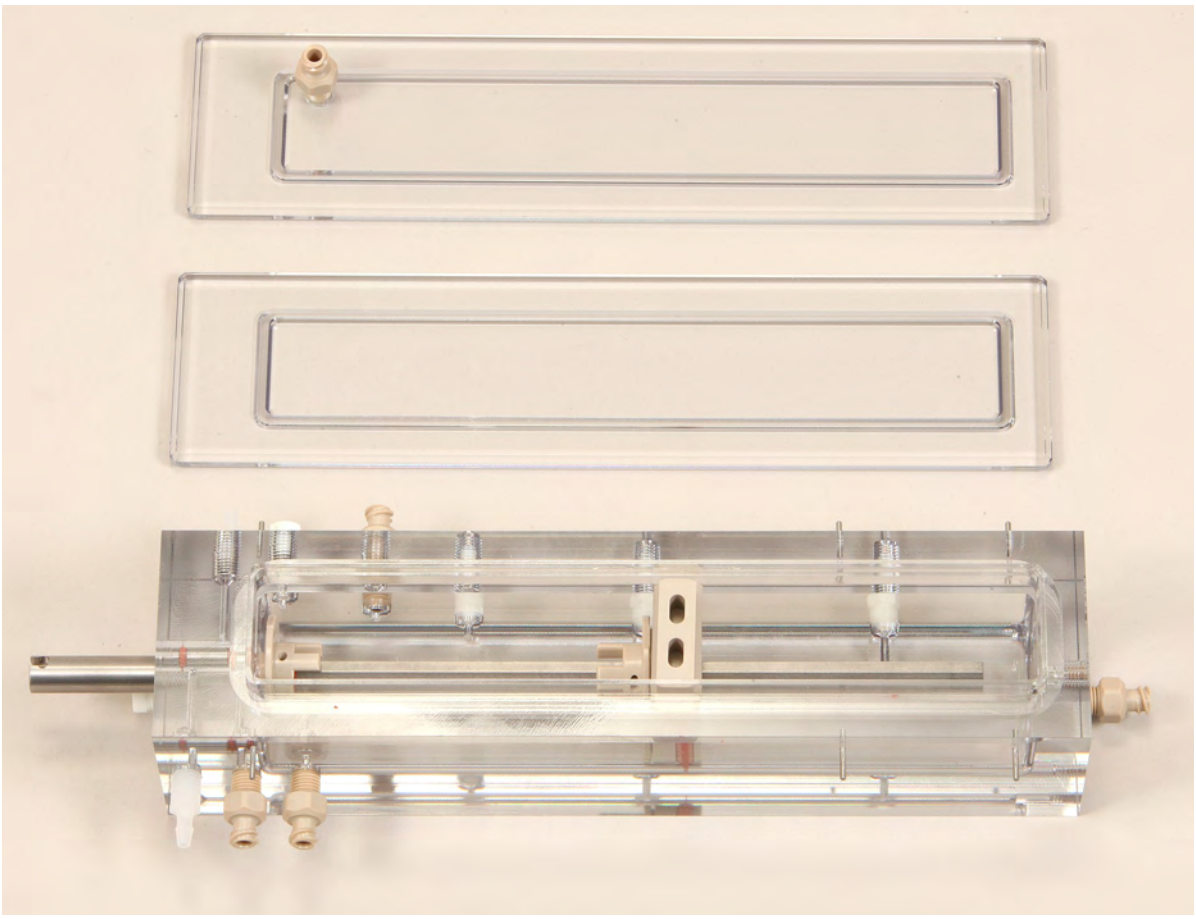
Large Animal Hollow Organ Bioreactor



HARVARD APPARATUS
REGENERATIVE TECHNOLOGY

Regenerated Organs for Transplant

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HARVARD APPARATUS
REGENERATIVE TECHNOLOGY

Harvard Apparatus Regenerative Technology (HART) partners with leading global scientists to provide specialized solutions.

The company is uniquely positioned to develop advanced instrumentation to accelerate regenerative medicine, tissue engineering and cell therapy experimentation. From the beginning, HART works closely with leading global researchers to produce products with the highest levels of performance, quality and support necessary for the new challenges of your life science research.

We look forward to working with you to develop new tools to assist you in solving these new challenges of regenerative medicine from the lab bench to the patient. There are hundreds of publications, in regenerative medicine and stem cell research, utilizing HART products. These products will serve the researcher and the physician to accelerate basic research and clinical translation.

Disclaimer:

Use of the LARGE ANIMAL HOLLOW ORGAN Bioreactor should be conducted by a trained operator. Harvard Apparatus Regenerative Technology (HART) does not warrant unauthorized use of this product. HART warrants the operation of the bioreactor for a period of one year under normal laboratory conditions.

HART reserves the right to change the instructions for use and any related products at any time without any prior notice and is not liable for any damages arising out of any change and/or alteration of the contents or product.

This product is for RESEARCH USE ONLY and NOT FOR HUMAN USE.

U.S., international, and foreign patent applications are pending.

Warning and Caution Statements

The use of a **WARNING** statement in this User Manual alerts you to a potential safety hazard. Failure to observe a warning may result in a serious injury to the user.

The use of a **CAUTION** statement in this User Manual alerts you to where special care is necessary for the safe and effective use of the product. Failure to observe a caution may result in minor injury to the user or damage to the product or other property.

Intended Use

The Hollow Organ Small Animal Bioreactor is a vessel used to support a hollow organ scaffold for the purpose of cell-seeding; it is intended for **research use only NOT FOR HUMAN USE.**

WARNING:

Use of this device in non-research settings must be conducted under local Regulatory requirements; consult your local Regulatory Authority.

The following conditions must be met prior to using the Bioreactor:

General Safety Requirements

WARNING:

The Bioreactor should only be used by qualified personnel who have been trained by the manufacturer or other authorized representative. Unauthorized use of this device is not recommended.

WARNING:

To prevent contamination, aseptic procedures must be followed and personal protective equipment must be worn at all times when handling and using the Bioreactor.

WARNING:

Wherever blood products are used, Universal Precautions must be followed.

