ab175813 – 14,15 DHET ELISA Kit for Human Urine

Instructions for Use

A competitive immunoenzymatic assay for the quantitative measurement of free and glucuronidated 14,15 DHET in urine.

This product is for research use only and is not intended for diagnostic use.

Version 2 Last Updated 2 March 2015

Table of Contents

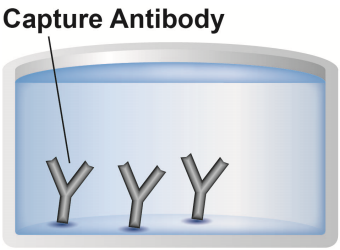
1. BACKGROUND

Abcam's 14,15 DHET competitive *in vitro* ELISA Kit for Human Urine is designed for determination of free and glucuronidated 14,15 DHET levels in urine.

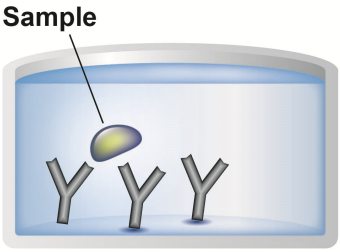
This competitive ELISA kit is based on competition between the 14,15 DHET epitope and the 14,15 DHET-HRP conjugate for a limited number of binding sites available from the anti-14,15 DHET antibody; which is coated to the wells of the 96 well ELISA plate. The conjugate concentration is held as a constant in each well, while the concentration of the 14,15 DHET is variable, based on the concentration of the sample or standard. Thus the amount of the 14,15 DHET conjugate which is able to bind to each of the wells is inversely proportional to the concentration of 14,15 DHET in the standard or sample. The amount of the conjugate which is bound to each well is then determined by the amount of color obtained, when TMB is added. The TMB reacts with the HRP available in the well. With the addition of sulfuric acid, the blue colored product is converted into a yellow colored product, which can be read on a plate reader at 450 nm.

The 14,15 DHET is a representative metabolite of soluble epoxide hydrolase-mediated metabolism of EETs, which are generated by arachidonic acid epoxygenase activity of cytochromes P450. 14,15 DHET level exhibited strong positive correlation with hypertension in rat and human and brain injury and stroke in rodents. High levels of the glucuronidated form of 14,15 DHET have been found in human urine but not in urine collected from rodents.

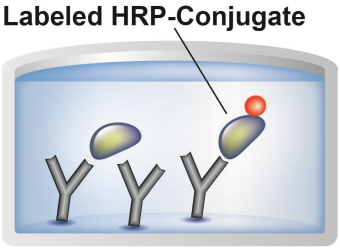
2. ASSAY SUMMARY



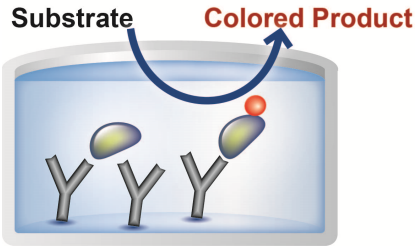
Prepare all reagents and samples as instructed.



Add standards and samples to each well used.



Add prepared HRP conjugate to each well and incubate at room temp.



Add TMB substrate to each well. Incubate at room temperature. Add Stop Solution to each well. Read immediately.

3. PRECAUTIONS

Please read these instructions carefully prior to beginning the assay.

All kit components have been formulated and quality control tested to function successfully as a kit. Modifications to the kit components or procedures may result in loss of performance.

4. STORAGE AND STABILITY

Store kit at 2-8°C or -20°C immediately upon receipt.

Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in section 9. Reagent Preparation.

5. MATERIALS SUPPLIED

Item	Amount	Storage Condition (After Preparation)
14,15 DHET ELISA plate	96 Wells	2-8°C
14,15 DHET Standard (1 mg/mL)	2 µL	2-8°C
1,000X 14,15 DHET HRP Conjugates	12 µL	2-8°C
10X Sample Dilution Buffer	25 mL	2-8°C
HRP Buffer	15 mL	2-8°C
10X Wash Buffer Solution	25 mL	2-8°C
TMB Substrate	24 mL	2-8°C
Beta-Glucuronidase enzyme	8 mg	2-8°C

