

# Magnetic Luminex<sup>®</sup> Performance Assay

## Human High Sensitivity Cytokine Base Kit B

Catalog Number LBHS000

For the simultaneous quantitative determination of multiple human cytokine concentrations in cell culture supernates, serum, and plasma.

**Note: IL-10 is no longer suitable for use in this assay. Please use Human High Sensitivity Cytokine Panel A to assay IL-10.**

This package insert must be read in its entirety before using this product.  
For research use only. Not for use in diagnostic procedures.

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## INTRODUCTION

Cytokines are intercellular signaling proteins released from a wide variety of cells and tissues. They play an integral role in regulating growth and cellular proliferation as well as modulating host response to infection, injury, and inflammation. Cytokines also influence reproduction and bone remodeling. A large number of cytokines are pleiotropic and share similar functions. In addition, many cytokines influence the production of other cytokines. Analysis and quantification of cytokines in biological fluids has thus become increasingly important. Methods such as bioassay, enzyme-linked immunosorbent assay (ELISA), intracellular staining, ribonuclease protection assay (RPA) and polymerase chain reaction (PCR) have all been used for quantifying cytokines, however, each of these techniques has limitations associated with it. These techniques are not capable of measuring multiple cytokines simultaneously in a limited sample volume.

Assessing the levels of multiple cytokines may be more revealing than analyzing a single protein. Quantifying multiple cytokines on an individual level can be time consuming and expensive. When combined with separately available analyte-specific microparticle sets, this kit is an excellent tool for simultaneously assessing the levels of multiple human cytokines in a single sample.

Any combination of the following microparticle sets are suitable for use with this base kit:

Analyte	Catalog Number	Microparticle Region
GM-CSF	LHSCM215	29
IFN- $\gamma$	LHSCM285	30
IL-1 $\beta$	LHSCM201	20
IL-2	LHSCM202	19
IL-5	LHSCM205	22
IL-6	LHSCM206	25
IL-7	LBHS207	33
IL-13	LBHS213	34
IL-15	LBHS247	35
IL-17A	LBHS317	36
IL-17F	LBHS1335	37
IL-22	LBHS5782	38
IL-23	LBHS1716	39
IL-31	LBHS2824	42
IL-33	LBHS3625	18
IL-36 $\beta$	LBHS1099	15
TNF- $\alpha$	LHSCM210	12

## PRINCIPLE OF THE ASSAY

Magnetic Luminex® Performance Assay multiplex kits are designed for use with the Luminex MAGPIX® CCD Imager. Alternatively, kits can be used with the Luminex 100/200™ or Bio-Rad® Bio-Plex®, dual laser, flow-based sorting and detection platforms.

Analyte-specific antibodies are pre-coated onto color-coded magnetic microparticles. Microparticles, standards and samples are pipetted into wells and the immobilized antibodies bind the analytes of interest. After washing away any unbound substances, a biotinylated antibody cocktail specific to the analytes of interest is added to each well. Following a wash to remove any unbound biotinylated antibody, streptavidin-phycoerythrin conjugate (Streptavidin-PE), which binds to the biotinylated antibody, is added to each well. A final wash removes unbound Streptavidin-PE, the microparticles are resuspended in buffer and read using the Luminex MAGPIX Analyzer. A magnet in the analyzer captures and holds the superparamagnetic microparticles in a monolayer. Two spectrally distinct Light Emitting Diodes (LEDs) illuminate the microparticle. One LED identifies the analyte that is being detected and the second LED determines the magnitude of the PE-derived signal, which is in direct proportion to the amount of analyte bound. Each well is imaged with a CCD camera.

