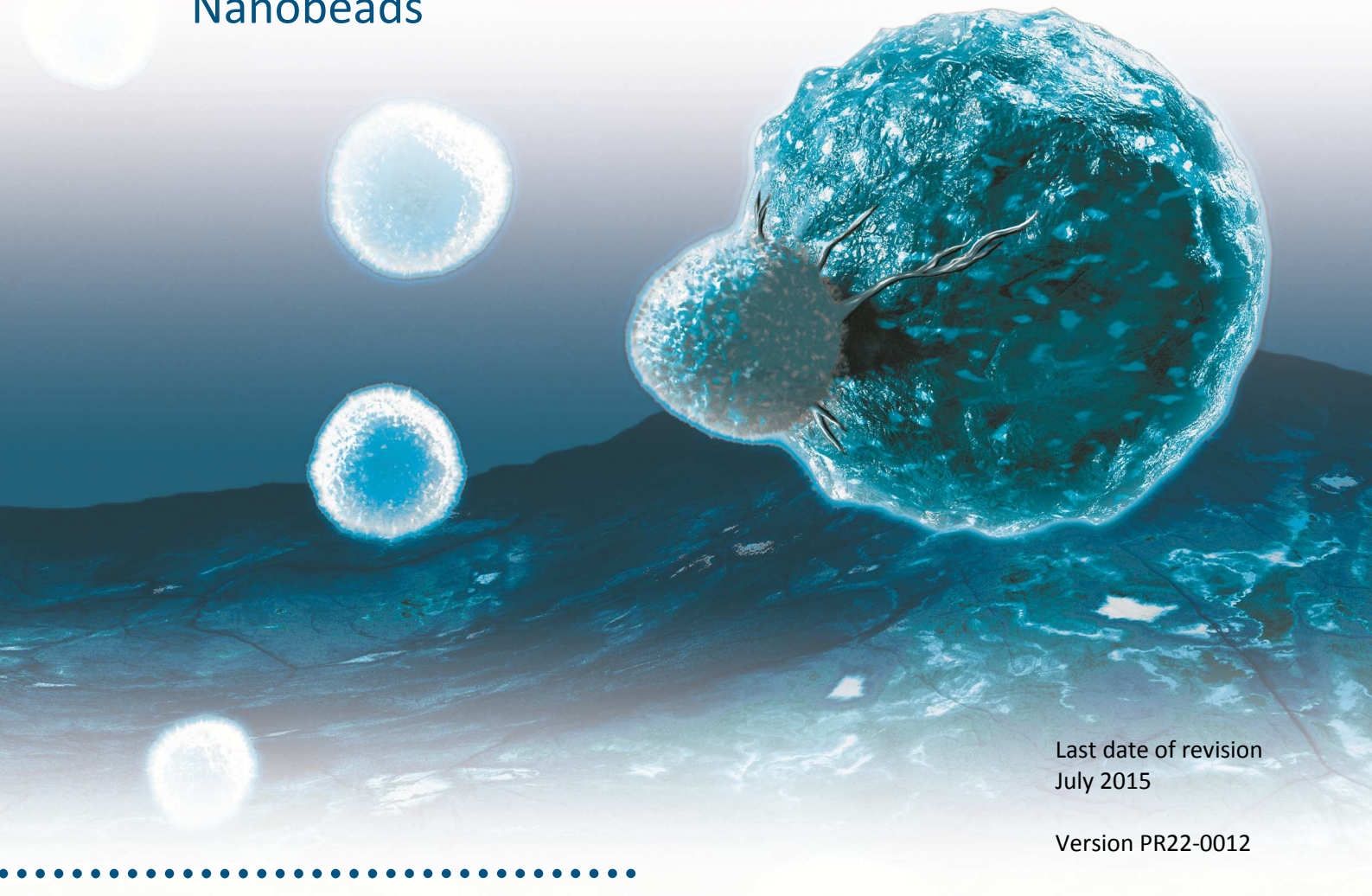


MHC I *Streptamer*[®] Manual

- I) Staining of antigen-specific CD8⁺ T cells with reversible MHC I *Streptamers*[®] and FACS isolation
- II) Isolation of antigen-specific CD8⁺ T cells with reversible MHC I *Streps* and *Strep-Tactin*[®] Magnetic Nanobeads



