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Urine - Based HSV-1&2 PCR Detection Kit

Product Insert

Product # 31700

Herpes Simplex Virus 1 and 2 (HSV-1&2) are both members of the herpes virus family, Herpesviridae. HSV-1 and HSV-2 have relatively large double-stranded DNA genomes. HSVs are primarily transmitted by sexual intercourse, direct contact with lesions or perinatally. Most HSV positive cases are characterised by lesions on the skins and mucous membranes of the mouth and genitals. HSV infection can be either primary or a recurrence of a previous infection. More than 90% of the primary HSV infections are asymptomatic. Primary infection with HSV-1 can lead to gingivostomatitis, eczema herpeticum, keratoconjunctivitis and encephalitis. The primary symptoms of a secondary infection are skin lesions in the nose, mouth and genital regions. The infection is contagious, mainly during an epidemic.

Principle of the Test

Norgen's Urine-Based HSV-1&2 PCR Detection Kit constituents a ready-to-use system for the isolation, detection and differentiation of HSV-1 and HSV-2 using end-point PCR. The kit first allows for the isolation of total DNA, including viral DNA, from the urine samples using spin-column chromatography based on Norgen's proprietary resin. The viral DNA is isolated free from inhibitors, and can then be used as the template in a PCR reaction for HSV-1 and HSV-2 detection using the provided HSV-1&2 Master Mix. The HSV-1&2 Master Mix contain reagents and enzymes for the specific amplification of a 275bp region of HSV-1 and 350 bp region of HSV-2. In addition, Norgen's Urine-Based HSV-1&2 PCR Detection Kit contains a second heterologous amplification system to identify possible PCR inhibition and/or inadequate isolation. The amplification and detection of either the *HSV-1&2 Isolation Control (IsoC)* or the *PCR control (PCRC)* does not reduce the detection limit of the analytical HSV-1 or HSV-2 PCR. The kit is designed to allow for the testing of 24 samples.

Kit Components:

Component	Contents
Binding Solution I	20 mL
Proteinase K	0.6 mL
Pronase	0.6 mL
Binding Solution II	3 mL
Wash Solution I	4 mL
Wash Solution II	12 mL
Elution Buffer	3 mL
Mini Filter Spin Columns	24
Collection Tubes	24
Elution tubes (1.7 mL)	24
HSV-1&2 2x PCR Master Mix	0.35 mL
HSV-1&2 Isolation Control (IsoC)[a]	0.4 mL
HSV-1&2 Positive Control (PosC)[b]	0.1 mL
HSV-1&2 Negative Control (NegC)	1.25 mL
Norgen's DNA Marker	0.1 mL
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[a] *The positive control is a cloned HSV-1 and HSV-2 product*
[b] *The isolation control is a cloned PCR product*

Customer-Supplied Reagents and Equipment

- Disposable powder-free gloves
- Centrifuge with a swinging bucket rotor capable of 2000 x g
- Benchtop microcentrifuge
- Micropipettors
- Sterile pipette tips with filters
- PCR tubes
- 96 – 100% ethanol
- 60°C incubator
- 15 mL tubes

Storage Conditions and Product Stability

All buffers should be kept tightly sealed and stored at room temperature (15-25°C) for up to 1 year without showing any reduction in performance.

Norgen's Urine-Based HSV-1&2 PCR Detection Kit contains ready-to-use Proteinase K and Pronase solutions, which are dissolved in a specially prepared storage buffer. The Proteinase K and the Pronase are stable for up to 1 year after delivery when stored at room temperature. To prolong the lifetime of Proteinase K and Pronase, storage at 2–8°C is recommended.

The HSV-1&2 2x PCR Master Mix, the *HSV-1&2 Isolation Control* (*IsoC*), the *HSV-1&2 Positive Control* (*PosC*) and the *HSV-1&2 Negative Control* (*NegC*) should be kept tightly sealed and stored at -20°C for up to 1 year without showing any reduction in performance. Repeated thawing and freezing (> 2 x) should be avoided, as this may reduce the sensitivity. If the reagents are to be used only intermittently, they should be frozen in aliquots.

General Precautions

The user should exercise the following precautions while using the kit:

- Use sterile pipette tips with filters.
- Store and extract positive material (specimens, controls and amplicons) separately from all other reagents and add it to the reaction mix in a spatially separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Work quickly on ice.