WARNING:

DO NOT SUPPLY EXPLOSIVE GASES TO THE BIOREACTOR

Facility Requirements

Assure that the facility is able to provide a clean, safe, and suitable area for aseptic cell processing. It is recommend that all manipulations of the unit once sterile are performed in a biological safety cabinet (laminar flow hood).

WARNING:

Failure to provide a means to conduct aseptic cell processing may result in harmful contamination.

Chapter 1: Introduction

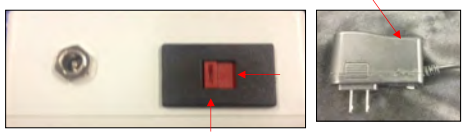
The **LARGE ANIMAL HOLLOW ORGAN Bioreactor** is a rotating, double chamber bioreactor designed for cell seeding and culturing both surfaces of a tubular matrix and includes rotational movement of the scaffold around its longitudinal axis.

A polymeric chamber houses the biological materials throughout the culture period. Cylindrical scaffold holders are constructed with working end of different diameters—to house matrices of diverse dimensions—and a central portion of smaller diameter to expose the luminal surface of the matrix for cell seeding and culturing.

A co-axial conduit links the inner chamber to the external environment through the chamber wall. This provides access to seed and feed the luminal surface of the construct. Secondary elements moving with the scaffold holder induce continuous mixing of the culture medium to increase oxygenation and mass transport.

1.1 Controller

The power module {1032418} should be connected to the power port



The controller can be used on various models of bioreactors. Confirm that the switch is in the clockwise position. Otherwise the shaft will be rotated out of the bioreactor.

Connect the Communication Cable from the controller to the bioreactor motor housing {1030280}

Alarm can be switched to the ON position which allows the controller to verify that the motor is turning. The TEST position will test the audible alarm.

The RPM switch allows selection of the revolutions per minute the shaft will be turned. The switch is turned clockwise to increase the rotational speed from 0.5 to 5 rpm. In order to select the rpm required, it is recommended that a stop watch is used to measure the turns made by the drive shaft and the knob adjusted accordingly. This should be done after the unit is in the incubator.

The motor – tray is placed in an incubator with the connection wire normally fed through an access port or by compressing the door gasket. If the door gasket is compressed to allow the wire to pass, it is recommended that a piece of foam is glued to the wire to eliminate a space being formed above the wire and the inside door of the incubator.

Place the bioreactor on the motor plate with the drive shaft connected to the motor. Note the clover leaf coupling should be in place.

1.2 Medium

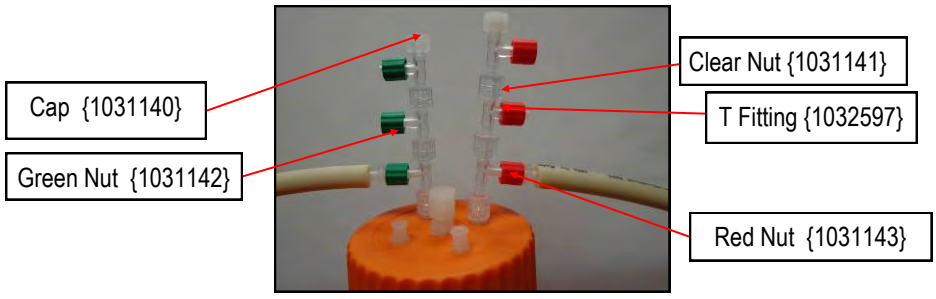
Replenishing feed medium can occur in a number of ways. When deciding on the manner of medium replenishment and removal that will be employed several factors should be taken into consideration including minimizing:

- The number of times the sterile flow path is opened to minimize the risk of contamination
- The volume of medium changed at a single time to not change the environment the cells are exposed due after they have conditioned the medium with the particular growth factors and cytokine cocktails they are accustomed to.
- The volume changed if the fresh medium temperature is different from the operating medium temperature.

One manner to deal with the considerations above is to have a medium feed bottle large enough supply the entire amount of medium required throughout the experiment. This may slow the initial growth phase of the culture as it may take a bit more time for the cells to secrete the needed levels of cytokines to condition a larger volume of medium. However, in general this is the safest manner to minimize contamination.

A second manner is to pour a volume, typically not more than 25-50% of the existing medium out and pour fresh medium in. Considerations for aseptic technique when handling large mouth reservoirs should be seriously studied.

Another manner is to have an external bag (s) or large bottle (s) of medium and have the system automatically feed and remove small volumes of medium on a continuous basis. Fittings can be used to allow multiple inlet and outlet lines to feed into a single bottle.