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1 The *Streptamer*[®] Principle

Strep-tag[®], *Strep-Tactin*[®] and *Streptamer*[®]

Strep-tags are short peptides with high binding selectivity for *Strep-Tactin*[®], an engineered streptavidin. The binding affinity of e.g. *Strep-tag II* to *Strep-Tactin*[®] (K_D = 1 μM) is nearly 100 times higher than to streptavidin. *Strep-tags* may be fused to recombinant proteins which allows efficient one-step purification of such fusion proteins on immobilized *Strep-Tactin*[®] under physiological conditions, thus preserving their bioactivity. As the *Strep-tag* binds to the biotin binding pocket of *Strep-Tactin*[®], purified proteins may be mildly eluted from the column by the addition of minute amounts of biotin.

Further information is available at www.strep-tag.com.

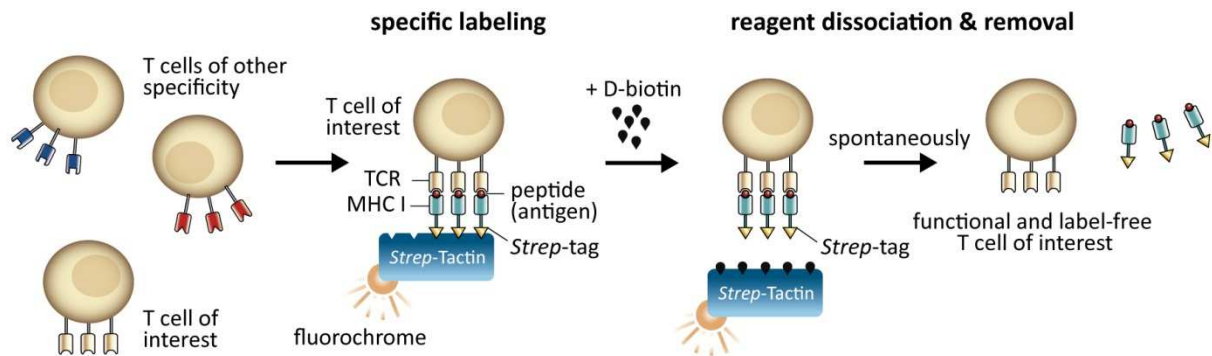
A special application of the *Strep-tag*[®]:*Strep-Tactin*[®] technology is the oligomerization of MHC I-*Strep-tag*[®] fusion proteins (MHC I-*Strep* proteins) on *Strep-Tactin*[®]. Multimers of MHC I-*Streps* complexed with either fluorescently or magnetically labeled *Strep-Tactin*[®], so-called *Streptamers*, are used for efficient staining or isolation of antigen-specific T cells. After separation of the labeled T cells from non-labeled cells by flow-cytometric or magnetic cell isolation, the *Streptamers* are efficiently disrupted on the cell by addition of biotin. Subsequently, the dissociation and removal of the *Strep-Tactin*[®] backbone leaves monomeric MHC I-*Strep* proteins on the surface of the T cell. As the interaction of monovalent MHC I:T cell receptor is weak, MHC I-*Strep* proteins spontaneously dissociate from the T cell receptor and may be removed from the T cells simply by washing. Keeping cells at cooled conditions as well as performing the rapid and complete removal of the *Streptamers*[®] from the T cells assures the isolation of fully functional, non-induced T cells.

Further information is available at www.streptamer.com.