Quality Control

In accordance with Norgen's ISO 9001 and ISO 13485-certified Quality Management System, each lot of Norgen's Urine-Based HSV-1&2 PCR Detection Kit, the HSV-1&2 2x PCR Master Mix, the *HSV-1&2 Isolation Control* (*IsoC*), the *HSV-1&2 Negative Control* (*NegC*) and the *HSV-1&2 Positive Control* (*PosC*) are tested against predetermined specifications to ensure consistent product quality.

Product Use Limitations

Norgen's Urine-based HSV-1&2 PCR Detection Kit is designed for research purposes only. It is not intended for human or diagnostic use.

Product Warranty and Satisfaction Guarantee

NORGEN BIOTEK CORPORATION guarantees the performance of all products in the manner described in our product manual. The customer must determine the suitability of the product for its particular use.

Safety Information

Ensure that a suitable lab coat, disposable gloves and protective goggles are worn when working with chemicals. For more information, please consult the appropriate Material Safety Data Sheets (MSDSs). These are available as convenient PDF files online at *www.norgenbiotek.com*.

CAUTION: DO NOT add bleach or acidic solutions directly to the sample-preparation waste.

The **Binding Solution I, Binding Solution II, Wash Solution I** and **Wash Solution II** contain guanidine hydrochloride, and should be handled with care. Guanidine hydrochloride forms highly reactive compounds when combined with bleach, thus care must be taken to properly dispose of any of these solutions.

If liquid containing these buffers is spilled, clean with suitable laboratory detergent and water. If the spilled liquid contains potentially infectious agents, clean the affected area first with laboratory detergent and water, and then with 1% (v/v) sodium hypochlorite.

1. Protocol

A. Specimen Collection, Storage and Transport

Precaution: All samples have to be treated as potentially infectious material.

1. Specimen Collection and Sample Storage

- Midstream urine samples should be collected, as the first flow of urine has been shown to have a higher rate of contamination (Morimoto *et al.,* 2003).
- It is highly recommended that urine samples be collected using Norgen's Urine Collection and Preservation Tubes (Cat# 18111). The urine samples can be stored for at least one year at room temperature when collected directly using Norgen's Urine Collection and Preservation Tubes.
- Alternatively, urine samples collected using any other collection and preservation systems or reagents are also compatible with this kit.

2. Sample Transport

- Sample material should be transported in a shatterproof, leak-proof transport container as a matter of principle. Thus, a potential danger of infection due to a leakage of sample can be avoided.
- The samples should be transported following the local and national instructions for the transport of pathogen material.

B. Isolation of DNA from Urine

Notes:

- Do not spin down or filter the urine sample before proceeding with the isolation, as this could negatively affect the isolation of viral DNA.
- Ensure that all solutions are at room temperature prior to use, and that no precipitates have formed. If necessary, warm the solutions and mix well until the solutions become clear again.
- Always **vortex** both the **Proteinase K** and the **Pronase** before use.
- Preheat an incubator or heating block to 60°C.
- Prepare a working concentration of **Binding Solution II** and **Wash Solution I** by adding the proper volume of 96-100% ethanol (provided by the user) indicated in Table 1 below to the supplied bottle containing the concentrated **Binding Solution II and Wash Solution I**. The label on the bottle has a box that may be checked to indicate that the ethanol has been added.
- Elevated levels of bilirubin (≥15 mg/dl) and lipids (≥800 mg/dl) and haemolytic samples do not influence the system

Table 1. Volume of Ethanol to be added to Binding Buffer II and Wash Buffer I

	Volume Provided	Ethanol (96-100%) Volume to Add	Final Volume
Binding Solution II	3 mL	7 mL	10 mL
Wash Solution I	4 mL	11 mL	15 mL

- An *HSV-1&2 Isolation Control* (*IsoC*) is supplied. This allows the user to control the DNA isolation procedure. For this assay, add the *HSV-1&2 Isolation Control* (*IsoC*) to the lysate during the isolation procedure.
    - The *HSV-1&2 Isolation Control* (*IsoC*) **must not** be added to the sample material directly
    - Do not freeze and thaw the *HSV-1&2 Isolation Control* (*IsoC*) more than 2 times.
    - The *HSV-1&2 Isolation Control* (*IsoC*) must be kept on ice at all times during the isolation procedure.

1. Obtain a 5 mL midstream urine sample. Add 700 µL of **Binding Solution I** to the urine sample and mix well by inversion. ***(Note*: *Binding Solution I contains resin and must be mixed well before every pipeting).***

2. Centrifuge for **5 minutes at 2,000 x g**. Discard the supernatant.

3. Add 20 µL of both **Proteinase K** and **Pronase** to the precipitated slurry pellet resulting from 5 mL of the urine sample. **Vortex for 10 seconds**.