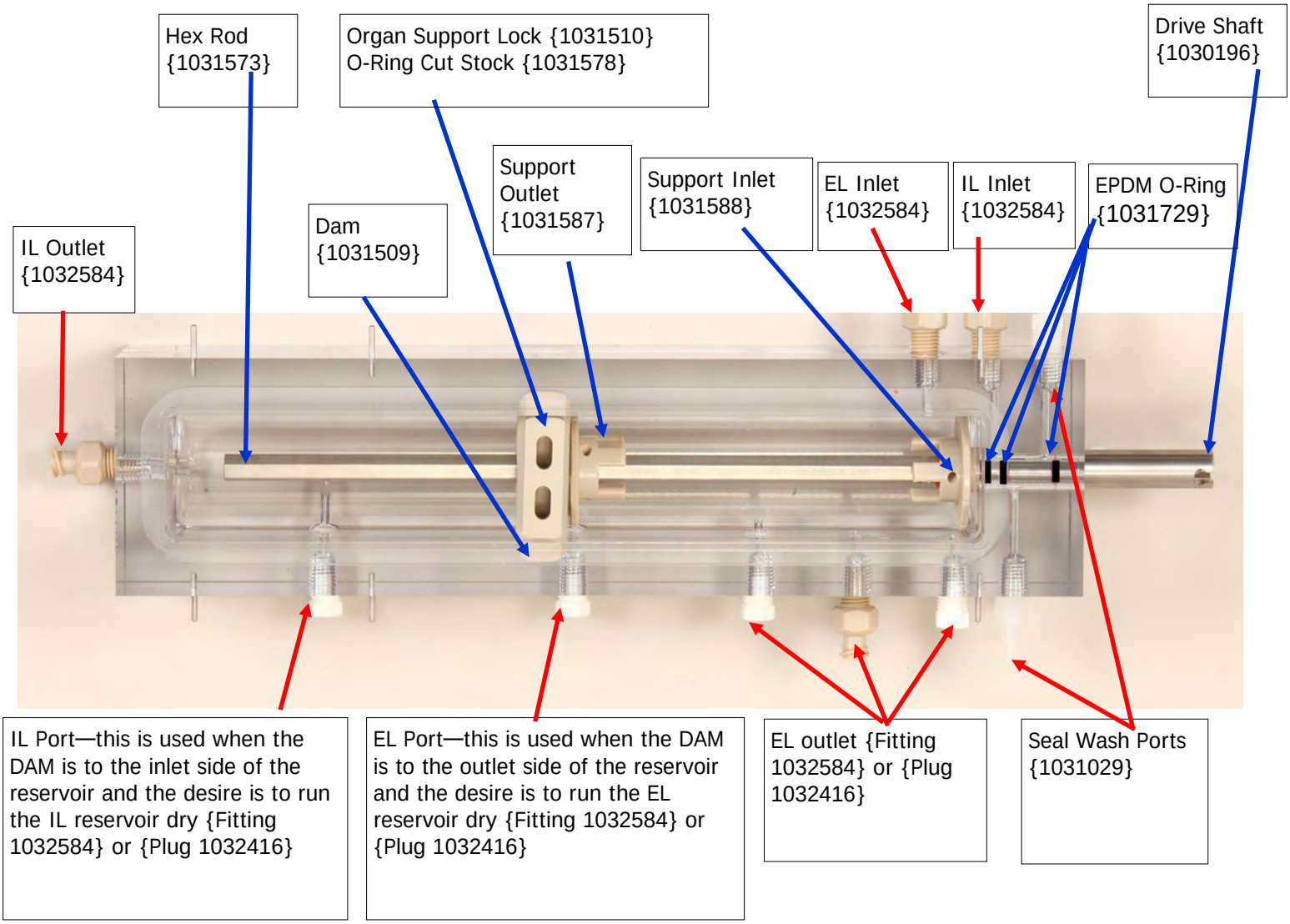
Please note that it is recommended that the tubing inside the bottle be long enough to always be below the medium level.

1.3 Equipment Components

(A) Drive Motor Base Plate: Plate that aligns the reservoir with the drive motor.

(B) Control Unit: Independent controller of rotational speed. The bioreactor may also be controlled using an ORCA Controller.

(C) Motor Drive: Drive motor responsible for rotation of the Arbor and organ.



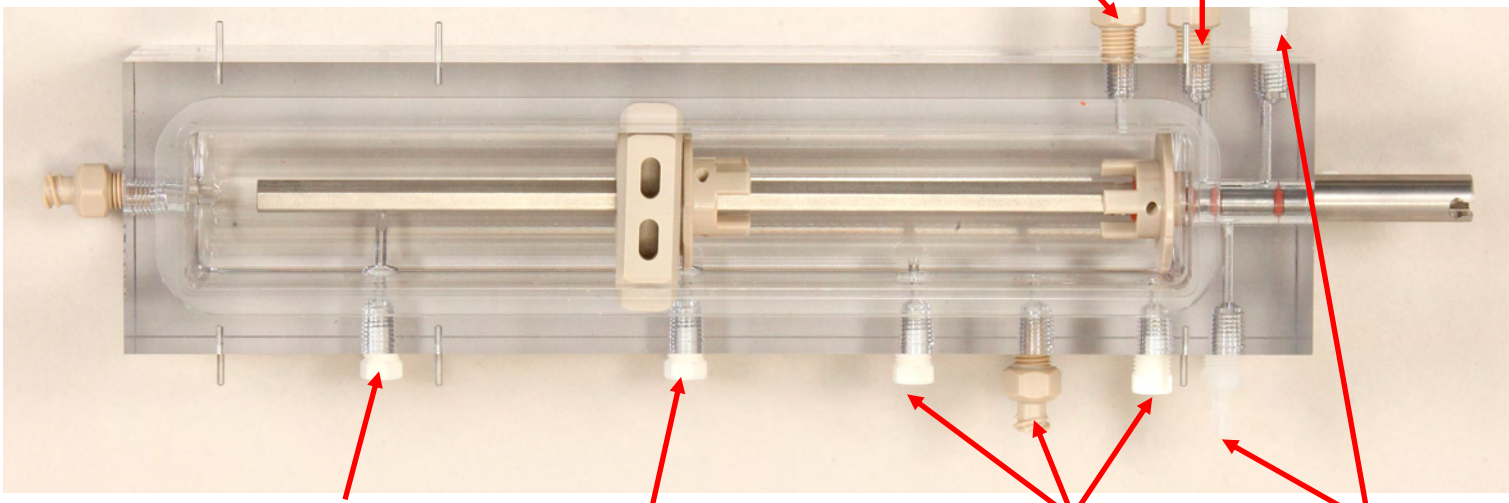
Chapter 2: Operating Instructions

2.1 Fluid Connections / Flow Path Options

It is optional to apply **RTV Sealant** {1030566} to the threads of every port and fitting. A small amount, the size of a match head, should be applied to the thread to minimize the potential for leaks. Do not use too much RTV; a single drop will spread out and cover the entire area. Do not get sealant on the top of the fitting or the lids. When changing the fittings, use a Q-tip to clear debris while holding upside down to allow residue to fall out of chamber.

Seal Wash Ports should be connected to a source of distilled water. An elevated inlet will provide enough pressure to allow flow through the ports. This will continuously wash the drive shaft and keep medium from drying and leaving salt crystals behind the seal which can cause damage to the seals and leaks.

Be careful to insert the fittings gently and not force them or you could cross-thread the fittings



IL Port—this is used when the DAM is to the inlet side of the reservoir and the desire is to run the IL reservoir dry

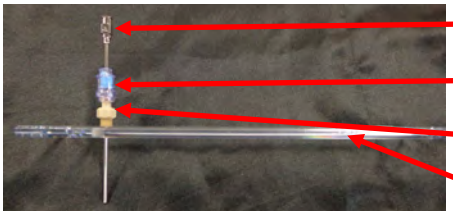
EL Port—this is used when the DAM is to the outlet side of the reservoir and the desire is to run the EL reservoir dry

EL outlet {Fitting 1032584} or {Plug 1032416}

Seal Wash Ports {1031029}

The level of the medium in the EL reservoir is selected by connecting a fitting to one of the three EL outlet ports on the reservoir or using the variable level outlet on the reservoir cover.

