

# **Bordetella pertussis ELISA IgG / IgA Testkit**

**Order No.: EC115.00**

**Color Coding: silver**

**FOR IN VITRO DIAGNOSIS ONLY**

**Sekisui Virotech GmbH  
Löwenplatz 5  
65428 Rüsselsheim / Germany**

**Tel.: +49-6142-6909-0  
Fax: +49-6142-966613  
<http://www.sekisuivirotech.com>**



# Contents

|  |          |
|--|----------|
| <b>1. Intended Use .....</b>   | <b>3</b> |
| <b>2. Diagnostic Relevance.....</b>  | <b>3</b> |
| <b>3. Test Principle.....</b>  | <b>3</b> |
| <b>4. Package Contents (IgG and IgA Testkit) .....</b>                             | <b>3</b> |
| <b>5. Storage and Shelflife of the Testkit and the ready to use reagents .....</b> | <b>4</b> |
| <b>6. Precautions and Warnings .....</b>   | <b>4</b> |
| <b>7. Material required but not supplied.....</b>                                  | <b>4</b> |
| <b>8. Test Procedure .....</b>   | <b>4</b> |
| 8.1 Examination Material.....  | 4        |
| 8.2 Preparation of Reagents .....  | 5        |
| 8.3 Virotech ELISA Test Procedure.....   | 5        |
| 8.4 Usage of ELISA processors .....  | 5        |
| <b>9. Test Evaluation.....</b>   | <b>6</b> |
| 9.1 Test function control.....   | 6        |
| 9.2 Calculation of the Virotech Units (VE) .....                                   | 6        |
| 9.3 Interpretation Scheme IgG and IgA .....  | 6        |
| 9.4 Limits of the Test.....  | 6        |
| <b>10. Performance Data.....</b>   | <b>7</b> |
| 10.1 Analytical Sensitivity and Specificity .....                                  | 7        |
| 10.2 Prevalence (Expected Values).....   | 7        |
| 10.3 Intra-assay-Coefficient of Variation (Repeatability).....                     | 7        |
| <b>11. Literature .....</b>  | <b>7</b> |
| <b>12. Test Procedure Scheme .....</b>   | <b>8</b> |

## 1. Intended Use

---

The Bordetella pertussis ELISA is a screening test. It is intended for the qualitative and semiquantitative detection of IgG- and IgA-antibodies against PT and FHA in human serum.

## 2. Diagnostic Relevance

---

The main agent of the genus *Bordetella*, *B. pertussis*, causes the clinical picture of whooping-cough.

The Pertussis Toxin (PT) is of significant importance for the pathogenesis of whooping cough. It is a real exotoxin responsible for many physiological, immunological and pharmacological effects. In contrast to other exotoxins of the species *Bordetella*, that show high cross-reactivities in serum diagnostics, the Pertussis Toxin is high-specific (3). An important adherence protein for attaching *B. pertussis* to the mucosal cells of the respiratory tract is filamentous haemagglutinin (FHA). During primary infection, the IgM-antibodies can be detected at the earliest 5-10 days after the beginning of the convulsive stage and persist for 6-12 weeks; they are the expression of an acute disease. IgA-antibodies can be detected 11 days after disease started at the earliest and can persist 6-24 months. They are also developed in vaccinated adults during a natural re-infection (without clinical disease) and are therefore found in healthy adults as well. Infected infants up to an age of 12 months do usually not develop IgA antibodies against Pertussis Toxin. Infants between 1-4 years rarely develop IgA antibodies against Pertussis Toxin, at an age between 5-10 years they develop only very small concentrations of IgA antibodies against Pertussis Toxin (4). In this case the detection of specific IgM can be a notice for a recent infection (2). IgG antibodies occur 2-3 weeks after onset of the disease in the serum at the earliest. Re-infections are marked by increased antitoxin-IgG- and -IgA-antibodies as a rule. IgG- and secrete-IgA-antibodies are, beside the specific sensibilised T-lymphocytes, the carrier of the long-term-immunity (1).

