 'SLIST' cannot have 'MATERIAL_POINTER', 'TERM', or 'STEP_POINTER' elements.)

For any legal types t1, ..., tn, n>0, and for any legal type constructor U, U(t1,..., tn) is a legal type. Neither 'STEP_POINTER' nor 'TERM' can be used as part of the type of a tag.

4.2 Predicates

The available predicates are either built in (like insert and not), or are defined by the contents of the data dictionary.

As of April, 1997, it is possible to logically delete steps. The effect is to simply *mark* a step as deleted; no storage space is released, and no index entries for associated identifier tags are removed. It is possible to see logically deleted steps if the predicate see_deleted_steps has previously been evaluated in the current query.

Definition: A step is *visible* unless it is logically deleted or **see_deleted_steps** has been evaluated in the current query.

- $material_kind(X)$ [f],[b] True if X is a material with kind $material_kind$.
- tag(R, Id) [fb], [bb] Provided tag is an identifier-tag, true if R is a material with Id associated with tag at some step. (An identifier-tag is one with the tag id_tag set to 1 in its data-dictionary entry.)
- tag(M1,...,Mk,V) [b...bf] (For cases not subsumed by the previous entry.) We must have k>0. Each Mi must be bound to a value of type 'MATERIAL_POINTER'. Search the intersection of the histories of M1,...,Mk from the most recent step backward, until a visible step is found with tag tag. Bind V to the corresponding associated value. (The tag tag must not be a data-dictionary tag—one with the tag id_tag set to 1 in its data-dictionary entry).
- tag(S, V) [bf] Bind V to the value associated with tag in step S. (tag must not be a data-dictionary tag.)
- $step_kind(S)$ [b], True if S is a step of kind $step_kind$.
- $step_kind(t1=x1,...,tn=xn)$ [b...b] Predicates of this form allow greater precision than those based on tags: $step_kind$ predicates can determine which tags are collected in a particular step, and can determine the tags with which materials are associated, thereby differentiating the roles of the materials. At least one ti must have type 'MATERIAL_ POINTER' and the associated xi must be bound. In the current implementation there can be no more than one unbound ti with type 'MATERIAL_POINTER'. Bindings for unbound xi's are found as follows:
 - Let S be the set containing exactly those steps, s, such that all of the following obtain:
 - * s is visible.
 - * The step kind of s is $step_kind$.
 - * For all the ti's of type 'MATERIAL_POINTER', s is the most recent step with any particular mapping of the ti's to material pointers.
 - * For every bound xi, ti is associated with xi's value.
 - For each s in S do:

- * Bind each free xi to the value associated with ti in s and return true.
- +, -, *, / These symbols are not the names of predicates, but can be the principal functor of a term argument to the is predicate.
- >,<,>=, <=, <>, == [bb] Binary comparison operators. <> is the inequality predicate, and == is the equality predicate, needed because underbound methods are not yet supported. May be used as infix operators.
- Pattern ~ String [bb] True if String contains the regular expression Pattern. Pattern and String must have type 'STRING'. The syntax of Pattern is that of the Unix editor ed(1), except that newlines are allowed in Pattern; for documentation use the Unix command man ed. Please note that in order to get a backslash into Pattern, is necessary to use two backslashes in the quoted string. For example, '^\\(.).*\\1\$' ~ uveweru. (The pattern matches any string of length at least 2 with the same character at the beginning and the end.)
- $term\theta$;; term1 [bb] Evaluate term0. If there are any bindings that make it true, then return true for every binding that makes term0 true. If term0 is never true, then return true for every binding that makes term1 true. (;; is a short-circuit or operator.) In the query-language grammar, ;; binds more tightly than ,.
- all_steps(R,S) [bf],[fb] True if S is a visible step associated with material R. The bindings are guaranteed to produced in the order in which the steps appear in R's step history.
- baseline_rusage [] Create a baseline for measuring resource usage. (Resource usage is measured in lbback only.) See incremental_rusage.
- cardinality(Set, Cardinality) [bf] True if Cardinality is the cardinality of Set.
- commit [] Execute a commit in the underlying storage manager.
- count(*term1*,..., termn, C[b...bf] Binds Cto the number of times term1,...,termn is true. Any bindings produced in evaluating term1,...,termn are undone before evaluating the term following the count term.
- db_size(*Bytes*, *Blocks*) [ff] Bind *Bytes* to the number of bytes in the database, and *Blocks* to the number of blocks (as returned by Unix stat). Only the main database file or files are considered. Log files are excluded.
- delete(*term*) [b]