Place fitting into culture lid. Add swabble port to the fitting. Push cannula through the swabble port. Adjust the height to provide the level of medium in the reservoir desired.



Cannula {1032413}

Swabable Port {1031081}

Fitting {1031136}

Culture Lid {1032414}

2.2 Medium Connections

Peristaltic Pumps

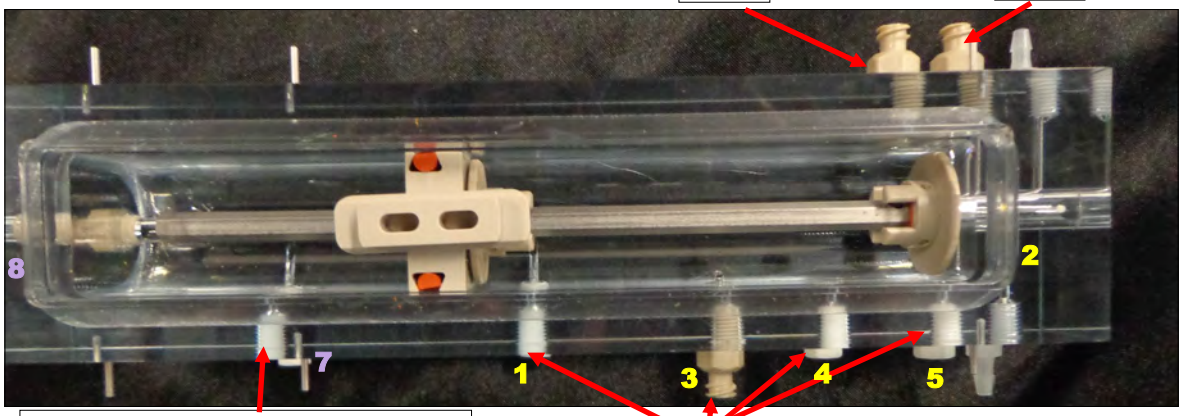
- Depending on the configuration of the system and how you set up the medium feed and waste will dictate the number of pumps and channels you will require. A typical configuration may be:
    - ◊ EL Inlet tubing is selected according to the flow rate range required {see chart}. The EL is normally fed from a bottle inside the reservoir.
    - ◊ EL Outlet tubing is selected according to the flow rate range required {see chart}. Typically the outlet flow rate is selected higher than the inlet to assure the unit does not over fill.
    - ◊ IL Inlet and outlet tubing is selected according to the flow rate range required {see chart}. The IL is normally fed from a bottle inside the reservoir.
    - ◊ IL Outlet tubing is selected according to the flow rate range required {see chart}. Typically the outlet flow rate is selected higher than the inlet to assure the unit does not over fill.

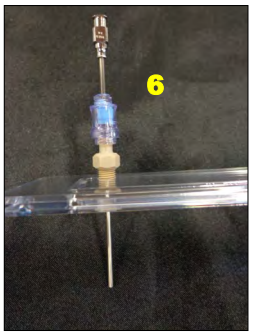
    This type of configuration would require 2 pumps with two channels each

⇒ Placing the drive unit of the peristaltic pump inside the incubator is usually recommended to minimize the tubing length and thermal effects of having liquid moving in and out of the incubator. This also minimizes the number of tubes that have to go through an aces port or that have to be squeezed against a door gasket.

Connecting the EL Circuit

- The **EL inlet** should be connected to a piece of Masterflex (see chart below) tubing through a Luer Barb Fitting to the EL reservoir. It is recommended that the user allows enough tubing length to not only complete the circuit to the EL outlet but with enough length to spare to allow the tubing to be moved an inch once per week. Moving the tubing an inch (out of the head) minimizes tubing wear on a single spot.
- **The EL outlet** should be selected according the to the medium level desired in the reservoir. The side of the chamber allows five choices. There are two choices that will allow running of the chamber virtually dry (1 & 2 below). The number 2 position can have issues with tubing fitting under the drive shaft if the tubing is too large. Both allow the exact functionality. Positions 3,4, & 5 allow the medium level to be fixed at three different levels. The variable level EL outlet on the Lid (6) may also be used. The level is selected by simply adjusting the level of the cannula tip.
- The **IL Inlet** should be connected to a piece of Masterflex {see chart} tubing through a Luer Barb Fitting to the IL reservoir. The positions #7 or #8 are identical. Position #7 may not be usable depending on the length of the scaffold which will dictate the dam location. We recommend that you allow enough tubing length to not only complete the circuit to the IL outlet but with enough length to spare to allow the tubing to be moved an inch each week. Moving the tubing an inch (out of the head) minimizes tubing wear on a single spot.





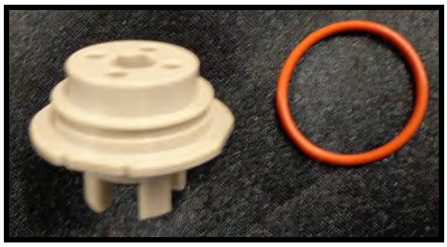
IL Port - this is used when the DAM is to the inlet side of the reservoir and the desire is to run the IL reservoir dry

Sealing Ports

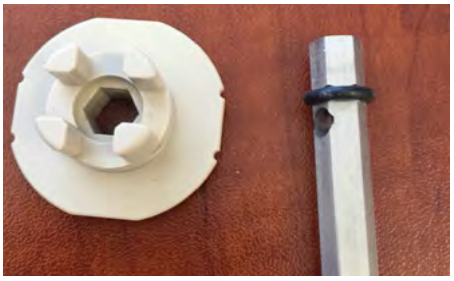
Swabble Ports {1031081} are available in bulk. These ports can be autoclaved once. They allow a sterile barrier to be maintained that can be connected to.

Sterile Caps {1032581} are available. These are shipped in individual sterile blisters. Each cap has both a male and female dead end port.