6. MATERIALS REQUIRED, NOT SUPPLIED

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Microplate reader capable of measuring absorbance at 450 nm
- Incubator at 37°C
- Multi- and single-channel pipettes to deliver volumes between 10 and 1,000 µL
- Optional: Automaticplate washer for rinsing wells.
- Storage bottles
- Rotating mixer
- Deionised or (freshly) distilled water.
- Disposable tubes
- Timer
- 2N Sulfuric acid
- 1M Citric acid

7. LIMITATIONS

- ELISA kit intended for research use only. Not for use in diagnostic procedures
- Use only clean pipette tips, dispensers, and lab ware
- Do not interchange screw caps of reagent vials to avoid cross-contamination
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination
- After first opening and subsequent storage check conjugate and control vials for microbial contamination prior to further use
- To avoid cross-contamination and falsely elevated results pipette patient samples and dispense conjugate, without splashing, accurately to the bottom of wells

8. TECHNICAL HINTS

- Avoid foaming or bubbles when mixing or reconstituting components
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions
- Ensure plates are properly sealed or covered during incubation steps
- Complete removal of all solutions and buffers during wash steps is necessary for accurate measurement readings
- Addition of the TMB Substrate solution initiates a kinetic reaction, which is terminated by the addition of the Stop Solution. Therefore, the TMB Substrate and the Stop Solution should be added in the same sequence to eliminate any time deviation during the reaction
- It is important that the time of reaction in each well is held constant for reproducible results. Pipetting of samples should not extend beyond ten minutes to avoid assay drift. If more than 10 minutes are needed, follow the same order of dispensation. If more than one plate is used, it is recommended to repeat the dose response curve in each plate
- The incomplete or inaccurate liquid removal from the wells could influence the assay precision and/or increase the background
- **This kit is sold based on number of tests. A 'test' simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions**

9. REAGENT PREPARATION

Equilibrate all reagents, samples and controls to room temperature (18-25°C) prior to use.

9.1 1X Wash Buffer

Mix the 10X Wash Buffer Solution with a stir bar, applying low, gentle heat until a clear colorless solution is obtained. Dilute the entire contents of the 10X Wash Buffer Solution (25 mL) with 225 mL of deionized water to yield a final volume of 250 mL of 1 X Wash Buffer. This can then be refrigerated for the entire life of the kit.

9.2 1X HRP Conjugate

Dilute 1 vial of the 14,15 DHET-HRP conjugate (12 µL) with 12mL of HRP Buffer. One vial makes enough conjugate for one plate. The conjugate must be used the same day and should not be stored for later use.

9.3 1X Sample Dilution Buffer

Prepare 1X Sample Dilution Buffer by adding 25 mL of 10X Sample Dilution Buffer to 225 mL of dH_20. Mix gently and thoroughly.

10. STANDARD PREPARATION

Prepare serially diluted standards immediately prior to use. Always prepare a fresh set of positive controls for every use.

10.1 Label 5 microtubes as Standard # 2 - 6.

10.2 Add 900µL of the 1X Sample Dilution Buffer to the microtubes for Standards # 2 to 6.

10.3 Prepare a 1 µg/mL **Standard #1** by first spinning down the enclosed 14,15 DHET standard vial (2 µL, filled with inert gas) and then adding 1.998 mL of 1X Sample Dilution Buffer to obtain 2 mL of solution.

10.4 Prepare **Standard #2** by adding 100µL of the Standard #1 to the microtube labeled **Standard #2**. Mix thoroughly and gently.

10.5 Prepare **Standard #3** by adding 100µL of the Standard #2 to the microtube labeled **Standard #3**. Mix thoroughly and gently.

10.6 Using the table below as a guide, repeat for tubes #4 through #6.

10.7 Standard B_o contains no protein and is blank control.

Standard #	Sample to Dilute	Volume to Dilute (µL)	Volume of Diluent (µL)	Starting Conc. (pg/mL)	Final Conc. (pg/mL)
1	Step 10.3				1,000,000
2	Standard #1	100	900	1,000,000	100,000
3	Standard #2	100	900	100,000	10,000
4	Standard #3	100	900	10,000	1,000
5	Standard #4	100	900	1,000	100
6	Standard #5	100	900	100	10
B_o	None		900	-	-

11. SAMPLE COLLECTION AND STORAGE

There are different protocols for isolating and purifying 14,15 DHET depending on the medium in which it is in. For optimal results follow the appropriate protocol based on the biological sample present.