2 Reversible staining of antigen-specific CD8⁺ T cells with MHC I *Streptamers*[®] and FACS isolation

2.1 Introduction: cell staining and removal of staining reagents

Scheme of a fluorescent *Streptamer*[®] labeled T cell and subsequent biotin induced removal of the *Streptamers*[®] to yield a functional, non-induced antigen specific T cell preparation.



2.2 Required reagents and materials

2.2.1 MHC I-*Streps* and *Strep-Tactin*[®] PE or APC

Cat.no	Product Name	Staining of	Size
6-7XXX-005	MHC I- <i>Strep</i>	2.5x10 ⁸ cells	0.2 ml
6-5000-005	<i>Strep-Tactin</i> [®] PE for MHC I <i>Streptamers</i> [®]	2.5x10 ⁸ cells	0.25 ml
6-5010-005	<i>Strep-Tactin</i> [®] APC for MHC I <i>Streptamers</i> [®]	2.5x10 ⁸ cells	0.25 ml

2.2.2 *Streptamer*[®] Solution Set and pre-separation filters

Cat.no.	Product Name	Content
6-5603-005	<i>Streptamer</i> [®] Solution Set Standard	Buffer IS, D-Biotin

The *Streptamer*[®] Solution Set Standard contains 50 ml Buffer IS as 10x concentrate for washing, and 1 ml of a D-Biotin stock solution (100 mM) for dissociation of the *Streptamers*[®] from the isolated cells. **Buffer IS has to be diluted with 9 volumes of water prior to use.** We recommend to add EDTA at a final concentration of 1 mM. Degas buffer before use. **The 100 mM Biotin stock solution has to be diluted with 99 volumes of Buffer IS prior to use (Biotin working solution is 1 mM; see 2.6.).**

Pre-separation filters (IBA GmbH, cat.-no.: 6-5601-010) are recommended for removal of cell clumps.

2.3 Use and storage of MHC I-*Streps* and fluorescent *Strep-Tactin*[®]

MHC I-*Streps* are shipped on dry ice and then stored at -80°C until use. After initial thawing prepare aliquots for long-term storage at -80°C. Aliquots for immediate use should be kept permanently on ice. **Aliquotation is mandatory to avoid freeze thaw cycles which denature the MHC I-*Streps*.**

Strep-Tactin[®] PE or APC for MHC I *Streptamers*[®] is shipped on blue ice and stored at 4°C.

2.4 Staining procedure

The procedure is optimized for staining of antigen-specific CD8⁺ T cells from fresh or frozen peripheral blood mononuclear cells (PBMCs). When working with anti-coagulated peripheral blood or buffy coats, PBMCs should first be isolated by density gradient centrifugation and separated from platelets.

Please adjust cell density to 10⁷ cells / 100 µl before starting the protocol.

Important: All steps have to be performed at 4°C! Please make sure that all your reagents and the cells have reached the temperature before starting the protocol. Protect labeled cells and fluorochrome reagents from light by incubating in the dark.

Protocol for staining of ca. 5x10⁶ cells (1 test):

1. Prepare ca. 3 ml Buffer IS from 10 x stock.
2. Incubate 5 µl *Strep-Tactin*[®] -PE or APC and 4 µl MHC I-*Strep* in a final volume of 50 µl Buffer IS for 45 minutes.
3. Add the pre-incubated *Streptamers*[®] (complex from *Strep-Tactin*[®] -PE (or APC) and MHC I-*Strep*, step 2) to the cell pellet.
4. Incubate for 45 minutes.
5. Wash cells twice with 200 µl Buffer IS.
6. Cells are ready for FACS-analysis or FACS-sorting.

Dead cell exclusion is strongly recommended (e.g. propidium iodide, 7-AAD, etc.)