The pertussis serology cannot replace antigen detection, but should be performed in addition. The anti-pertussis antibodies are produced later in comparison to other infectious diseases.

## 3. Test Principle

---

The antibody searched for in the human serum forms an immune complex with the antigen coated on the microtiter-plate. Unbound immunoglobulins are removed by washing processes. The enzyme conjugate attaches to this complex. Unbound conjugate is again removed by washing processes. After adding the substrate solution (TMB), a blue dye is produced by the bound enzyme (peroxidase). The color changes to yellow when the stopping solution is added.

## 4. Package Contents (IgG and IgA Testkit)

---

1. **1 Microtiter-Plate** consisting of 96 wells with antigen coated, breakable single wells, lyophilised
2. **PBS-Dilution Buffer (blue, ready to use) 2x50ml**, pH 7,2, with preservative and Tween 20
3. **PBS-Washing Solution (20x concentrated) 50ml**, pH 7,2, with preservative and Tween 20
4. **IgG negative Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
5. **IgG cut-off Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
6. **IgG positive Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
7. **IgA negative Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
8. **IgA cut-off Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
9. **IgA positive Control, 1300µl**, human serum with protein-stabilizer and preservative, ready to use
10. **IgG-Conjugate (anti-human), 11ml**, (sheep or goat)-horseradish-peroxidase-conjugate with protein-stabilizer and preservative in Tris-Buffer, ready to use
11. **IgA-Conjugate (anti-human), 11ml**, (sheep or goat)-horseradish-peroxidase-conjugate with FCS and preservative in Tris-Buffer, ready to use
12. **Tetramethylbenzidine substrate solution (3,3',5,5'-TMB), 11ml**, ready to use
13. **Citrate-Stopping Solution, 6ml**, contains an acid mixture

## 5. Storage and Shelflife of the Testkit and the ready to use reagents

---

Store the testkit at 2-8°C. The shelf life of all components is shown on each respective label; for the kit shelf life please see Quality Control Certificate.

1. Microtiter strips/single wells are to be resealed in package after taking out single wells and stored with desiccant at 2-8°C. Reagents should immediately be returned to storage at 2-8°C after usage.
2. The ready to use conjugate and the TMB-substrate solution are sensitive to light and have to be stored in dark. Should there be a color reaction of the substrate dilution due to incidence of light, it is not useable anymore.
3. Take out only the amount of ready to use conjugate or TMB needed for the test insertion. Additional conjugate or TMB taken out may not be returned but must be dismissed.

| Material                      | Status                        | Storage  | Shelflife |
|-------------------------------|-------------------------------|--|-----------|
| Test Samples                  | Diluted                       | +2 to +8°C   | max. 6h   |
|                               | Undiluted                     | +2 to +8°C   | 1 week    |
| Controls                      | After Opening                 | +2 to +8°C   | 3 months  |
| Microtitreplate               | After Opening                 | +2 to +8° (storage in the provided bag with desiccant bag) | 3 months  |
| Rheumatoid factor - Absorbent | Undiluted, After Opening      | +2 to +8°C   | 3 months  |
|                               | Diluted                       | +2 to +8°C   | 1 week    |
| Conjugate                     | After Opening                 | +2 to +8°C (protect from light)                            | 3 months  |
| Tetramethylbenzidine          | After Opening                 | +2 to +8°C (protect from light)                            | 3 months  |
| Stop Solution                 | After Opening                 | +2 to +8°C   | 3 months  |
| Washing Solution              | After Opening                 | +2 to +8°C   | 3 months  |
|                               | Final Dilution (ready-to-use) | +2 to +25°C  | 4 weeks   |

## 6. Precautions and Warnings

---

1. Only sera which have been tested and found to be negative for HIV -1 antibodies, HIV -2 antibodies, HCV antibodies and Hepatitis-B surface-antigen are used as control sera. Nevertheless, samples, diluted samples, controls, conjugates and microtiter strips should be treated as potentially infectious material. Please handle products in accordance with laboratory directions.
2. Those components that contain preservatives, the Citrate Stopping Solution and the TMB have an irritating effect to skin, eyes and mucous. If body parts are contacted, immediately wash them under flowing water and possibly consult a doctor.
3. The disposal of the used materials has to be done according to the country-specific guidelines.