4. Incubate the mixture at **60°C for 20 minutes**.

5. After the 20 minute incubation, add 260µL **Binding Solution II**,

6. Add 15 µL **HSV-1&2 *Isolation Control (IsoC)*** to the lysate, mix well by pipeting and then transfer the entire contents into a Mini Filter Spin column (provided).

7. Centrifuge for **2 minutes at 6,700 x g**, and discard the flow-through.

8. Apply 450 µL of **Wash Solution I** to the column and centrifuge for **1 minute**. Discard the flowthrough and reassemble the spin column with its collection tube.

9. Apply 450 µL of **Wash Solution II** to the column and centrifuge for **1 minute**. Discard the flow-through and reassemble the spin column with its collection tube.

10. Spin the column for 1 minute in order to thoroughly dry the resin. Discard the collection tube.

11. Transfer the spin column to a fresh 1.7 mL Elution tube. Apply 100 µL of **Elution Buffer** to the column and centrifuge for **2 minutes at 200 x g (~2,000 RPM),** followed by **1 minute at 14,000 x g (~14,000 RPM)**.

C. HSV-1&2 PCR Assay Preparation

Notes:

- It is recommended that 10 µL of the DNA elution be used as the PCR sample input volume
- Sample volume can be varied between 2 µL – 10 µL of the DNA elution. PCR grade water should be added to make up the final volume of the PCR reaction to 20 µL.
- Using a lower volume from the sample than recommended may affect the sensitivity of the HSV-1&2 Limit of Detection.

- *HSV-1&2 Negative Control* (*NegC*) and *HSV-1&2 Positive Control* (*PosC*) must be included during every run.
- The *HSV-1&2 Negative Control* (*NegC*) and *HSV-1&2 Positive Control* (*PosC*) provided are sufficient for eight PCR runs.
- Before each use, all reagents need to be thawed completely, mixed (by repeated up and down pipetting or quick vortexing), and centrifuged briefly.

1. Prepare PCR reactions as outlined in Table 2 below. For each sample to be run, pipette 10 µL of the eluted DNA and 10 µL of the Master Mix into a PCR tube. Each PCR reaction will have a final volume of 20 µL.
2. *HSV-1&2 Negative Control* (*NegC*) and *HSV-1&2 Positive Control* (*PosC*) must be included in every run. Pipette 10 µL of *HSV-1&2 Negative Control* (*NegC*) into a PCR tube and add 10 µL of Master Mix. Pipette 10 µL of *HSV-1&2 Positive Control* (*PosC*) into a PCR tube and add 10 µL of Master Mix.
3. Program the PCR machine according to the program shown in Table 3 below.
4. Run PCR.

Table 2. PCR Assay Preparation

Preparation of PCR assay	Volume Per PCR Reaction		
HSV-1&2 2X PCR Master Mix	10 µL	10 µL	10 µL
Sample (Eluted DNA)	10 µL		
HSV-1&2 Positive Control (PosC)	----------	10 µL	----------
HSV-1&2 Negative Control (NegC)	----------	----------	10 µL
Total Volume	20 µL	20 µL	20 µL

Table 3. HSV-1&2 PCR Assay Program

PCR Cycle	Step	Temperature	Duration
Cycle 1	**Step 1**	**95°C**	**3 min**
Cycle 2 (40x)	**Step 1**	**94°C**	**15 sec**
	Step 2	**60°C**	**30 sec**
	Step 3	**72°C**	**45 sec**
Cycle 3	**Step 1**	**72°C**	**5 min**
Cycle 4	**Step 1**	**4°C**	**∞**

D. HSV-1&2 PCR Assay Interpretation

- For the analysis of the PCR data, the entire 20 µL PCR reaction should be loaded on a 1X TAE, 2% Agarose DNA gel along with 10 µL of Norgen's DNA Marker (provided).
- The PCR products should be resolved on the 1X TAE, 2% Agarose gel at 150V for 30 minutes
- Figure 1 and Table 4 explain how to interpret the PCR assay results