## LIMITATIONS OF THE PROCEDURE

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- The kit should not be used beyond the expiration date on the kit label.
- Do not mix or substitute reagents with those from other lots or sources.
- If samples fall outside the dynamic range of the assay, further dilute the samples with the appropriate Calibrator Diluent and repeat the assay.
- Any variation in standard diluent, operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding.
- Variations in sample collection, processing, and storage may cause sample value differences.
- This assay is designed to eliminate interference by other factors present in biological samples. Until these factors have been tested in the Luminex Performance Assay, the possibility of interference cannot be excluded.
- Luminex Performance Assays afford the user the benefit of multianalyte analysis of biomarkers in a single complex sample. For each sample type, a single multipurpose diluent is used to optimize recovery, linearity, and reproducibility. Such a multipurpose diluent may not optimize any single analyte to the same degree that a unique diluent selected for analysis of that analyte can optimize conditions, therefore, some performance characteristics may be more variable than those for assays designed specifically for single analyte analysis.
- **Only the analytes listed on the Standard Value Card can be measured with this base kit.**

## TECHNICAL HINTS

- When mixing or reconstituting protein solutions, always avoid foaming.
- To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Protect microparticles and Streptavidin-PE from light at all times to prevent photobleaching.

## MATERIALS PROVIDED & STORAGE CONDITIONS

Store the unopened kit at 2-8 °C. Do not use past kit expiration date.

PART	PART #	DESCRIPTION	STORAGE OF OPENED, DILUTED, OR RECONSTITUTED MATERIAL
Standard Cocktail Human HS Cytokine Panel B	894489	2 vials of recombinant human cytokines in a buffered protein base with preservatives; lyophilized.	Discard after use. Use a fresh standard for each assay.
Diluent RD2-5	895982	2 vials (6 mL/vial) of a buffered protein base with blue dye and preservatives.	May be stored for up to 1 month at 2-8 °C.* <i>Once diluted, any unused microparticle cocktail must be discarded.</i>
Calibrator Diluent RD5K	895119	21 mL of a buffered protein base with preservatives. <i>For cell culture supernate samples.</i>	May be stored for up to 1 month at 2-8 °C.*
Calibrator Diluent RD6-65	895098	21 mL of a buffered protein base with preservatives. <i>For serum/plasma samples.</i>	
Streptavidin-PE	895613	5.5 mL of streptavidin-phycoerythrin conjugate with preservatives.	
Wash Buffer Concentrate	895003	21 mL of a 25-fold concentrated solution of buffered surfactant with preservative. <i>May turn yellow over time.</i>	
Microplate	641385	1 flat-bottomed 96-well microplate used as a vessel for the assay.	
Mixing Bottles	895505	2 empty 8 mL bottles used for mixing microparticles with Microparticle Diluent.	
Plate Sealers	640445	6 adhesive foil strips.	
Standard Value Card	749810	1 card listing the Standard reconstitution volume and working standard concentrations for this lot of base kit.	

\*Provided this is within the expiration date of the kit.

## OTHER SUPPLIES REQUIRED

- Luminex Performance Assay analyte-specific kit(s) (see Introduction on page 1).
- Luminex MAGPIX, Luminex 100/200, or Bio-Rad Bio-Plex analyzer with X-Y platform.
- Hand-held microplate magnet or platewasher with a magnetic platform.
- Pipettes and pipette tips.
- Deionized or distilled water.
- Multi-channel pipette, manifold dispenser, or automated dispensing unit.
- 500 mL graduated cylinder.
- Horizontal orbital microplate shaker (0.12" orbit) capable of maintaining a speed of 800 ± 50 rpm.
- Microcentrifuge.
- **Polypropylene** test tubes for dilution of standards and samples.
- Human HS Cytokine Panel B Controls (R&D Systems, Catalog # QC19; optional).

## PRECAUTIONS

Some components in this kit contain ProClin® which may cause an allergic skin reaction. Avoid breathing mist.

Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling. Please refer to the MSDS on our website prior to use.

## SAMPLE COLLECTION & STORAGE

**The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.**

**Cell Culture Supernates** - Remove particulates by centrifugation and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifuging for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at  $\leq -20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles.

**Note:** *Citrate plasma has not been validated for use in this assay.*

*Hemolyzed, icteric, and lipemic sample are not suitable for use in this assay.*

## SAMPLE PREPARATION

Serum and plasma samples require a 2-fold dilution. A suggested 2-fold dilution is 150  $\mu\text{L}$  of sample + 150  $\mu\text{L}$  of Calibrator Diluent RD6-65. Mix thoroughly.