The only legal argument is a *term* of the form value_set(*string*, *value*) (or material_set(*string*, *valu*)), where both *string* and *value* are bound. The result is to delete *value* from the value set with name *string*.

- delete_entire_material_set(Set_name) [b] Delete the material set with name Set_name (which must be a 'STRING'). Always true (whether or not there is material set named Set_name).
- delete_entire_temp_material_set(*Set_name*) [b] Like delete_entire_material_set, except or temporary material sets.
- delete_step(*Step*) [b] Logically delete *Step*. (No error is raised if *Step* is already deleted.) See also undelete_step.
- element(collection, V1,..., Vn) [bx...x], x in {b,f} The collection must be of type 'LIST', 'SLIST', or 'SET'. If n=1, each element of collection is bound (in order, if collection is of type 'LIST' or 'SLIST') to V1. If n>1, each element of collection must be a list, [x1,...,xn] containing exactly n elements. For each element of collection (in order, if collection is of type 'LIST') bind Vj to xj.
- exists(term1,..., termn) [b...] True if term1,..., termn evaluated as a query (using any already-established bindings) is true. The argument query is not evaluated after one set bindings is found which makes it true; thus using exists might be more efficient than evaluating the argument query directly. It is an error to use the second kind of step_kind query within the argument of an exists predicate. In the current implementation the exists predicate can cause some storage leakage in the lbback server, so it should be used only when there is a compelling efficiency rationale.
- gather_in_list(term1,..., termn, Element, List) [b...bff] For each set of bindings for which term1,..., termn is true, take the value bound to Element and make it an element of List. The order of elements in List is determined by the order in which they are bound to Element by evaluating term1,..., termn.
- gather_in_set(term1,..., termn, Element, Set) [b...bff] For each set of bindings for which term1,..., termn is true, take the value bound to Element and make it an element of Set. All the values bound to Element must be comparable as if by ==.
- hex_escape(c) [b] Please use hex_escape_and_quote for all future coding. hex_escape is maintained only for backward compatibility. When printing out strings from the database, replace the characters

[,],{,},comma,newline,

and c itself by c followed by the character's 2-digit hex code. The intent of this predicate is to make it easy to parse LabBase output by means of simple regular expressions. The effect of evaluating this predicate is limited to the current query.

• hex_escape_and_quote(c) [b] When printing out strings from the database, replace the characters

[,],{,},(,),comma,newline,single quote,

and c itself by c followed by the character's 2-digit hex code. In addition enclose all strings in single quotes (including material-kind names and principal material ids when printing a MATERIAL_POINTER and step-kind names when printing a STEP_POINTER).

The intent of this predicate is to make it easy to parse LabBase output by means of simple regular expressions. The effect of evaluating this predicate is limited to the current query. The local perl module ../sitelisp/LabBase.pm is designed to turn the output of a query produced using this predicate into a normal perl5 data structure.

The format in which step "identifiers" are printed out is designed to make parsing by ../site_lisp/LabBase.pm reliable; they are printed out in the form

step('<kind>''%2c'(when=<timestamp>)) (%2c is the hex code for comma) (or step('<kind>''%2c'(when=<timestamp>' %5bdeleted%5d')) if the step is logically deleted).