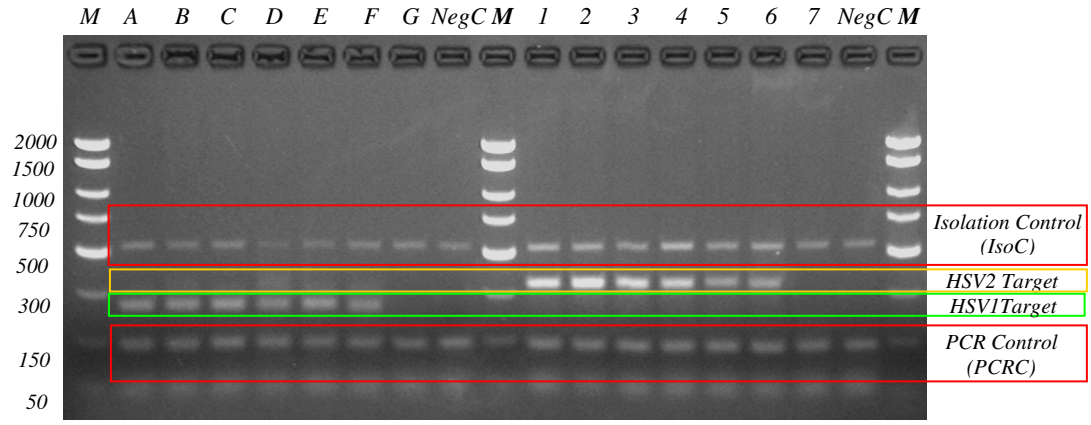

Figure 1: A representative 1X TAE, 1.7% agarose gel showing the amplification of HSV-1 and HSV-2 at different concentrations (*Target*). The size of the HSV-1 target amplicon corresponds to the 275bp band represented by the provided DNA Marker (M).The size of the HSV-2 target amplicon corresponds to the 350bp band represented by the provided DNA Marker (M). The size of the *HSV-1&2 Isolation Control* (*IsoC*) corresponds to the 500bp band represented by the provided DNA Marker (M). The HSV-1&2 2X PCR Master Mix contains a HSV-1&2 PCR Control (*PCRC*). The HSV-1&2 PCRC Controls for PCR inhibition. The size of the HSV-1&2 PCRC corresponds to the 150bp band represented by the provided DNA Marker (*M*). Lanes A-G represents samples spiked with different HSV-1 concentrations isolated from 5mL urine samples (interpreted as positive results). Lanes 1-7 represents samples spiked with different HSV-2 concentrations isolated from 5mL urine samples (interpreted as positive results).The HSV-1 and HSV-2 spiked in urine samples is a cloned PCR product.

Table 4. Interpretation of PCR Assay Results

Input Type	HSV-1&2 IsoC Band (500 bp)	HSV-1 (275 bp) or HSV-2 (350 bp) Target Band	HSV-1&2 PCRC Band (150 bp)	Interpretation
Positive Control	X	X	X	Valid
Negative Control			X	Valid
Sample	X	X	X	Positive
Sample	X		X	Negative
Sample		X	X	Positive
Sample	X	X		Positive
Sample		X		Positive

** For results obtained that are not covered in Table 4 above, please refer to the Troubleshooting Section.

E. Specificity

The specificity of Norgen's Urine-Based HSV-1&2 PCR Detection Kit is first and foremost ensured by the selection of the HSV-1 and HSV-2-specific primers, as well as the selection of stringent reaction conditions. The primers were checked for possible homologies in GenBank published sequences by sequence comparison analyses. Furthermore, the specificity of the HSV-1 and HSV-2-specific primers were tested against most of the known sexually-transmitted pathogens.

F. Linear Range

- The linear range (analytical measurement) of Norgen's Urine-Based HSV-1&2 PCR Detection Kit was determined by analyzing a dilution series of HSV-1 and HSV-2 quantitative standard ranging from 8.46 x 10^9 VP/µl to 1 x 10^{-1} IU/µl.
- Each dilution has been tested in replicates (n = 4) using Norgen's Urine-Based HSV-1&2 PCR Detection Kit on 1X TAE, 1.7% Agarose gels.
- The linear range of Norgen's Urine-Based HSV-1&2 PCR Detection Kit has been determined to cover concentrations from 0.2 VP/µl to at least 8 x 10^6 VP/µl
- Under the conditions of Norgen's Urine DNA Isolation procedure, Norgen's Urine-Based HSV-1&2 PCR detection Kit covers a linear range from 200VP/mL urine to at least 8 x 10^9 VP/mL urine.

G. Frequently Asked Questions

1. How many samples should be included per PCR run?

- Norgen's Urine-Based HSV-1&2 PCR Detection Kit is designed to test 24 samples. For every 6 samples, a Negative Control and a Positive Control must be included. It is preferable to pool and test 6 samples at a time. If not, the provided Negative Control and Positive Control are enough to run 3 samples at a time.