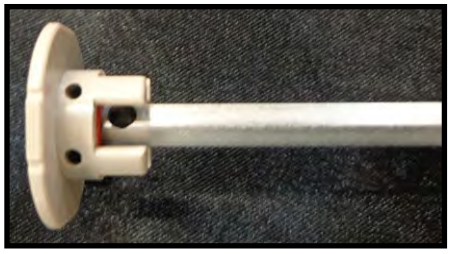
2.3 Chamber Assembly



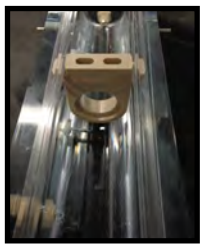
Insert the O-ring {1031577} onto the Organ Support Outlet {1031587}.



Place the small O-Ring {1030195} onto the end of the hex rod {1031573}.



Push O-Ring in flush against the flex hole of the Support Inlet {1031588}



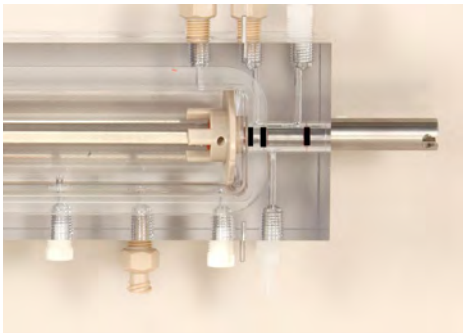
Lock, Organ Support {1031510} should be placed on the Dam {1031509}, Organ Support with the dowel Pin.

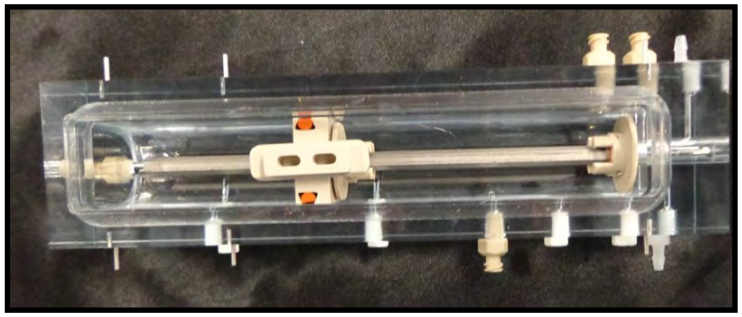
The Lock should swivel parallel to the Dam.

{1032467} O-Ring removal tool is used to replace O-Rings on the drive shaft

Place three black EPDM O-Rings {1031729} onto the driveshaft {1030196}.

Insert the hex rod {1031573} and the driveshaft.





Operating Instructions

Prior to Starting:

Assure that all the requirements identified in this manual have been successfully met.

Personnel using the LARGE ANIMAL HOLLOW ORGAN Bioreactor should read through this procedure in its entirety prior to using the device.

Personnel should train on all processes and equipment prior to operating the Bioreactor.

WARNING:

Failure to follow aseptic techniques and failure to train on all processes and procedures prior to using the bioreactor may result in critical delays, contamination, and other harmful events

2.4 Sterilization

Bioreactor Equipment for Sterilization

Sterile Pouch	Equipment
Bag #1	Culture vessel
Other bags	Organ Holder Parts
Wipe down DO NOT AUTOCLAVE	Drive Motor
	Base Plate
	Control Unit
	Power Supply

Before autoclaving, foil or autoclave wrap the following:

1. Reservoirs and connectors

2. Tubing ends

3. Luer fittings of all bioreactor connections (do not unscrew).

Then, remove valves and drive shaft from the operating ports and place them into the storage ports.

2.5 Surface Disinfection

Equipment
Drive Motor & Base Plate
Control unit
Power Supply

NOTE:

Two operators are recommended for unpacking, disinfection and assembly.

CAUTION:

In the case where several bioreactors are being used simultaneously, a unique identifier should be placed on each bioreactor component to avoid parts being exchanged. Colored stickers have typically been used on each pouch. Similar precautions should be taken with components post sterilization.

WARNING:

Failure to follow the defined sterilization procedure may damage the bioreactor and make it unsuitable for use.

2.6 Post-Sterilization Assembly

Assembly Preparation

The bioreactor parts are delivered in sterile pouches. Unpacking and bioreactor assembly should be performed in a clean room environment provided by a laminar airflow cabinet.

CAUTION:

To prevent contamination, aseptic procedures must be followed and personal protective equipment must be worn at all times when handling and using the Bioreactor.

CAUTION:

Failure to strictly follow aseptic techniques may result in critical delays, contamination, and other harmful events.

Sterile Bioreactor Equipment

Bioreactor
Arbor & organ sterilized in sterile pouch
Driveshaft
Reservoir Cover
Transport Sealing Cover

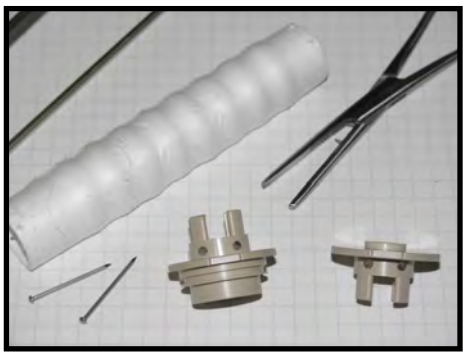
Other Materials

Surgical suture thread with needle
Sterile serological pipette
Sterile bottle (1000 ml)
PBS buffer
Culture medium
Inoculum

Disinfected Bioreactor Equipment

Drive unit
Control unit
Power Supply

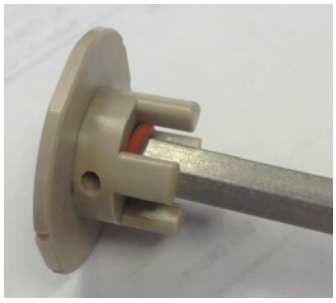
2.7 Affixing a Synthetic Scaffold / Organ to the Organ Holder



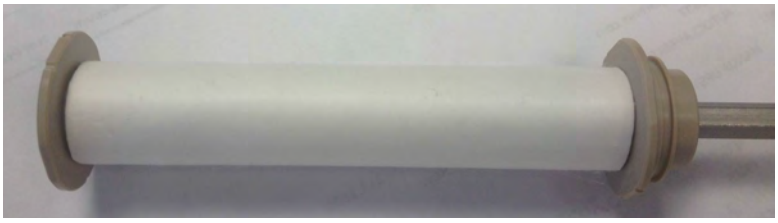
First, lay out the relevant materials you will be needing in a laminar flow hood to protect sterility. The bioreactor parts will be sterile after autoclaving.