- incremental_rusage(List) [f] Bind List to a list of triples. The first element of each triple is a string describing the resource. The second element is the amount of resource used since the most recent evaluation of baseline_rusage, and the third element is (ususally) the amount of resource used since lbback was invoked. The first three elements of List are user CPU time, system CPU time, and elapsed time (as in the default for csh's time command). Remaining elements (if any) are from getrusage(2).
- insert(*term*) [b] The insert predicate is discussed in various sections above.
- insist(*term*) [b] True whenever *term* is true. If there are no bindings that make *term* true, then insist prints out an error message.
- V is expression [fb] If expression is a non-term value, binds V to expression. If expression is a term with one of principal functors +, -, *, or / is evaluates expression according to C-like rules of arithmetic. In particular, / operating on 'INTEGER's yields an 'INTEGER', and the result of any expression containing a 'FLOAT' will be a 'FLOAT'. For the current implementation, you must use the standard (i.e. parenfix) syntax for +, -, *, and /, which are all binary operators.
- ith(*List*,*I*,*V*) [bbf], [bff] For adornment [bbf] the second bound argument must be of type 'INTEGER'. *List* must be of type 'LIST', 'SLIST', or 'SET'. Bind each element, *ei*, in *List* (in order) to *V*, and bind *i* to *I*.

Simple array subscription is performed by ith. For example, ith([a,b,c],0,Z) binds Z to a. Some more examples are:

```
1> ith([a,b,c],I,Z).
I=0,Z=a
I=1,Z=b
```

```
I=2,Z=c
YES
2> ith([[a,b],[1,2]],1,Z).
Z=[1,2]
YES
3> ith([[a,b],[1,2]],I,Z)
I=0,Z=[a,b]
I=1,Z=[1,2]
YES
```

- {left,right}_primer_sequence(M,Primer_sequence) [bf]
 - Special-purpose predicates for the CGR. $Primer_sequence$ becomes bound to the left or right primer sequence of M (a 'MATERIAL_POINTER') according to the rules detailed below. (Also see pcr_product_length.) These predicates signal an error in database state by binding $Primer_sequence$ to the empty string. They signal an incompatible database schema by the usual LabBase error mechanism (which aborts all query processing for the current query).
 - Let A be left or right.
 - Let s be an arbitrary step in the history of M such that s contains either the $A_$ start or the $A_$ primer tag, and such that no later step contains either tag.
 - Signal an error in database state if s contains both A_start and A_primer.
 - If s contains $A_{\texttt{start}}$ then
 - * Let *i* be the value associated with *A_start*, and let *j* be the value associated with *A_length* tag in *s*. Signal an error in database state if *A_length* is absent in *s*.
 - * Find the most recent insert_start, insert_length and sequence tags in any step at or before s. If any of these tags is missing signal an error in database state.
 - * Let q be the value associated with insert_start, let r be the value associated with insert_length, and let, S, bet the value associated with the sequence tag, respectively.
 - * For left_primer_sequence, bind *Primer_sequence* to the substring of *S* starting at position *i* (0-based) and of length *j*.
 - * For right_primer_sequence, bind *Primer_sequence* to the reverse complement of the substring of S starting at *i*-*j*+1 and of length *j*.
 - Otherwise, if s contains an A_primer tag, bind its value to Primer_sequence.
- length(X,Length) [bf] True if Length is the length of X (which must be of type 'LIST', 'SLIST', 'STRING', or 'DNA_SEQUENCE'.
- make_list(V1, ..., Vn, L) [b...bf] Bind L to the list containing V1, ..., Vn, where n must be greater than 0 and each Vi must be a value or a variable bound to a value.

- material_set(Set_name) [f],[b] True if Set_name (which must be a 'STRING') is a material set.
- material_set(Set_name, R) [bf],[bb] True if material R is in permanent material set Set_name (a 'STRING'). With adornment bb, can also be the argument to insert and delete.
- next(M, Step_kind1, Step_kind2) [bbb] True iff the step history of material M contains a step of Step_kind1 and the most recent step of kind Step_kind1 in the history is not followed by a step of kind Step_kind2. The value of next is false if the most recent steps of both kinds have the same when value
- not(term1,..., termn) [b...b] True if term1,..., termn has no true bindings. Any bindings produced in evaluating term1,..., termn are undone before evaluating the term following the not term.
- only(*Variable1,...,Variablen*) (Adornment is irrelevant.) Cause the binding of *only* the argument variables to be printed. The effects of the side-effecting predicate persist during the evaluation of the query. Multiple evaluations cause the union of the variables in all the evaluations to be printed.
- pcr_product_length(*M*,*Length*) [bf] A special-purpose predicate for the CGR. *Length* becomes bound to the PCR product size (in base pairs) of *M* (a 'MATERIAL_POINTER') according to the rules detailed below.