2.5 Titration (optional)

If the staining protocol is not suitable for your application, a titration of the *Streptamers*[®] should be performed. Our recommendation for the titration is:

Keep the cell concentration of 10^7 cells / 100 μ l constant and increase the amount of *Streptamers*[®] stepwise (2-, 3- and 4-fold). Add the following volumes of pre-incubated (45 min) *Streptamers*[®] to your cell pellet:

- 2-fold increase (8 μ l MHC I-*Strep* + 10 μ l *Strep*-Tactin[®] PE in 100 μ l buffer IS)
- 3-fold increase (12 μ l MHC I-*Strep* + 15 μ l *Strep*-Tactin[®] PE in 150 μ l buffer IS)
- 4-fold increase (16 μ l MHC I-*Strep* + 20 μ l *Strep*-Tactin[®] PE in 200 μ l buffer IS)

The assay can be conducted in a 96-well round bottom microplate.

2.6 Dissociation of *Streptamers*[®] with D-Biotin

Important: All steps have to be performed at 4°C! Please make sure that all your reagents and the cells have reached the temperature before starting the protocol.

1. Collect cells by centrifugation.
2. Prepare ca. 1 ml Buffer IS containing 1 mM D-Biotin (1 mM Biotin working solution).
3. Resuspend cells in 200 μ l Biotin working solution and incubate for 10 minutes.
4. Collect cells by centrifugation.
5. Repeat step 3 and 4.
6. Wash cells 4 times with 200 μ l Buffer IS.
7. Transfer cells into the appropriate buffer or medium for further applications.

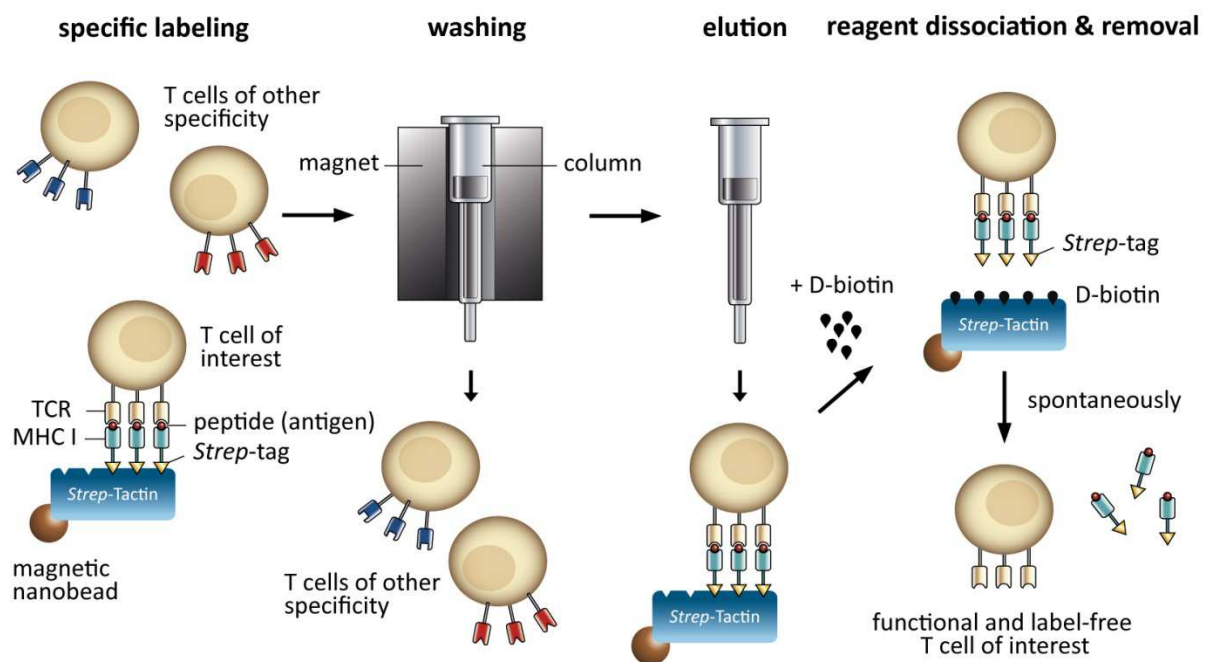
2.7 Short Protocol

Please request a copy of our Short Protocol PR38 for MHC I *Streptamer*[®] Staining at info@streptamer.com or download it from www.streptamer.com