It is important to have the correct size organ holder. If the organ holder is too small then there will not be a seal for the IL space and all the liquid will run into the EL chamber.

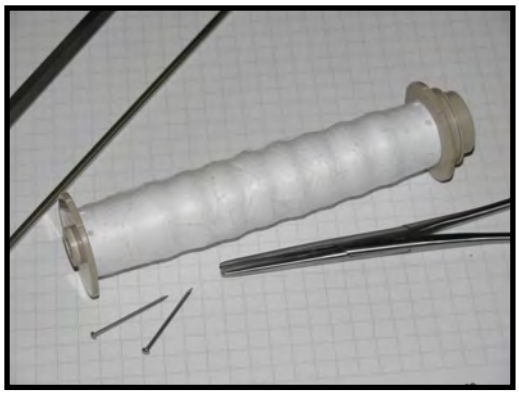
Place the O-Ring onto the shaft. A sterile pipette tip may be used to push the O-Ring into place.



Place the organ on the other organ holder. There are flat areas on each of the Organ Supports that allow the organ to lie stationary.

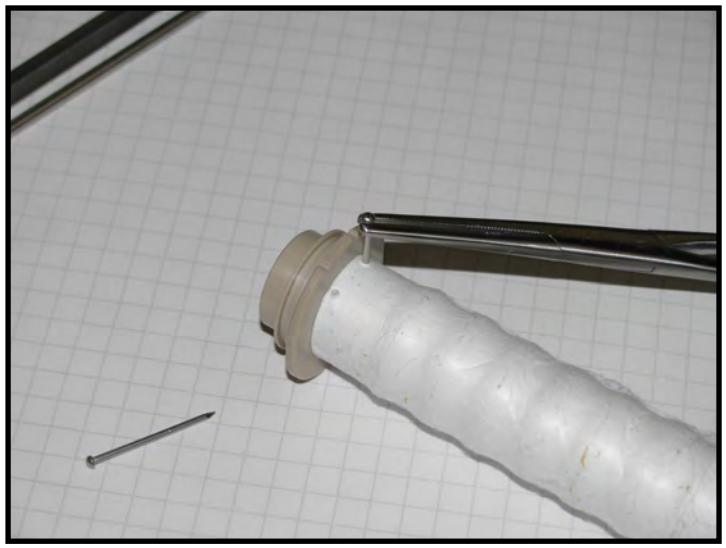


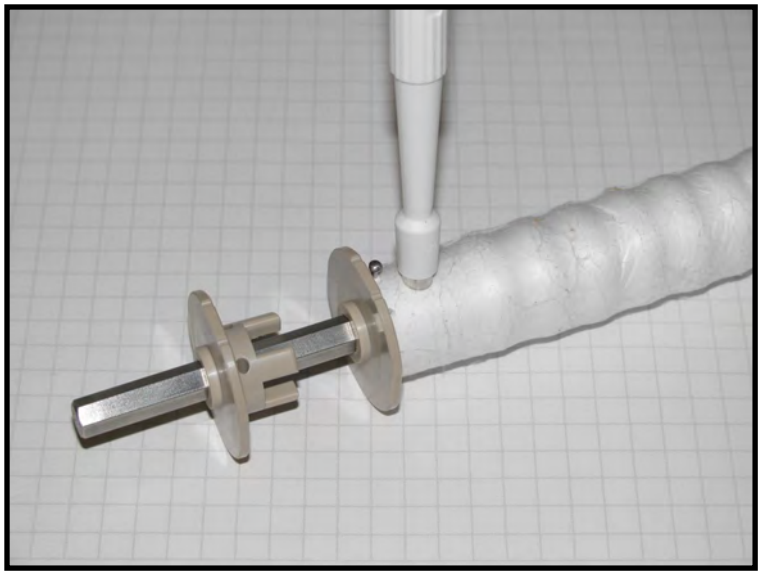
Slide the driveshaft through the organ and into the initial organ holder.



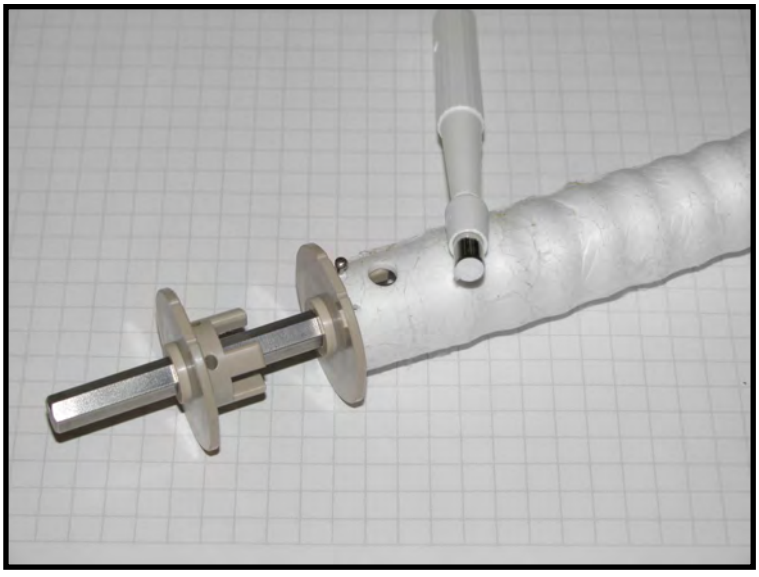
Use of pins to attach the trachea may be enough to hold a synthetic scaffold in place. Suturing may be needed to close the ends.

If pins are chosen, the notches indicate where the pins should be placed. Use the forceps to place the pins, one at a time, in the four dedicated locations. An alternative is to suture the scaffold into place by wrapping the thread around the groove in the organ holder and pulling it tight.





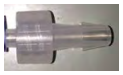
Using a biopsy punch, a sample of tissue can be taken post-incubation for further analysis if desired.

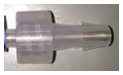


This sample can now be used for testing.