(Also see left_primer_sequence, and right_primer_sequence.)

pcr_product_length binds Length to either:

- The value associated with the most recent product_length tag in *M*'s history, provided that there is no more or equally recent left_start or right_start tag.
- Otherwise the PCR product length computed from the most recent left_start and right_start tags in *M*'s history.

pcr_product_length binds Length to -1 if there is a left_start (or right_start) tag at least as recent as the most recent product_length tag, but no right_start (or, respectively, left_start) tag. pcr_product_length signals an incompatible database schema by the usual LabBase error mechanism (which aborts all query processing for the current query). pcr_product_length does *not* treat as an error nonsensical values for left_start and right_start; for example if right_start is less than left_start pcr_product_length will silently bind Length to a non-positive value.

• polymorphic(Avg_allele_sizes, I0, I1, Delta) [bbbb] Avg_allele_sizes is a list of integers, each at least -1. Yield true if the I0th and I1th (0-based) elements of Avg_allele_sizes differ by at least Delta and neither element is -1, or if one but not both of I0 and I1 is -1. (See also strictly_polymorphic).

- reversec(X, Y) [bf] [fb] Bind the free argument to the reverse complement of the bound argument. The bound argument must have type 'STRING' or 'DNA_SEQUENCE', and an error is reported if a 'STRING' argument contains any character other than A, T, G, C, or N. The free argument becomes bound to a value of type 'DNA_SEQUENCE'.
- right_primer_sequence(*M*, *Primer_sequence*) [bf] A special-purpose predicate for the CGR. Documented under left_primer_sequence.
- see_deleted_steps [b] Make logically deleted steps visible for the remainder of the current query.
- strictly_polymorphic(Avg_allele_sizes, I0, I1, Delta) [bbbb] Avg_allele_sizes is a list of integers, each at least -1. Yield true if the I0th and I1th (0-based) elements of Avg_allele_sizes differ by at least Delta and neither element is -1. (See also poly-morphic).
- substring(String,Start,Length,Substring) [bbbf] Bind Substring to the substring of String starting at (0-based) index Start and of length Length. It is an error if any part of the specified Substring falls outside of String. String must be bound to a value of type 'STRING' or 'DNA_SEQUENCE', and Substring becomes bound to a value of the same type as that of String.
- tag_value(*step*,*List*) [bf] Bind *List* to a list containing all the tag-and-value pairs (each represented as a list) in 'STEP_POINTER' *step*.
- tag_value(material,List) [bf] Bind List to a list containing all the most-recent tag-and-value pairs (each represented as a list) in 'MATERIAL_POINTER' material.
- temp_material_set(*Set_name*) [f],[b] True if *Set_name* (which must be a 'STRING') is a temporary material set.
- temp_value_set(*Set_name*, *R*) [bf],[bb] Same as material_set, except that the material set dissappears at the end of a database session—that is, when lbserv is used to shut down the database server.
- undelete_step(*Step*) [b] Undo the logical deletion of *Step*. (No error is raised if *Step* is not logically deleted.)
- var(X) [b],[f] True iff X is an unbound variable.

4.3 Built-In Tags and Step Kinds

Built-In Tags Certain tags are required by LabBase, and are built-in. Some are *data-dictionary tags*, because they are tags in data-dictionary entries. These are dd_identifier, type, dd_tag, id_tag. The other built-in tags are: who, when, created_material, material.

Built-In Step Kinds The step kind create is built-in.

4.4 Proposed but Unimplemented Predicates

LabBase users who require predicates not available above should contact:

steve@genome.wi.mit.edu.