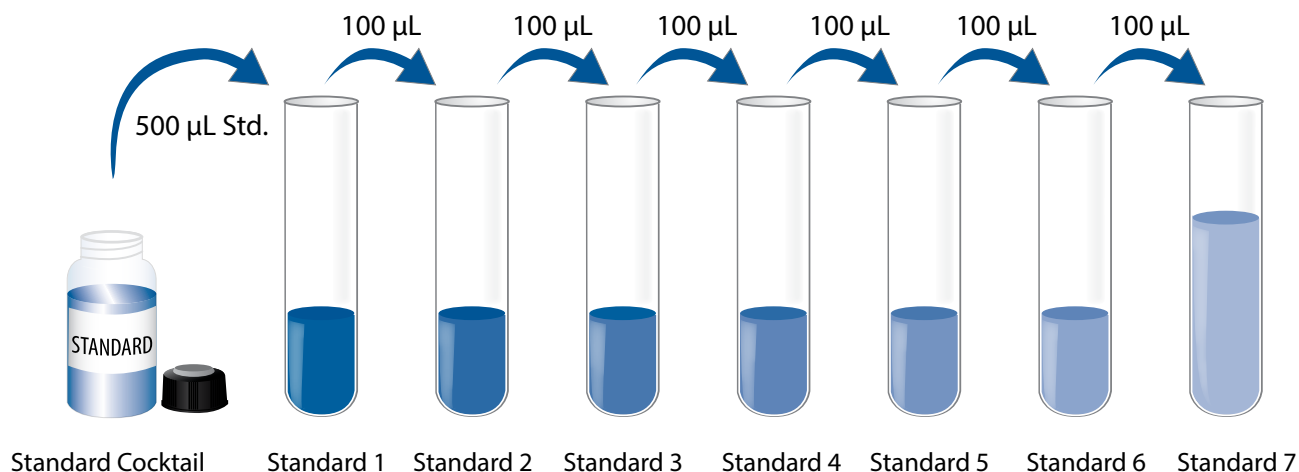
## REAGENT PREPARATION

**Bring all reagents to room temperature before use.**

**Wash Buffer** - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 20 mL of Wash Buffer Concentrate to deionized or distilled water to prepare 500 mL of Wash Buffer.

**Standard** - Reconstitute the Standard Cocktail with Calibrator Diluent RD5K (*for cell culture supernate samples*) or Calibrator Diluent RD6-65 (*for serum/plasma samples*). Refer to the Standard Value Card for the reconstitution volume and assigned values. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions.

**Use polypropylene tubes.** Pipette 500  $\mu$ L of the reconstituted Standard into the Standard 1 tube. Pipette 300  $\mu$ L of the appropriate Calibrator Diluent into the remaining tubes. Use Standard 1 to produce a 4-fold dilution series (below). Refer to *analyte specific datasheet for details*. Mix each tube thoroughly before the next transfer. Standard 1 serves as the high standard. The appropriate Calibrator Diluent serves as the blank.



## DILUTED MICROPARTICLE COCKTAIL PREPARATION

1. Centrifuge each Microparticle Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vials to resuspend the microparticles, taking precautions not to invert the vials.
3. Dilute the Microparticle Concentrates in the mixing bottle provided. The volume of the Microparticle Concentrate listed in the table below is for each analyte (e.g. if measuring a full plate of IL-1 $\beta$  and IL-6, add 50  $\mu$ L of IL-1 $\beta$  Microparticle Concentrate and 50  $\mu$ L of IL-6 Microparticle Concentrate to 2.5 mL of Diluent RD2-5).

Number of Wells Used	Microparticle Concentrate	+	Diluent RD2-5
96	50.0 $\mu$ L	+	2.50 mL
72	37.5 $\mu$ L	+	1.875 mL
48	25.0 $\mu$ L	+	1.25 mL
24	12.5 $\mu$ L	+	0.625 mL

**Note:** *Protect microparticles from light during handling. Diluted microparticles cannot be stored. Prepare microparticles within 30 minutes of use.*

## DILUTED BIOTIN ANTIBODY COCKTAIL PREPARATION

1. Centrifuge each Biotin Antibody Concentrate vial for 30 seconds at 1000 x g prior to removing the cap.
2. Gently vortex the vials, taking precautions not to invert the vials.
3. Add 50  $\mu$ L of each Biotin Antibody Concentrate to one vial of the Diluent RD2-5. Mix gently.



