



E.Z.N.A.® Cycle Pure Kit

D6492-01	50 preps	V-spin
D6492-02	200 preps	V-spin
D6493-00	5 preps	Q-spin
D6493-01	50 preps	Q-spin
D6493-02	200 preps	Q-spir

July 2014

For research use only. Not intended for diagnostic testing.

E.Z.N.A.® Cycle Pure Kit

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Introduction

The E.Z.N.A.® family of products is an innovative system that radically simplifies the extraction and purification of nucleic acids from a variety of sources. The key to this system is the HiBind® matrix that specifically, but reversibly, binds DNA or RNA under optimized conditions allowing proteins and other contaminants to be removed. Nucleic acids are easily eluted with deionized water or a low salt buffer.

The E.Z.N.A.® Cycle Pure Kit is a convenient system for the fast and reliable purification of PCR products. The E.Z.N.A.® Cycle Pure Kit uses HiBind® technology to recover DNA bands from 100 bp to 10 kb free of oligonucleotides, nucleotides, and polymerase with yields exceeding 80%. The binding conditions of the HiBind® DNA Mini Columns are adjusted by the addition of a specially formulated buffer before adding the sample. Following a rapid wash step, DNA is eluted with deionized water or a low salt buffer. Purified DNA can be directly used for most downstream applications include T-A ligations, PCR sequencing, restriction enzyme digestion, or various labeling reactions.

Benefits of the E.Z.N.A.® Cycle Pure Kit

- Fast DNA recovery from enzymatic reactions in less than 10 minutes
- Reliability Optimized buffers that guarantee pure DNA
- Safety No organic extractions
- Quality Purified DNA is suitable for any application

Q-spin column vs. V-spin column

The E.Z.N.A.® Cycle-Pure Kit is available with two different types of columns. V-Spin columns have an attached cap, while Q-spin columns are capless. The columns are otherwise identical in use and application. Either column can be used with either the vacuum or centrifugation protocols. D6492 is the V-Spin version of the Cycle Pure Kit, while D6493 is the Q-Spin version.

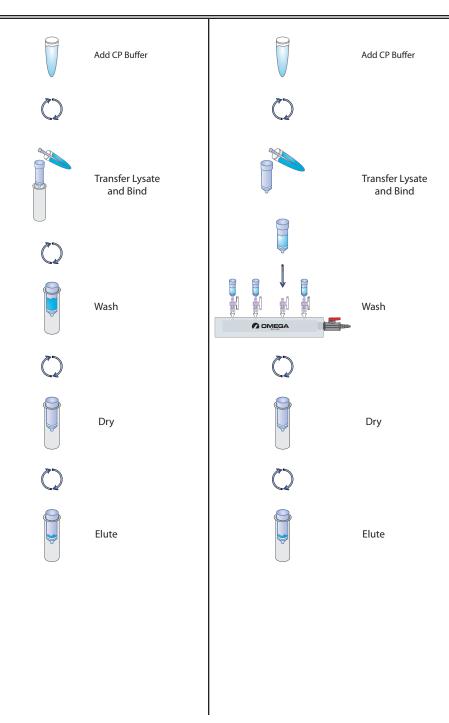
Binding Capacity

Each HiBind® DNA Mini Column can bind ~30 μg DNA.

New in this Edition: Protocol has been edited for fragments less than 200 BP.

Centrifugation Protocol

Vacuum Protocol



Kit Contents

Product Number	D6492-00 D6493-00	D6492-01 D6493-01	D6492-02 D6493-02
Preps	5	50	200
HiBind® DNA Mini Columns	5	50	200
2 mL Collection Tubes	5	50	200
CP Buffer	5 mL	40 mL	150 mL
Elution Buffer	5 mL	10 mL	20 mL
DNA Wash Buffer	1.5 mL	15 mL	3 x 25 mL
User Manual	√	√	√

Storage and Stability

All of the E.Z.N.A.® Cycle Pure Kit components are guaranteed for at least 12 months from the date of purchase when stored at room temperature. During shipment or storage in cool ambient conditions, precipitates may form in CP Buffer. Dissolve such deposits by warming the solution at 37°C and gently shaking.