2.8 Equipment Components

Part Name	# Included	Part #
Chamber Only		1031570
Drive Motor Assembly & Controller only		1032420
Barb, 1/8" x NPT, 1/16" Seal Wash	2	1031029
Cannula, 14 Gauge	1	1032413
Chamber	1	1031570
Chamber Culture Lid	1	1032414
Chamber Sealing Lid	1	1032415
Driveshaft	1	1030196
Fitting Tapered PEEK 10 each		1031136
Fitting, Fem Luer Lock x 1/16 NPT	4	1032584
Hex Rod Organ Support, Driveshaft Side	1	1031573
Kit, Lid Closures	3	1031631
Kit, Pins, Scaffold Retention	10	1031634
O-Ring, EPDM .375 x .25 x .065 (Drive shaft) 10 each		1031729
O-Ring Dam cut stock		1031578
Organ Support Dam	1	1031509
Organ Support Lock	1	1031510
Organ Support, Outlet Side	1	1031587
Organ Support, Inlet Side	1	1031588
Pin Dowel, 3mm OD x 10mm length	1	1031567
Pin Dowel, 0.06" diameter x 0.5" length	6	1031624
Plug, Nylon 10 each	5	1032416
Power Module		1032418
RTV Sealant		1030566
Tool, Lock and Support Removal	1	1031575
Tool ORing insertion	0	1032467
Organ Holder, 16 mm ID	1	1032431
Organ Holder, 18 mm ID	1	1032427
Organ Holder, 20 mm ID	1	1032428
Organ Holder, 22 mm ID	1	1032429
Organ Holder, 24 mm ID	1	1032430

	Part Name		Qty	Part #
	Fitting Luer Female - Barb 	1/16"	25	1032297
		1/8 "	25	1032298
		3/16"	25	1032299
		1/4"	25	1032293
	Fitting Luer Male – Barb 	1/16"	25	1032294
		1/8 "	25	1032295
		3/16"	25	1032996
		1/4"	25	1032300
	Y Fitting Barb 	1/16"	10	1032075
		1/8 "	10	1032077
		3/16"	10	1031124
		1/4 "	10	1031125
	 T Fitting Barb	1/16"	10	1032076
		1/8 "	10	1032078
		3/16"	10	1032079
		1/4 "	10	1032082
	Clamp 	1/8 "	25	1031087
		¼"	25	1031088
		3/8"	25	1031089
		½"	25	1031091

	Part Name		Qty	Part #
	Fitting Luer Female - Barb 	1/16"	25	1032297
		1/8 "	25	1032298
		3/16"	25	1032299
		1/4"	25	1032293
	Fitting Luer Male – Barb 	1/16"	25	1032294
		1/8 "	25	1032295
		3/16"	25	1032996
		1/4"	25	1032300
	Y Fitting Barb 	1/16"	10	1032075
		1/8 "	10	1032077
		3/16"	10	1031124
		1/4 "	10	1031125
	 T Fitting Barb	1/16"	10	1032076
		1/8 "	10	1032078
		3/16"	10	1031079
		1/4 "	10	1032082
	Clamp 	1/8 "	25	1031087
		¼"	25	1031088
		3/8"	25	1031089
		½"	25	1031091

	Part Name		Qty	Part #
	 Swabble Port		(25 ea)	1031081
	 Sterile Sealing Caps		(25 ea)	1032581
	 Tapered Luer Fittings		(5 ea)	1032584
	 Nut Clear Cap		25	1031140
	 Green Nut		25	1031142
	 Red Nut		25	1033143
	 Clear Nut		25	1031141
	 T Fitting Luer – Slip - Slip		25	1032597
	Reservoirs 	2 L	1	1031100
		1 L	1	1031104
		500 mL	1	1031105

Chapter 3: Care & Maintenance

Cross-Infection Prevention, Biohazardous Waste, and Product Disposal

3.1 Cross-Infection Prevention / Universal Precautions

All blood products or products potentially contaminated by blood or other body/animal fluids should be treated as potentially infectious materials. Personal protective equipment should be worn at all times when using the LARGE ANIMAL HOLLOW ORGAN Bioreactor to protect personnel from becoming contaminated as well as to help prevent cross-infection and cross-contamination.

Bench tops, equipment, and other potentially contaminated surfaces should be cleaned and disinfected according to the manufacturers' and/or the facility's procedures. Any article used to clean potentially contaminated surfaces should be disposed of as Biohazardous Waste.

CAUTION:

Failure to use the manufacturers' cleaning and disinfecting procedure could result in damage to the surface or equipment.

Biohazardous Waste

Dispose of biohazardous waste according to local Regulatory requirements.

3.2 Cleaning

Flush all fluid paths with deionized water and either follow with or wipe with 70% IPA.

These materials of construction will withstand virtually all biological reagents and cold sterilization agents.

Stainless steel parts may be sonicated.

3.3 Specifications

Size of Organ	0.7—9 mm ID, up to 65 mm in length is standard
Rotation Speed	0 - 5 rpm
Diagnostics	Positional Monitoring
Power	100-240 VAC, 50/60 Hz

3.4 Troubleshooting

Arbor not turning:

Check to see if the reservoir is in place and the drive shaft is connected to the motor. The retaining clamp is tightened during installation to maintain the connection.

Check to see if the medium—blood being used has fouled the arbor. This can occur in high fibrinogen solutions.

Excess Medium / Reagent build up in the reservoir:

Check for clogging or fouling of the reservoir outlet line or port especially during a decellularization procedure.

Medium level in the IL or EL flow path has gone dry:

Confirm that there is sufficient medium in the inlet reservoirs and that the inlet tubing inside the reservoirs is below the medium level.

Leaks:

DO NOT USE BLEACH WHEN CLEANING THE BIOREACTOR AS IT HAS BEEN SHOWN TO CAUSE FAILURE OF COMPONENTS DURING AUTOCLAVE OPERATIONS.

Also, be sure to check O-Rings frequently and replace as necessary.

3.5 Frequently Asked Questions

- **What is the size of the standard organ holder?**

  The typical organ holder has an ID of the hole leading to the intraluminal space of 12 mm.

  The OD can vary on the standard unit from 16 - 24 mm. For organs requiring 11– 12 mm OD a custom organ holder can be made.