5 Grammar

The grammar of the LabBase query language is similar to those of standard Prologs, with some exceptions. The most notable exceptions are:

- LabBase does not support the definition of rules.
- All infix operators are right associative and all have the same precedence, and parentheses cannot be used to group. When in doubt, use the parenfix syntax: for example, ;(3 < 5, foo(X)) rather than 3 < 5 ; foo(x) (equivalent to <(3,;(5,foo(x)))). The infix operator = plays a special role in LabBase, in arguments to the insert predicate and in certain queries based on step kinds. It is not possible to define new operators in the LabBase query language; they are all built in.
- Beware of the following:

3 < 4.

The term above compares the integer 3 with the *float* 4. The dot does *not* end a query.

5.1 Lexical Elements

Null characters (ASCII code 0) are illegal anywhere in the input. Control-A (ASCII code 1) is used as a synchronization character between clients and the LabBase server: it terminates the current query in any context. The perl API sees to it that each query sent from a client is terminated with a control-A.

5.1.1 String Literals

A string literal is one of the following:

- A (maximal) sequence of the characters [a-zA-ZO-O_] beginning with a lower-case letter ([a-z]).
- A sequence of the characters from the set $+-*\^<>=`~:.?@#$&.$
- A sequence of characters enclosed in single quotes.

Escapes are available within quoted strings: t, n, b expand to tab, newline, and back space, respectively, c causes the whitespace character c to be ignored, for example 'ab c' is another way of writing 'abc'. Every other character preceded by a i is replaced by itself, so 'foo' is another way of writing 'foo'. To write a the string ab'c write 'ab''c' or 'ab'c'. Octal escapes are not yet implemented—you cannot write '1' to get a string consisting of the character control-A, but they can be implemented quickly on request.

6 User Extension

It is possible to extend LabBase by adding new builtin predicates or new types to LabBase executable. Please refer to [7] for an overview of LabBase's system architecture before reading the material in this section.

6.1 Adding New Builtin Predicates

If you want to add a builtin that takes terms as arguments, you can find examples in builtin1.C. The remainder of this section discusses a simple interface for adding builtins that use only values (as opposed to terms) as input or output arguments. Examples can be found in builtin2.C.

To add a builtin predicate one must write a single function if the predicate produces no more than 1 set of bindings for a single input, or two functions if the predicate can produce more than 1 set of bindings for a single input.

To write a single-function predicate:

- 1. Let *first* be the name of the function implementing the predicate. (The name of the predicate is determined later, and need not be the same.) This function will set up the first bindings (if any). It returns an object of type QL_Iterator_State*, but a 1-function predicate should return 0 or 1 (suitably cast), to indicate whether the predicate is true or not.
- 2. Put the following line in a header file to be included by dd_ops.C (and which must also include builtin.H).

```
BUILTIN_FIRST_EXTERN(first);
```

3. In the file that will contain the function definition, create a static, file-global array variable, *types*, that contains the types expected of an initial segment of the bound arguments:

```
static Value::tv_type_id types[] = {Value::T1,...,Value::Tk};
```

Each Ti is a type from the enum tv_type_id defined in value.H. The variable types must contain at least one value.

4. Use the macro (defined in builtin.H): BUILTIN_FIRST_DEFINITION(*first*, bound_number, free_number, types, type_number, unused)
(

$\dot{f}unction_body$

The arguments to BUILTIN_FIRST_DEFINITION are:

- (a) first The C++ identifier for a global function to be created.
- (b) *bound_number* The number of bound arguments expected. Use -1 to indicate a variable number of bound arguments.
- (c) *free_number* The number of free arguments expected. Use -1 to indicate a variable number of free arguments.
- (d) types The array of Value::tv_type_ids defined above. Use Value::UNDEFINED to indicate that any type is acceptable. Otherwise an error will be generated if the corresponding bound argument is of a different type.
- (e) type_number The number of elements in types, which must at least 1 unless bound_number=0. If type_number is less than the number of bound arguments found at run time, the last element in types is taken to specify the required type of the remaining bound arguments.
- (f) function_body Computes whether the predicate is true or false, and produces bindings for any unbound arguments. The following variables are available in function_ body:
 - i. QL_Term* t is the term being evaluated; t would be the first argument in a call to ql_eval_error, which is how *first* should report a user error.
 - ii. int m is the number of bound arguments.
 - iii. int n is the number of free arguments.
 - iv. const Value in_tv[] has length m. The contents are the bound arguments. It is guaranteed that each element of in_tv is the address of a legal Value.
 - v. Value out_tv[] has length m. The elements of this array must be assigned to create the update bindings if *first* returns a non-0 result.
- 5. Add the following line to function DD_Ops::add_builtins() in dd_ops.C:

add_builtin(predicate_name, predicate_adornment, first);

where

- *predicate_name* is of type **char***, and is the name by which the predicate will be known in the LabBase query language.
- predicate_adornment is either the C++ string "*", indicating a variadic predicate, or a (possibly empty) string accepted by the regular expression [bf]*, which denotes the expected adornment of a predicate of fixed arity.
- *first* is the C++ identifier discussed above.

- 6. Garbage Collection Values produced using the static member functions *Value_Class*::make are automatically garbage collected. The constructors for Value classes are not public.
- 7. Reporting User Errors Use ql_eval_error(QL_Term*, char*m1,...) to report user errors. (There can be up to a total of 5 char* arguments.) The error message reported to the user is the concatenation of the char* arguments, followed by the text " predicate 'name' near line line_number, column column_number". As a consequence, most error messages should end in a preposition, such as "in" or "for". For example, calling ql_eval_error with char* arguments "a total " and "disaster occurred in", the user will see ERROR=a total disaster occurred in predicate name near line line_number, column_number. Do not end the error message with a period.
- 8. Stick with class Value You should be able to do everything you need to do in class Value. For most elements, X of the enum Value::tv_type_id there is a member function down_to_X that should be used for down-casts. These functions will call abort if the downcast is illegal based on the type of the Value. Also, for collection classes (lists and sets), stick with the class TV_Collection_Value, (which is a Collection_Value of Values). Lists and sets have the same C++ class! but do differ in how they respond to Value::tv_type(), and some member functions raise run-time errors if invoked on sets as opposed to lists (à la Smalltalk).

To write a two-function predicate, follow all steps for writing a one-function predicate, except step 5. In addition:

- 1. Let *next* be the name of the second function implementing the predicate. The *first* function must return some state that the **next** function receives as an argument, and that allows *next* to set up the next set of bindings (if any).
- 2. Put the following line in a header file to be included by dd_ops.C (and which must also include builtin.H).

BUILTIN_NEXT_EXTERN(next);

3. Place the macro

BUILTIN_NEXT_DEFINITION(next, unused) { function_body }

in the same file as the use of the macro BUILTIN_FIRST_DEFINITION. Within *function_body* the variables t, n, and out_tv have the same types and semantics as for the *function_body* argument of BUILTIN_FIRST_DEFINITION. In addition, variable s has type QL_Iterator_State*, and contains state information returned by a call to *first* or a previous call to *next*.

4. Add the following line to function DD_Ops::add_builtins() in dd_ops.C: add_builtin(predicate_name, predicate_adornment, first, next); where

- predicate_name, predicate_adornment, and first are as for the 1-function case, and
- next is the C++ identifier introduced above.

Please add appropriate regression tests to the directory \$LABBASE_ROOTDIR/src/lbback/ tests, and add a description of the new builtin to this document. Regression tests end in extension .test, and the baseline output is stored files with the same name and extension .last.

6.2 Adding New Types

This section is still *incomplete* and has not been debugged much. Please contact the authors for help in adding a new type.

All new types must be derived (in the C++ sense) from class Value. Let the new class be New_Class, and let the corresponding enum value for Value::tv_type_id be NEW_CLASS. For consistency with existing LabBase types, the enum name should be all upper-case, since the LabBase user will see this as the type name.