3 Magnetic isolation of antigen-specific CD8⁺ T cells with MHC I *Streptamers*[®]

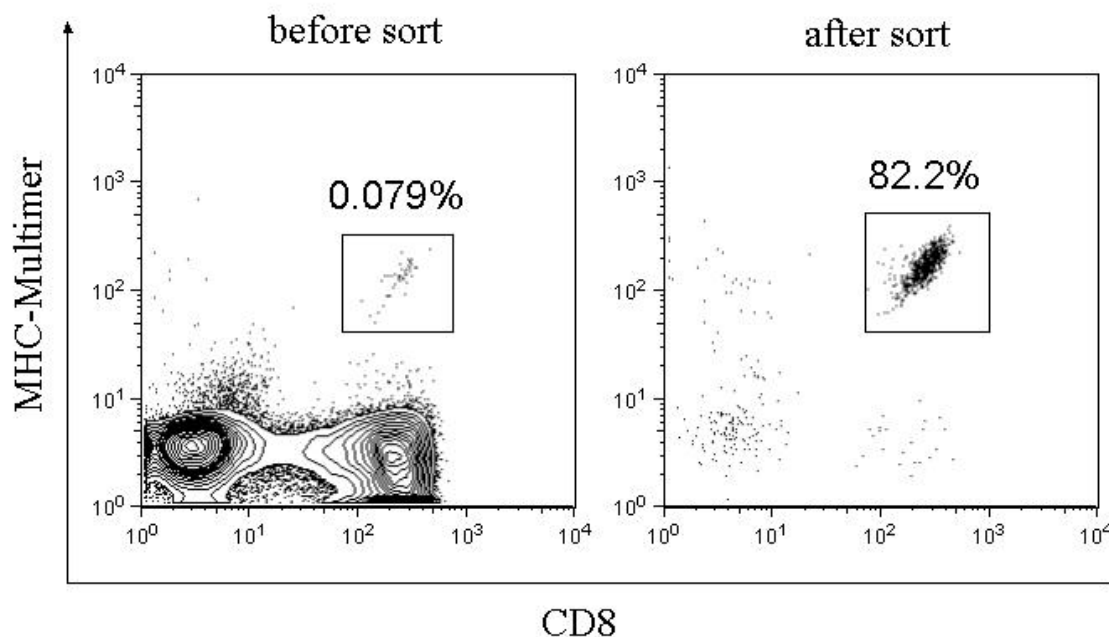
3.1 The principle: magnetic isolation and removal of labeling reagents

CD8⁺ T cells are labeled according to their antigen specificity with *Strep-Tactin*[®] Magnetic Nanobeads coupled to the specific MHC I-*Strep*. Labeled cells are separated from other cells by a magnetic field and retained target cells are eluted after removal of the magnet. All *Streptamer*[®] reagents are then released from the target cells by the addition of biotin (vitamin H) to yield a functional, non-induced antigen specific CD8⁺ T cell preparation.



Example: Isolation of CMV-antigen specific T cells

Isolation of human antigen specific T cells from PBMC with MHC I *Streptamers*[®]. Antigen-specific T cells were positive-selected with *Strep-Tactin*[®] Magnetic Nanobeads for MHC I *Streptamers*[®], which were coupled to MHC I-*Strep* HLA-A*0201, CMV pp65₄₉₅₋₅₀₃. Magnetically labeled cells were isolated on a MACS column. Thereafter, the *Streptamer*[®] reagents were removed from the target cells with 1 mM D-Biotin and cells were analyzed by flow cytometry. Before sort the antigen-specific T cells represented only 0.079 % of the total cell population. After sort an enormous enrichment by a factor of 1.000 and a high purity of 82 % became evident. These two important aspects were achieved with positive selection in only one step!