## 7. Material required but not supplied

---

1. Aqua dest./demin.
2. Eight-channel pipette 50µl, 100µl
3. Micropipettes: 10µl, 100µl, 1000µl
4. Test tubes
5. Paper towels or absorbent paper
6. Cover for ELISA-plates
7. Disposal box for infectious material
8. ELISA hand washer or automated EIA plate washing device
9. ELISA plate spectrophotometer, wavelength = 450nm, reference length = 620nm (Reference Wavelength 620-690nm)
10. Incubator

## 8. Test Procedure

---

Working exactly referring to the Sekisui Virotech user manual is the prerequisite for obtaining correct results.

### 8.1 Examination Material

Either serum or plasma can be used as test material, even if only serum is mentioned in the instructions. Any type of anticoagulant can be used for plasma.

Always prepare patient-dilution freshly.

For a longer storage the sera must be frozen. Repeated defrosting should be avoided.

1. Only fresh non-inactivated sera should be used.
2. Hyperlipaemic, haemolytic, microbially contaminated and turbid sera should not be used (false positive/negative results).

## 8.2 Preparation of Reagents

The Sekisui Virotech System Diagnostica offers a high degree of flexibility regarding the possibility to use the dilution buffer, washing solution, TMB, citrate stopping solution as well as the conjugate for all parameters and for all different lots. The ready to use controls (positive control, negative control, cut-off control) are **parameter specific** and **only to use** with the plate lot indicated in the Quality Control Certificate.

1. Set incubator to 37°C and check proper temperature setting before start of incubation.
2. Bring all reagents to room temperature before opening package of microtiter strips.
3. Shake all liquid components well before use.
4. Make up the washing solution concentrate to 1 L with distilled or demineralised water. If crystals have formed in the concentrate, please bring the concentrate to room temperature before use and shake well before use.

## 8.3 Virotech ELISA Test Procedure

1. For each test run, pipette 100µl each of ready to use dilution buffer (blank), IgG- and IgA-positive, negative and cut-off controls as well as diluted patient sera. We propose a double insertion (blank, controls and patient sera); for cut-off control a double insertion is absolutely necessary. Working dilution of patient sera: 1+100; e.g. 10µl serum + 1ml dilution buffer.
2. After pipetting start incubation for 30 min. at 37°C (with cover).
3. End incubation period by washing microtiter strips 4 times with 350 – 400µl washing solution per well. Do not leave any washing solution in the wells. Remove residues on a cellulose pad.
4. Pipette 100µl of ready to use conjugate into each well.
5. Incubation of conjugates: 30 min. at 37°C (with cover).
6. Stop conjugate incubation by washing 4 times (pls. refer to point 3 above).
7. Pipette 100µl of ready to use TMB into each well.
8. Incubation of substrate solution: 30 min. at 37°C (with cover, keep in dark).
9. Stopping of substrate reaction: pipette 50µl of citrate stopping solution into each well. Shake plate carefully and thoroughly until liquid is completely mixed and a homogeneous yellow color is visible.
10. Measure extinction (OD) at 450/620nm (Reference Wavelength 620-690nm). Set your photometer in such a way that the blank value is deducted from all other extinctions. Extinctions should be measured within 1 hour after adding the stopping solution!

**Pls. refer to last page for Test Procedure Scheme**

## 8.4 Usage of ELISA processors

All Sekisui Virotech ELISAs can be used on ELISA processors. The user is bound to proceed a validation of the devices (processors) on a regular basis.