- In value.H:
 - Add the forward declaration "class New_Class;" just before the definition of class value.
 - In the definition of class Value:
 - * Add the enum value named *NEW_CLASS* to the definition of enum tv_type_id.
 - * Add the declaration "New_Class *down_to_NEW_CLASS() const;"
 - Add the necessary declaration of *New_Class*. This declaration should offer the following public member functions:
 - * compare [more to come]
 - * print [more to come]
 - * zero [more to come]
 - * make [more to come]
 - * save [more to come]
 - * [more to come]
- In value.C:
 - Define the function down_to_New_Class() by the top-level macro call DEFINE_ DOWNTO_FUNCTION(STEP_POINTER,Step)
 - Add the macro call V_PRINT(*New_Class*) to the body of Value::print().
 - Add the macro call V_SAVE(*New_Class*) to the body of Value::save() (assuming you want to save values of type *New_Class* in the database).
 - Add the macro call V_COMPARE(*New_Class*) to the body of Value::compare().

- Add an appropriate branch to the switch in the body of delete_value() to call the correct destructor for New_Class.
- As necessary, add definitions for the member functions of *New_Class*. (These definitions can also be in a separate file, of course.)
- In dd_ops.C:
 - Add the appropriate call to add_type_constructor to the body of DD_Ops::add_ type_constructors, to enable lbback to parse type expressions. The optional second argument to add_type_constructor, if non-0, indicates that the new type is in fact a type constructor that takes arguments.
- In ql_term.C: Fix the type-checking routines as necessary.
- In the implementations of builtin predicates: If any existing builtin predicates should operate on the new type, their implementations must be changed.

You will probably want to add a new set of regression tests for the new type.

References

- [1] W. F. Clocksin and C. S. Mellish. Programming in Prolog. Springer-Verlag, 1987.
- [2] N. Goodman. An object oriented DBMS war story: Developing a genome mapping database in C++. In W. Kim, editor, Modern Database Management: Object-Oriented and Multidatabase Technologies. ACM Press, 1994.
- [3] N. Goodman, M.-P. Reeve, and L. Stein. The design of MapBase: An object oriented database for genome mapping, Dec. 1992. Whitehead Institute for Biomedical Research, technical report.
- [4] N. Goodman, S. Rozen, and L. Stein. Requirements for a deductive query language in the MapBase genome-mapping database. In R. Ramakrishnan, editor, Applications of Logic Databases, pages 259-278. Kluwer, 1994. Available at ftp://genome.wi.mit.edu/ pub/papers/Y1994/requirements.ps.
- [5] A. R. Kerlavage, M. D. Adams, J. C. Kelly, M. Dubnick, J. Powell, P. Shanmugam, J. C. Venter, and C. Fields. Analysis and management of data from high-throughput expressed sequence tag projects. In T. N. Mudge, V. Milutinovic, and L. Hunter, editors, *Proceedings of the 26th Annual Hawaii International Conference on System Sciences*, volume 1, pages 585-594. IEEE Computer Society Press, Jan. 1993.

- [6] A. R. Kerlavage, W. FitzHugh, A. Glodek, J. Kelley, J. Scott, R. Shirley, G. Sutton, M. Wai-Chiu, O. White, and M. D. Adams. Data management and analysis for high-throughput DNA sequencing projects. *IEEE Engineering in Medicine and Biology*, Nov./Dec. 1995.
- [7] S. Rozen, L. Stein, and N. Goodman. Constructing a domain-specific DBMS using a persistent object system. In M. Atkinson, V. Benzaken, and D. Maier, editors, *Persistent Object Systems*. Springer-Verlag and British Computer Society, Workshops in Computing Series, 1995. Presented at POS-VI, Sep. 1994. Available at ftp://genome.wi.mit.edu/ pub/papers/Y1994/labbase-design.ps.Z.
- [8] S. Rozen, L. Stein, and N. Goodman. LabBase: A database to manage laboratory data in a large-scale genome-mapping project. *IEEE Engineering in Medicine and Bi*ology, 14(6):702-709, Nov./Dec. 1995. Preprint available at ftp://genome.wi.mit.edu /pub/papers/Y1995/labbase.ps.gz.
- [9] L. Stein, S. Rozen, and N. Goodman. Managing laboratory workflow with LabBase. In Proceedings of the 1994 Conference on Computers in Medicine (CompMed94). World Scientific Publishing Company, 1995. In press. Available at ftp://genome.wi.mit.edu /pub/papers/Y1995/workflow.ps.Z.