human PBMCs, CMV+
MHC: HLA A2 pp65₄₉₅₋₅₀₃

3.2 *Streptamer*[®] reagents and magnetic columns for cell isolation

3.2.1 MHC I-*Streps* and *Strep-Tactin*[®] Magnetic Nanobeads

Cat.no	Product Name	Isolation from	Size
6-7XXX-005	MHC I- <i>Strep</i>	5 x 10 ⁸ cells human 2.5 x 10 ⁸ cells mouse	1 x 0.2 ml
6-5500-005	<i>Strep-Tactin</i> [®] Magnetic Nanobeads for MHC I <i>Streptamers</i> [®]	1 x 10 ⁸ cells	1 x 0.25 ml

3.2.2 *Streptamer*[®] Solution Set Standard for washing & dissociation

Cat.no.	Product Name	Content
6-5603-005	<i>Streptamer</i> [®] Solution Set Standard	Buffer IS, D-Biotin

The *Streptamer*[®] Solution Set Standard contains 50 ml Buffer IS as 10x concentrate for washing and 1 ml of a D-Biotin stock solution (100 mM) for dissociation of the *Streptamers*[®] from the isolated cells. **Buffer IS has to be diluted with 9 volumes of water prior to use. We recommend to add EDTA at a final concentration of 1 mM.** Degas buffer before use, as air bubbles may block the column. The 100 mM Biotin stock solution has to be diluted with 99 volumes of Buffer IS prior to use (**Biotin working solution is 1 mM**).

3.2.3 Magnetic columns and pre-separation filters

For magnetic separation we recommend the MS or LS columns with the MACS® Manual Separators or the AutoMACS® from Miltenyi Biotech GmbH.

For removal of cell clumps prior to column loading we recommend our pre-separation nylon filters (IBA GmbH, cat.no. 6-5601-010, includes 10 filters).

3.3 Use and storage of MHC I-*Streps* and magnetic beads

MHC I-*Streps* are shipped on dry ice and then stored at -80°C until use. After initial thawing prepare aliquots for long-term storage at -80°C. Aliquots for immediate use should be kept permanently on ice. **Aliquotation is mandatory to avoid freeze thaw cycles which denature the MHC I-*Streps*.**

Strep-Tactin® Magnetic Nanobeads for MHC I *Streptamers*® are shipped on blue ice and stored at 4°C (do not freeze).

3.4 Experimental procedure

The procedure is optimized for T cell isolation from 2×10^7 cells. For cell numbers higher than 2×10^7 we suggest a linear upscale of beads and MHC I-*Streps*.

Some cells like monocytes or natural killer cells may be co-purified due to their ability to bind MHC I; they can be depleted by density gradient centrifugation or CD8+ pre-selection prior to T cell isolation.

Important: All steps – the isolation of cells as well as the following dissociation of *Streptamers*® – have to be performed at 4°C. Please make sure that all your reagents and the cells have reached the temperature before starting the protocol.

Avoid foaming, which interferes with proper bead retention on the magnet!

3.4.1 Preparation of cells and *Streptamers*®

Human cells: The procedure is optimized to isolate antigen-specific CD8⁺ T cells from 2×10^7 freshly isolated or frozen peripheral blood mononuclear cells (PBMCs). When working with anti-coagulated peripheral blood or buffy coat, PBMCs should be isolated by density gradient centrifugation first.

Mouse cells: When working with cells from spleen or lymph node, be careful to resuspend cells completely. Other organ preparations may require protease digestion and/or gradient centrifugation. Mouse T cell separation protocol is established for 2×10^7 cells. Higher cell numbers require larger amounts of beads and MHC I-*Streps*.

Cell preparation (step 1-2):

1. Collect prepared cells and resuspend in 10 ml Buffer IS.
2. Pass cells through a nylon filter tube to remove cell clumps, which may clog the columns, and place cells on ice.