Dissolve 8 mg of Beta-Glucuronidase in 8 mL of 1 M Citric acid and adjust to pH 5.5 (400 U/mL).

11.1 **Measurement of free and glucuronidated 14,15 DHET in sample extraction with ethyl acetate.**

11.1.1 **Measurement of free 14,15 DHET:** Extract 4 mL of urine with ethyl acetate using the **Extraction Protocol** described in 11.1.4 below.

11.1.2 **Measurement of free and 14,15 DHET glucuronide:** This method is for determining the level of glucuronidated 14,15 DHET in urine after digestion of the molecule with glucuronidase. Collect the first sample as soon as the Beta-Glucuronidase is added to a reaction mixture (0 hour digestion) and then a second sample at a time when digestion of the glucuronic acid moiety of the molecule is completed. Subtract the level of the molecule in the first sample at 0 hour from the levels in the second sample after complete digestion (usually 3 hrs) to obtain the level of glucuronidated molecule.

11.1.3 **Beta-Glucuronidase digestion:**

11.1.3.1 To 4 mL of urine add 1 mL of the Beta-Glucuronidase solution, pH 5.5, to each tube (pH < 6.0).

11.1.3.2 Immediately transfer 2 mL of urine to a clean tube and flash-freeze. This is the zero time point.

11.1.3.3 Incubate the remaining 2 mL at 37°C for 3 hours. This is the 3 hour time point.

11.1.3.4 Extract.

11.1.4 **Extraction protocol:**

11.1.5 Combine an equal amount of urine sample from steps 11.1.3.2 (Zero time point) and 11.1.3.3 (3 hour time point) above (adjusted with approximately 20 µL of acetic acid to pH 4) and ethyl acetate. Vortex thoroughly. Centrifuge at 2000 rpm for ten minutes at 22°C. Three phases should result:

11.1.6 Upper organic phase – ethyl acetate phase (lipoproteins).

11.1.7 Interphase – proteins.

11.1.8 Lower phase – aqueous phase.

11.1.9 Collect the upper organic phase (a) and set aside.

11.1.10 Discard the interphase. Transfer the lower phase with a glass pipette to a new tube and repeat the ethyl acetate extraction step two more times.

11.1.10.1 Evaporation of pooled organic phase: There should be approximately 5-6 mL of the ethyl acetate phase (a). Dry the pooled organic phase in a Speedvac to get the extracted sediment (b).

11.1.10.2 Saponification (to cleave fatty acid from glycerol backbone): Dissolve the dried residues (b) in 2 mL of 20% KOH solution (for preparation see 14,15 DHET measurement in cells). Vortex

thoroughly and incubate for 1 h at 50°C. This will yield an aqueous solution (c).Vortex and incubate for 1 h at 50°C.This will yield an aqueous solution (c).

11.1.10.3 Dilute 2 mL of the aqueous solution (c) with 3 mL of H_2O. Adjust the pH using 20% formic acid (132 µL) to pH~5.5. Add ethyl acetate (1 part aqueous solution (c) + 1 part ethyl acetate), vortex thoroughly, and centrifuge at 2000 rpm for ten minutes at 22°C. Repeat the procedure twice more using an equal volume of ethyl acetate per sample. Collect the upper phase containing saponified lipids.

11.1.10.4 Dry the pooled ethyl acetate upper phase (d) and dry in a Speedvac, yielding the dried sample-sediment (e). Store the sediment (e) at -20°C. For ELISA assay, dissolve the sediment (e) in 20 µL of DMF or ethanol, then add 130 µL of 1X Sample Dilution Buffer.

11.1.10.5 For the competitive 14,15 DHET ELISA, the above 150 µL sample needs to be further diluted: Dilute 1:4 (e.g., 80 µL sample + 320 µL 1x Sample Dilution Buffer). Check the final pH (should be pH 7.4). When calculating the final concentration, consider all dilution factors.