- **Should I sterilize using plasma sterilization or does autoclaving work?**

  The materials of construction are a polycarbonate reservoir, stainless steel baseplate, driveshaft, and valves, silicone rubber O-rings, Kynar Luer fittings, polycarbonate cover, and PEEK arbor components. All of these materials will withstand steam sterilization.

  One CAUTION, however…. **Please disassemble all the valve and driveshaft parts from the block before steam sterilization**. The high temperature may cause the machined holes in the reservoir to become deformed by the parts pressed into them. Reinstall the valves and driveshaft after the reservoir cools down.

  The Kynar Luer fittings have been installed using silicone adhesive to hold them in place and prevent leaks. It should not be necessary to remove these fittings from the reservoir because of the sterilization process.

- **Can washing be done with [reagent]?**

  These materials of construction will withstand virtually all biological reagents and cold sterilization agents (such as Cidex, Cidex OPA, Mucasol, etc.)

- **How does the "seal wash inlet" and outlet work?**

  This passage is designed to take a non-salt solution and flush it through a separate space between the two outside O-rings on the driveshaft. This will prevent any buildup of evaporated salt crystals from forming on the turning shaft during long periods of use. As an example, an intravenous drip bag with water set to between 0.1mL and 0.5mL per minute to continuously flush the seal wash path.

- **Can I use the bioreactor for natural and synthetic scaffolds and organs?**

  The bioreactor can be used for both types of organs.

3.5 Frequently Asked Questions (cont.)

- **Can I use Tygon tubing to save money?**

  Tygon tubing is not recommended but can be used. The issue that arises all too frequently (even with the autoclave grade Tygon tubing) is that a piece of the tubing will crease and partially block during an autoclave cycle. It is very difficult to undo the crease and can cause issues.

  If Tygon tubing is used, it is highly recommended that tubing be removed from the fittings during the autoclave process. Otherwise the tubing should be taken off the fitting and the end clipped to present fresh tubing and then pushed back on the fitting. Otherwise the fatigue experienced as the different plastics expand at differential rates may cause the tubing to come undone during an experiment.

- **Autoclave cycle settings**

  It is highly recommended that the temperature cycle not exceed 121°C as this can cause damage to various components of the bioreactor.

- **What type of pump must I use for medium delivery?**

  There is not actual requirement on the type of pump selected. In most cases people will use a peristaltic pump that allows the electronics to be outside the incubator with the drive motor inside.

  If you are using multiple pumps in the incubator or multiple bioreactors with multiple pumps, the ability of the incubator to maintain temperature may be an issue. Many types of incubators only heat and the use of multiple pumps may cause too much heat inside the incubator. If this is an issue, select an incubator that will heat and cool or contact the customer service group as the power to the pump motor may be lowered and this may aid with minimizing the heat.

- **How do I vary the length of the scaffold holder?**

  There is a sliding dam which divides the chamber into an IL and EL chamber. The slide allows a maximum length of 140 mm of an organ.

  The organ holder slides on a hexagonal drive rod. The standard organ holder has a 12 mm ID opening. There is a nipple where the organ is attached to is made with an OD diameter from 14 – 24 mm. Custom organ holders down to 11 mm OD with a smaller ID can be designed.

3.5 Frequently Asked Questions (cont.)

- **What do I do if I want to use organs with an ID of less than 10 mm?**

  A small hollow organ bioreactor chamber is available which can be used with the controller and motor drive of the large hollow organ bioreactor.

- **How to seed the Extra - Luminal cavity**

  Protocols will vary depending on many factors including the type of scaffold, medium and cell types. Typically, the cells are introduced in droplets of 100 - 250 μL. Synthetic scaffolds tend to absorb the medium and cells faster than natural scaffolds, so smaller droplets are recommended on natural scaffolds. One loading procedure is to apply droplets across the horizontal axis of the scaffold. Then rotate the arbor to the next number and repeat. Cells will fall off into the EL reservoir. They can be picked up with a pipette and the process repeated until the cells required by the protocol are attached to the scaffold.

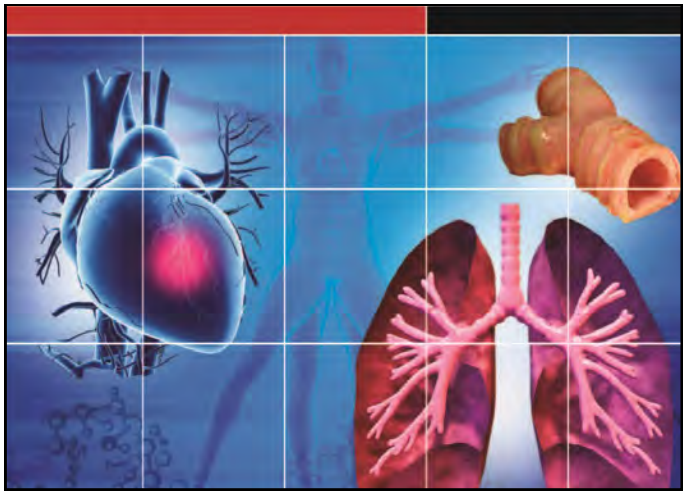
- **How to seed the Intra-Luminal cavity**

  Protocols may dictate which type of loading procedure is used. One method involves using a sterile stopcock and loading the cells directly into the IL via a syringe. Once the volume of cells required is loaded the stop cock is turned to trap the volume in the IL. Rotating at 0.5—1 rpm may assist the adherence of the cells. Some methods call for the selector valve to be set {position #1} with no outlet of the IL fluid to assist in holding the medium in the IL. Caution should be used as some seepage will occur through the scaffold walls.

  Once the cells are attached the selector can be switched back to allow normal flow from the IL reservoir through the IL. Some protocols connect two stopcocks, one on the inlet and one on the outlet and allow a piece of by pass tubing to bypass the reservoir and minimize the fluid path volume in the initial stages of the culture after the cell seeding.

Notes

Rev	Date	Changes
1.1	September 23, 2014	
1.2	December 1, 2014	O-Ring changes & O-Ring Tool
1.3	February 20, 2015	Add sealing caps
1.4	March 10, 2015	General Updates and picture with new black O-Rings
1.5	April 21, 2015	Add Spare Parts
1.6	July 15, 2015	Correction on Tee fitting P/N



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