MHC I *Streptamer*[®] preparation for human cells (step 3):

3. Incubate 50 μ l *Strep*-Tactin[®] Magnetic Nanobeads, 8 μ l MHC I-*Strep*, and 90 μ l Buffer IS at least 45 minutes at 4°C (or overnight).
4. Proceed to 3.4.2

MHC I *Streptamer*[®] preparation for mouse cells (step 3):

3. Incubate 50 μ l *Strep*-Tactin[®] Magnetic Nanobeads, 16 μ l MHC I-*Strep* and 80 μ l Buffer IS at least 45 minutes at 4°C (or overnight).
4. Proceed to 3.4.2

3.4.2 Cell Labeling with *Streptamers*[®]

Purification of *Streptamers*[®]: removal of unbound MHC I-*Strep* (step 1-4):

1. Place MS column in the magnetic field and prepare column by rinsing with 2 ml Buffer IS.
2. Add 1 ml Buffer IS to *Streptamers*[®] (from 3.4.1 step 3, human or mouse) and load on MS column.
3. Wash MS column while in magnetic field with 2 ml Buffer IS to remove unbound MHC I-*Streps*.
4. Add 250 μ l Buffer IS to MS column and elute retained *Streptamers*[®] (beads with bound MHC I-*Streps*) outside the magnetic field into a fresh vial; firmly flush out the purified *Streptamers*[®] using the plunger supplied with the column.

Cell Labeling with purified *Streptamers*[®] (step 5-7):

5. Centrifuge cell suspension (300 x g) and resuspend the cells in 250 μ l purified *Streptamers*[®] from 3.4.2 step 4. Incubate 45 minutes on ice.
6. Add 1.5 ml Buffer IS, centrifuge cell and *Streptamer*[®] mixture and carefully wash with 2 ml Buffer IS to eliminate unbound magnetic beads, which may trap cells on the column unspecifically.
7. Resuspend cells in 2 ml Buffer IS. Proceed to 3.4.3. or 3.4.4.

3.4.3 Magnetic separation with LS column

8. Place LS column in the magnetic field and prepare column by rinsing with 3 ml Buffer IS.
9. Apply cell suspension (3.4.2 step 7) onto the column. Allow unlabeled cells to pass through.
10. Wash column with 3 x 3 ml Buffer IS, adding buffer each time once the column reservoir is empty.
11. Remove column from magnetic field, add 5 ml Buffer IS and flush out labeled cells (positive fraction) into a fresh vial by firmly applying the plunger supplied with the column.

To increase purity, the magnetically labeled fraction can be passed over a new MS column (for up to 10^7 labeled cells) or LS column (for up to 10^8 labeled cells).

3.4.4 Optional: Magnetic separation with the AutoMACS[®] separator

For detailed instructions on how to use the AutoMACS[®] please refer to the corresponding user manual. Choose program "Posseld" and collect positive cell fraction.

3.4.5 Dissociation of *Streptamers*[®] with D-Biotin

12. Centrifuge eluted cells (positive fraction), resuspend in 2 ml Buffer IS containing 1 mM D-Biotin and incubate for 10 minutes.
13. Repeat step 12.
14. Wash cells 3 times with 5 ml Buffer IS.

3.4.6 Staining of T cells with *Streptamers*[®]

Antigen-specific cells can be visualized by utilizing the same MHC I-*Strep* which was used for magnetic isolation in combination with *Strep*-Tactin[®]-PE or APC. Proceed as described under 2.4. When a combinatorial staining with antibodies (especially anti-CD3 or anti-CD8 mAbs) is desired, add the respective antibodies only for the last 20 min of the staining protocol (2.4.; in total 45min) to avoid interference with the *Streptamers*[®].

A live/dead discrimination is suggested.

3.4.7 Short Protocol

Please request a copy of our Short Protocol PR36 (Human) or PR37 (Mouse) for MHC I *Streptamer*[®] Isolation at info@streptamer.com or download it from <http://www.iba-lifesciences.com/technical-support.html>

4 References

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For more references please visit www.streptamer.com

5 Warranty

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