11.1.10.6 Perform the ELISA for 14,15 DHET.

11.2 Measurement of Free and Glucuronidated 14,15 DHET without extraction

11.2.1 **Measurement of free 14,15 DHET:** Dilute 1 mL urine 4-fold with 1X Sample Dilution Buffer and apply to ELISA plate (100 µL/well). A 4X dilution is recommended although other dilution factors may be tried, too.

11.2.2 **Measurement of glucuronidated 14,15 DHET**: This method is for determining the level of glucuronidated 14,15 DHET in urine after digestion of the molecule with glucuronidase. Collect the first sample as soon as the Beta-Glucuronidase is added to a reaction mixture (0 hour digestion) and then a second sample at a time when digestion of the glucuronic acid moiety of the molecule is completed. Subtract the level of the molecule in the first sample at 0 hour from the levels in the second sample after complete digestion (usually 3 hrs) to obtain the level of glucuronidated molecule.

11.2.3 **Beta-Glucuronidase digestion:**

11.2.3.1 Dilute 1 mL of urine 4-fold with 1X Sample Dilution Buffer.

11.2.3.2 To 4 mL of urine add 1 mL of the Beta-Glucuronidase solution, pH 5.5, to each tube (pH < 6.0).

11.2.3.3 Immediately transfer 2 mL of urine to a clean tube and flash-freeze. This is the Zero time point.

11.2.3.4 Incubate the remaining 2 mL at 37ºC for 3 hours. This is the 3 hour time point.

11.2.3.5 Follow the instructions for the ELISA kit.

11.2.3.6 To calculate the amount of glucuronidated 14,15 DHET, subtract the Zero time value from the 3 hour value.

12. PLATE PREPARATION

- The 96 well plate included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents
- Unused well strips should be returned to the plate packet and stored at 4°C
- For each assay performed, a minimum of 2 wells must be used as a blank, omitting sample and conjugate from well addition. Another 2 wells must be used for a maximum binding control
- For statistical reasons, we recommend each standard and sample should be assayed with a minimum of two replicates (duplicates)

13. ASSAY PROCEDURE

- **Equilibrate all materials and prepared reagents to room temperature prior to use**
- **Please read the test protocol carefully before performing the assay. Result reliability depends on strict adherence to the test protocol as described**
- **If performing the test on anautomatic ELISA system we recommend increasing the washing steps from three to five and the volume of 1X Wash Bufferfrom 300 µL to 350 µL to avoid washing effects**
- **Assay all standards, controls and samples in duplicate**

13.1 Add 200 µL of 1X Sample Dilution Buffer into the blank wells and 100 µL of1XSample Dilution Buffer into maximum binding control wells.

13.2 Add 100 µL of each of the standards or samples into the appropriate wells.

13.3 Add 100 µL of the 1X-HRP conjugate in the all wells except the blank control wells.

13.4 Incubate the plate at room temperature for two hours.

13.5 Wash the plate three times with 400 µLof 1X Wash Buffer per well.

13.6 After the last of the three wash cycles pat the inverted plate dry onto some paper towels.

13.7 Add 200 µL of the TMB substrate to all of the wells.

13.8 Incubate the plate at room temperature for 15-30 minutes.

13.9 Add 50 µL of 2 N sulfuric acid to all of the wells.

13.10 Read the plate at 450 nm.

14. CALCULATIONS

If data redaction software is not available on your plate reader then the results can be obtained manually as follows:

14.1 Average the absorbance (Abs) readings from the blank wellsand subtract that value from each well of the plate to obtain the corrected readings. (Note: Some plate readers do this automatically. Consult the user manual of your plate reader.)

14.2 Average the corrected absorbance readings from the maximum binding control wells. This is your maximum binding.

14.3 Calculate the % Abs for Standard 1 by averaging the corrected absorbance of the two wells; divide the average by the Maximum Binding Control well average absorbance, then multiply by 100. Repeat this formula for the remaining standards.