Sekisui Virotech recommends the following procedure:

1. Sekisui Virotech recommends to proceed the validation of device referring to the instructions of the device manufacturer during the implementation of the ELISA processor respectively after bigger reparations.
2. It is recommended to check the ELISA-processor with the Validationkit (EC250.00) afterwards. A regular check using the Validationkit shall be proceeded minimum once a quarter to test the accuracy of the processor.
3. The release criteria of the Quality Control Certificate of the product must be fulfilled for each test run.

With this procedure, your ELISA processor will function properly and this will support quality assurance in your laboratory.

## 9. Test Evaluation

The ready to use controls serve for a semiquantitative determination of specific IgG- and IgA-antibodies. Their concentration can be expressed in Virotech units = VE. Fluctuations resulting from the test procedure can be balanced with this calculation method and a high reproducibility is achieved in this way. Use the means of the OD values for calculation of the VE.

### 9.1 Test function control

#### a) OD-values

The OD of the blank should be < 0.15.

The OD-values of the negative controls should be lower than the OD-values mentioned in the Quality Control Certificate. The OD-values of the positive controls as well as of the cut-off controls should be above the OD-values mentioned in the Quality Control Certificate.

#### b) Virotech Units (VE)

The Virotech Units (VE) of the cut-off controls are defined as 10 VE. The calculated VE of the positive controls should be within the ranges mentioned in the Quality Control Certificate.

If those requirements (OD-values, VE) are not fulfilled, the test has to be repeated.

### 9.2 Calculation of the Virotech Units (VE)

The extinction of the blank value (450/620nm) has to be subtracted from all other extinctions.

$$\begin{aligned} \text{VE (positive control)} &= \frac{\text{OD (positive control)}}{\text{OD (cut - off control)}} \times 10 \\ \text{VE (patient serum)} &= \frac{\text{OD (patient serum)}}{\text{OD (cut - off control)}} \times 10 \end{aligned}$$

### 9.3 Interpretation Scheme IgG and IgA

| Result (VE) | Evaluation |
|-------------|------------|
| < 9,0       | negative   |
| 9,0 - 11,0  | borderline |
| > 11,0      | positive   |

1. If the measured values are above the defined borderline range, they are considered to be positive (please take notice of vaccination management!).
2. If the measured VE is within the borderline range, no significant high antibody concentration is present, the samples are considered to be borderline. For the secure detection of an infection it is necessary to determine the antibody concentration of two serum samples. One sample shall be taken directly at the beginning of the infection and a second sample 5 – 10 days later (convalescent serum). The antibody concentration of both samples has to be tested in parallel, that means in one test run. A correct diagnosis based on the evaluation of a single serum sample is not possible.
3. If the measured values are below the defined borderline range, no measurable antigen specific antibodies are present in the samples. The samples are considered to be negative.
4. It is recommended to use the Virotech Bordetella pertussis LINE Immunoblot to confirm a positive IgG- or positive IgA-result.

### 9.4 Limits of the Test

1. The interpretation of serological results shall always include the clinical picture, epidemiological data and all further available laboratory results.
2. Filamentous haemagglutinin is a group antigen which is also found in other agents of the species Bordetella (e.g. *Bordetella parapertussis*, *Bordetella bronchiseptica*) (5,6). A cross reactivity is therefore to be expected.

## 10. Performance Data

---

### 10.1 Analytical Sensitivity and Specificity

200 sera were tested in the Virotech Bordetella pertussis ELISA and then compared with the Virotech Pertussis Toxin ELISA to determine the analytical sensitivity and specificity. The sera collective comprises following sera:

25 sera from patients with suspected Bordetella pertussis infection

80 sera obtained from routine tests (respiratory diseases without suspicion of Bordetella pertussis)