## INSTRUMENT SETTINGS

### Luminex MAGPIX analyzer:

- a) Assign the microparticle region for each analyte being measured (see page 1)
- b) 50 events/bead
- c) Sample size: 50  $\mu$ L
- d) Collect Median Fluorescence Intensity (MFI)

### Luminex 100/200 and Bio-Rad Bio-Plex analyzers:

**Note:** *Calibrate the analyzer using the proper reagents for superparamagnetic microparticles (refer to instrument manual).*

- a) Assign the microparticle region for each analyte being measured (see page 1)
- b) 50 events/bead
- c) Minimum events: 0
- d) Flow rate: 60  $\mu$ L/minute (fast)
- e) Sample size: 50  $\mu$ L
- f) Doublet Discriminator gates at approximately 8000 and 16,500
- g) Collect MFI

**Note:** *The CAL2 setting for the Bio-Rad Bio-Plex analyzer should be set at the low RP1 target value.*

## ASSAY PROCEDURE

**Bring all reagents and samples to room temperature before use. It is recommended that all samples and standards be assayed in duplicate.**

**Note:** *Protect microparticles and Streptavidin-PE from light at all times.*

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Resuspend the diluted microparticle cocktail by inversion or vortexing. Add 25  $\mu\text{L}$  of the microparticle cocktail to each well of the microplate.
3. Add 100  $\mu\text{L}$  of Standard or sample\* per well. Securely cover with a foil plate sealer. Incubate for 3 hours at room temperature on a horizontal orbital microplate shaker (0.12" orbit) set at  $800 \pm 50$  rpm. A plate layout is provided to record standards and samples assayed.
4. Using a magnetic device designed to accommodate a microplate, wash by applying the magnet to the bottom of the microplate, removing the liquid, filling each well with Wash Buffer (100  $\mu\text{L}$ ) and removing the liquid again. Complete removal of liquid is essential for good performance. Perform the wash procedure three times.  
  
**Note:** *Refer to the magnetic device user manual for proper wash technique using a round bottom microplate.*
5. Add 50  $\mu\text{L}$  of diluted Biotin Antibody Cocktail to each well. Securely cover with a foil plate sealer and incubate for 1 hour at room temperature on the shaker set at  $800 \pm 50$  rpm.
6. Repeat the wash as in step 4.
7. Add 50  $\mu\text{L}$  of Streptavidin-PE to each well. Securely cover with a foil plate sealer and incubate for 30 minutes at room temperature on the shaker set at  $800 \pm 50$  rpm.
8. Repeat the wash as in step 4.
9. Resuspend the microparticles by adding 100  $\mu\text{L}$  of Wash Buffer to each well. Incubate for 2 minutes at room temperature on the shaker set at  $800 \pm 50$  rpm.
10. Read within 90 minutes using a Luminex or Bio-Rad Analyzer.

\*Serum and plasma samples require dilution. See Sample Preparation section.

## **CALCULATION OF RESULTS**

Use the Standard concentrations on the Standard Value Card and calculate 4-fold dilutions for the remaining levels. Average the duplicate readings for each standard and sample and subtract the average blank Median Fluorescence Intensity (MFI).

Create a standard curve for each analyte by reducing the data using computer software capable of generating a five parameter logistic (5-PL) curve-fit.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

## **CALIBRATION**

This assay is calibrated against highly purified recombinant human cytokines produced at R&D Systems.

## SPECIFICITY

The assay was tested for cross-reactivity and interference with the following factors. Less than 0.5% cross-reactivity and interference was observed unless otherwise noted on the analyte specific datasheet.