2. How can I interpret my results for a sample if neither the HSV-1&2 PCR control nor the HSV-1&2 Isolation Control amplifies?

- If neither the HSV-1&2 PCR control nor the *HSV-1&2 Isolation Control* (*IsoC*) amplifies, the sample must be re-tested. If the positive control showed amplification, then the problem occurred during the isolation, where as if the Positive control did not amplify the problem has occurred during the setup of the PCR assay reaction.

3. How should it be interpreted if only the HSV-1&2 PCR control showed amplification but neither the HSV-1 or HSV-2 target nor the HSV-1&2 Isolation Control (IsoC) amplified for a sample?

- This indicates a poor isolation. The isolation procedure must be repeated.

4. How should it be interpreted if only the HSV-1&2 Isolation Control (IsoC) was amplified in a sample?

- The sample tested can be considered as HSV-1 or HSV-2 negative.

5. How should it be interpreted if only the HSV-1 or HSV-2 target and the HSV-1&2 PCR control were amplified in a sample?

- The sample tested can be considered as HSV-1 or HSV-2 positive.

6. How should it be interpreted if only the HSV-1 target was amplified in a sample?

- The sample tested can be considered positive. At high HSV-1 viral load, the HSV-1 amplicon will be predominant and the HSV-1&2 PCR control as well as the HSV-1&2 Isolation control may not amplify.

7. How should it be interpreted if only the HSV-2 target was amplified in a sample?

- The sample tested can be considered positive. At high HSV-2 viral load, the HSV-2 amplicon will be predominant and the HSV-1&2 PCR control as well as the HSV-1&2 Isolation control may not amplify.

8. How should it be interpreted if only the HSV-1&2 PCR control and the HSV-1&2 Isolation Control (IsoC) showed amplification?

- The sample tested can be considered negative

9. Can I process a different urine volume?

- The reagents provided with the isolation kit are only sufficient to process 24 urine samples of 5mL each.

9. What If I added more or less of the specified reagents' volume during DNA isolation?

- Adding less volume may reduce your DNA yields. Adding more may not affect the DNA yields EXCEPT if more Elution Buffer was added. Eluting DNA in higher volumes of Elution Buffer will result in diluting your DNA.

10. What If my incubation temperature varied from the specified 60°C?

- The incubation temperature can be in the range of 55°C - 65°C. At other temperatures the activity of both the Proteinase K and the Pronase will be reduced. This will result in a reduction in your DNA yields.

11. What If my incubation varied from the 20 minutes specified in the product manual?

- Less than 20 minutes will result in lower DNA yields. More than 20 minutes may not affect your DNA yields.

12. What If I forgot to do a dry spin after my second wash?

- Your DNA elution will be contaminated with the Wash Solution. This may dilute the DNA yield in your elution and it may interfere with your down stream applications.

13. What If I forgot to add the HSV-1&2 Isolation control during the Isolation?

- The Isolation must be repeated.

Related Products	Product #
Urine DNA Isolation Kit	18100
Urine (Exfoliated Cell) DNA Purification Kit	22300
Urine (Exfoliated Cell) RNA Purification Kit	22500
Urine Bacteria DNA Purification Kit	22400
Urine Bacteria RNA Purification Kit	23400
Urine-Based HSV-1 PCR Detection Kit	32600
Urine-Based HSV-2 PCR Detection Kit	32400

Technical Assistance

NORGEN's Technical Service Department is staffed by experienced scientists with extensive practical and theoretical expertise in sample and assay technologies and the use of NORGEN products. If you have any questions or experience any difficulties regarding Norgen's Urine-based HSV-2 PCR Detection Kit or NORGEN products in general, please do not hesitate to contact us.

NORGEN customers are a valuable source of information regarding advanced or specialized uses of our products. This information is helpful to other scientists as well as to the researchers at NORGEN. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques.

For technical assistance and more information, please contact our Technical Support Team between the hours of 8:30 and 5:30 (Eastern Standard Time) at (905) 227-8848 or Toll Free at 1-866-667-4362 or call one of the NORGEN local distributors (www.norgenbiotek.com) or through email at techsupport@norgenbiotek.com.

References

Morimoto, M., Yanai, H., Chiba, H., Matsuno, K. and Shukuya, K. (2003). Importance of midstream clean-catch technique for urinalysis, reconfirmed by urinary flow cytometry. *Clin Chim Acta.* 333, 101-102.

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