95 blood donor sera

#### Sera collective (n= 200)

| Virotech Pertussis Toxin ELISA | Virotech Bordetella pertussis ELISA IgG |            |          | Virotech Bordetella pertussis ELISA IgA |            |          |
|--------------------------------|---|------------|----------|---|------------|----------|
|                                | Negative                                | Borderline | Positive | Negative                                | Borderline | Positive |
| Negative                       | 121                                     | 15         | 34       | 156                                     | 7          | 17       |
| Borderline                     | 0                                       | 0          | 8        | 6                                       | 2          | 2        |
| Positive                       | 0                                       | 0          | 22       | 2                                       | 0          | 8        |

Borderline results have not been considered for the calculation of the sensitivity and specificity. Referring to the Virotech Pertussis Toxin ELISA a sensitivity of >99,8 % resp. a specificity of 78,1 % for IgG and a sensitivity of 80,0 % resp. a specificity of 90,2 % for IgA have been obtained.

### 10.2 Prevalence (Expected Values)

The following table shows the results of the examination of 80 blood bank sera in IgG and 78 blood bank sera in IgA.

|            | IgG |      | IgA |     |
|------------|-----|------|-----|-----|
|            | No. | %    | No. | %   |
| Negative   | 54  | 67,5 | 74  | 95  |
| Borderline | 10  | 12,5 | 2   | 2,5 |
| Positive   | 16  | 20,0 | 2   | 2,5 |

### 10.3 Intra-assay-Coefficient of Variation (Repeatability)

In one assay, strips of different plates of one batch have been tested with the same serum sample. The obtained coefficient of variation for IgG is < 9% and for IgA is < 15%.

## 11. Literature

---

1. Wiersbitzky S. Pertussis Kostengünstige Prävention zu wenig genutzt Therapiewoche 25 (1995), p.1485 - 1486
2. Lehrbuch der Medizinischen Mikrobiologie, H. Brandis, W. Köhler, H.J. Eggers, G. Pulverer, 7. Auflage, p. 483
3. Mastrantonio et al., 1997, *Bordetella parapertussis* infections., Dev Biol Stand., (89):255-259
4. Wirsing von König et al., 1999, Evaluation of a single-Sample Serological Technique for Diagnosing Pertussis in Unvaccinated Children, Eur J Clin Microbiol Infect Dis, (18)341-345
5. Elisabeth Bergfors MD et al., Parapertussis and Pertussis: Differences and Similarities in Incidence, Clinical Course, and Antibody Responses, Intern. J. Infet. Dis., 3(3):1999
6. Jacob-Dubuisson F et al., Molecular characterization of *Bordetella bronchiseptica* filamentous haemagglutinin and its secretory machinery, Microbiology (2000), 146,1211-1221

## Preparation of Patient Samples and Washing Solution

▼ **Washing Solution:** Fill up concentrate to 1 liter with aqua dest./demin.

### **IgG/IgA-Samples – Dilution**

**1:101**

e.g.:

10 µl serum/plasma + 1000 µl Dilution Buffer

(Serum Dilution Buffer is ready to use)

## Testprocedure

|                      |                    |   |
|----------------------|--------------------|---|
| Samples Incubation   | 30 minutes at 37°C | <b>100 µl Patient Samples</b><br>blank value (Dilution Buffer) and controls |
| ↓                    |                    |   |
| Wash 4times          |                    | <b>400 µl Washing Solution</b><br>Remove Residues on a Cellulose Pad        |
| ↓                    |                    |   |
| Conjugate Incubation | 30 minutes at 37°C | <b>100 µl Conjugate</b><br>IgG, IgA   |
| ↓                    |                    |   |
| Wash 4times          |                    | <b>400 µl Washing Solution</b><br>Remove Residues on a Cellulose Pad        |
| ↓                    |                    |   |
| Substrate Incubation | 30 minutes at 37°C | <b>100 µl Substrate</b>   |
| ↓                    |                    |   |
| Stopping             |                    | <b>50 µl Stopping Solution</b><br>shake carefully                           |
| ↓                    |                    |   |
| Measure Extinctions  |                    | <b>Photometer at 450/620nm</b><br>(Reference Wavelength 620-690nm)          |