### Recombinant human:

4-1BB Ligand	IL-2 R $\alpha$	LIF R $\alpha$
Amphiregulin	IL-3	LIGHT
Angiogenin	IL-3 R $\alpha$	M-CSF
Angiopoietin-2	IL-4	MCP-1
APRIL	IL-4 R $\alpha$	Midkine
BAFF/BLyS	IL-5 R $\alpha$	MIF
BDNF	IL-6 R $\alpha$	MIP-1 $\alpha$
Cardiotropin-1	IL-7 R $\alpha$	MIP-1 $\beta$
CD4	IL-8	$\beta$ -NGF
CD40	IL-9	NT-3
CD40 Ligand	IL-10	NT-4
CNTF	IL-11	Oncostatin M (OSM)
EDA-A2	IL-12 p70	Osteoprotegerin
EDA	IL-12 p40	OX40 Ligand
$\beta$ -ECGF	IL-17B	PD-ECGF
EGF	IL-17C	PDGF-AA
Epo	IL-17D	PDGF-AB
Fas	IL-17E	PDGF-BB
Fas Ligand	IL-18	Pleiotrophin/PTN
FGF acidic	IL-19	RANTES
FGF basic	IL-20	SCF
FGF-4	IL-21	SLPI
FGF-5	IL-22 R $\alpha$ 1	SMDF
FGF-6	IL-23 R	Lymphotoxin $\alpha$ 1/ $\beta$ 2
G-CSF	IL-24	Lymphotoxin $\alpha$ 2/ $\beta$ 1
GDNF	IL-26	TGF- $\alpha$
GITR	IL-27	TGF- $\beta$ 1
GITR Ligand	IL-28A/IFN- $\lambda$ 2	TGF- $\beta$ 2
GRO- $\alpha$	IL-29/IFN- $\lambda$ 1	TGF- $\beta$ 3
GRO- $\beta$	IL-32 $\alpha$	TGF- $\beta$ 5
GRO- $\gamma$	IL-32 $\beta$	TGF- $\beta$ RII
HB-EGF	IL-32 $\gamma$	TNF- $\beta$
HGF	IL-33 (Pro)	TNF RI
IGF-I	IL-36 $\alpha$	TNF RII
IL-1 $\alpha$	IL-36 $\beta$	TRAIL
IL-1F7/FIL1 zeta	IL-36 $\gamma$ /IL-1F9	TRANCE
IL-1ra	IL-36Ra/IL-1F5	TWEAK
IL-1 RI	Leptin	VEGF
IL-1 RII	LIF	VEGI

## SPECIFICITY *CONTINUED*

### Recombinant mouse:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-7  
IL-10  
IL-13  
IL-15  
IL-17A  
IL-17F  
IL-22  
IL-31  
IL-33  
IL-32 $\beta$   
TNF- $\alpha$

### Recombinant rat:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-10  
IL-13  
IL-17F  
IL-22  
IL-23  
TNF- $\alpha$

### Recombinant porcine:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-10  
TNF- $\alpha$

### Recombinant cotton rat:

IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-10  
IL-13  
TNF- $\alpha$

### Recombinant feline:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-10  
TNF- $\alpha$

### Recombinant canine:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-10  
IL-13  
IL-17  
TNF- $\alpha$

### Other recombinants:

rhesus macaque IL-1 $\beta$   
rhesus macaque IL-5  
rhesus macaque IFN- $\gamma$   
rhesus macaque TNF- $\alpha$   
rabbit IL-2  
rabbit IL-6  
rabbit TNF- $\alpha$   
guinea pig IL-1 $\beta$   
guinea pig IL-10  
bovine IFN- $\gamma$   
bovine IL-2  
bovine IL-5  
bovine TNF- $\alpha$

### Recombinant human multiplex partners:

GM-CSF  
IFN- $\gamma$   
IL-1 $\beta$   
IL-2  
IL-5  
IL-6  
IL-7  
IL-13  
IL-15  
IL-17A  
IL-17F  
IL-22  
IL-23  
IL-31  
IL-33  
IL-36 $\beta$   
TNF- $\alpha$

**PLATE LAYOUT**

Use this plate layout to record standards and samples assayed.

12								
11								
10								
9								
8								
7								
6								
5								
4								
3								
2								
1								
	A	B	C	D	E	F	G	H



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