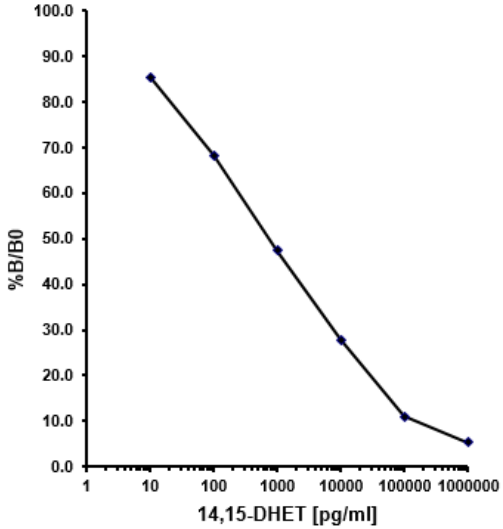
14.4 Plot the % Abs versus the concentration of 14,15 DHET from the standards using semi-log paper.

14.5 Calculate the % Abs for the samples and determine the concentrations, utilizing the standard curve.

14.6 Multiply the concentrations obtained for each of the samples by their corresponding dilution factor.

15. TYPICAL DATA

TYPICAL STANDARD CURVE – Data provided for demonstration purposes only. A new standard curve must be generated for each assay performed. The data shown here is an example of typical results obtained using the Abcam's14,15 DHET ELISA kit. These results are only a guideline, and should not be used to determine values from your samples.



Conc. (pg/mL)	% (B/B_0)
10	85.4
100	68.2
1,000	47.3
10,000	27.9
100,000	11.0
1,000,000	5.4

16. ASSAY SPECIFICITY

The specificity of the 14,15 DHET ELISA was investigated using authentic 14,15 DHET and a panel of eiconsanoids.

Eiconsanoid	Reactivity
14,15 DHET	100.00%
8,9-DHET	3.30%
11,12-DHET	3.30%
14,15-EET	1.5 %
15(s) HETE	1.00%
8,9-EET	0.40%
5(s)15(s)DiHETE	0.20%
11,12-EET	0.05%
Arachidonic Acid	0.05%
5,6-DHET	0.02%
5,6-EET	0.02%
Thromboxane B_2	0.02%
PGE_2	<0.01 %
PGF_{2a}	<0.01 %
6-keto-PGF_{1a}	<0.01 %

SENSITIVITY-

The calculated minimal detectable (MDD) dose is 3 pg/mL.The MDD was determined by calculating the mean of zero standard replicates.

17. TROUBLESHOOTING

Problem	Cause	Solution
No color present in standard wells	The HRP conjugate was not added	Redo the assay and add the conjugate at the proper step
	The HRP conjugate was not incubated for the proper time	Redo the assay and incubate for the proper time
No color in any wells	The TMB substrate was not added	Add substrate
	The TMB conjugate was not incubated for the proper time	Continue incubation until desired color is reached

Problem	Cause	Solution
The color is faint	One or all of the incubation times were cut short	Redo the assay with the proper incubation times
	The TMB substrate was not warmed up to room temperature	Redo the assay making sure all reagents are at room temperature
	The lab is too cold	Be sure the lab temperature is between 21-27°C and redo the assay
The background color is very high	The TMB substrate has been contaminated	Redo the assay with a fresh bottle of substrate
Scattered OD obtained from sample	Incorrect loading of samples	Redo assay using an 8 channel pipetman making sure the 8 channels are equal volume while loading

18. NOTES

UK, EU and ROW
Email: technical@abcam.com | Tel: +44-(0)1223-696000

Austria
Email: wissenschaftlicherdienst@abcam.com | Tel: 019-288-259

France
Email: supportscientifique@abcam.com | Tel: 01-46-94-62-96

Germany
Email: wissenschaftlicherdienst@abcam.com | Tel: 030-896-779-154

Spain
Email: soportecientifico@abcam.com | Tel: 911-146-554

Switzerland
Email: technical@abcam.com
Tel (Deutsch): 0435-016-424 | Tel (Français): 0615-000-530

US and Latin America
Email: us.technical@abcam.com | Tel: 888-77-ABCAM (22226)

Canada
Email: ca.technical@abcam.com | Tel: 877-749-8807

China and Asia Pacific
Email: hk.technical@abcam.com | Tel: 108008523689 (中國聯通)

Japan
Email: technical@abcam.co.jp | Tel: +81-(0)3-6231-0940

www.abcam.com | www.abcam.cn | www.abcam.co.jp