Preparing Reagents

Dilute DNA Wash Buffer with 100% ethanol as follows and store at room temperature.

Kit	100% Ethanol to be Added
D6492-00 D6493-00	6 mL
D6492-01 D6493-01	60 mL
D6492-02 D6493-02	100 mL per bottle

Guidelines for Vacuum Manifold

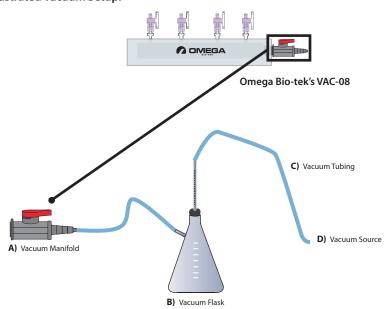
The following is required for use with the Vacuum Protocol:

- A) Vacuum Manifold (We recommend Omega Bio-tek's VAC-08)
 Other Compatible Vacuum Manifolds: Qiagen QIAvac24, Sigma AldrichVM20,
 Promega Vacman®, or manifold with standard luer connector
- B) Vacuum Flask
- C) Vacuum Tubing
- **D)** Vacuum Source (review tables below for pressure settings)

Manifold	Recommended Pressure (mbar)		
VAC-08	-200 to -600		

Conversion from millibars:	Multiply by:
Millimeters of mercury (mm Hg)	0.75
Kilopascals (kPa)	0.1
Inches of mercury (inch Hg)	0.0295
Torrs (Torr)	0.75
Atmospheres (atmos)	0.000987
Pounds per Square Inch (psi)	0.0145

Illustrated Vacuum Setup:



E.Z.N.A.® Cycle Pure Kit Centrifugation Protocol

E.Z.N.A. Cycle Pure Kit - Centrifugation Protocol

Materials and Equipment to be Supplied by User:

- Microcentrifuge capable of at least 13,000 x q
- Nuclease-free 1.5 mL microcentrifuge tubes
- 100% ethanol
- Optional: Sterile deionized water or TE Buffer
- For Fragments < 200 BP, 100% Isopropanol

Before Starting:

- Prepare DNA Wash Buffer according to the "Preparing Reagents" section on Page 4.
- Perform agarose gel/ethidium bromide electrophoresis to analyze PCR product.
- 2. Determine the volume of your PCR reaction.
- 3. Transfer the sample into a clean 1.5 mL microcentrifuge tube.
- Add 4-5 volumes CP Buffer. For PCR products smaller than 200 bp, add 5 volumes CP Buffer and 0.4 V 100 % Isopropanol

Note: Volume refers to the size of your PCR reaction. For example, if your PCR reaction is 100 μ L and is smaller than 200 bp, you would use 500 μ L CP Buffer abd 40 μ L Isopropanol.

- 5. Vortex to mix thoroughly. Briefly centrifuge to collect any drops from the inside of the lid.
- 6. Insert a HiBind® DNA Mini Column into a 2 mL Collection Tube (provided).
- 7. Add the sample from Step 5 to the HiBind® DNA Mini Column.
- 8. Centrifuge at maximum speed (\geq 13,000 x g) for 1 minute at room temperature.

E.Z.N.A.® Cycle Pure Kit Centrifugation Protocol

Note: DNA Wash Buffer must be diluted with ethanol before use. Please see the

Discard the filtrate and reuse collection tube.

11. Centrifuge at maximum speed for 1 minute.

Preparing Reagents section on Page 4 for instructions.

10. Add 700 μL DNA Wash Buffer.

9.

12. Discard the filtrate and reuse collection tube. 13. Repeat Steps 10-12 for a second DNA Wash Buffer wash step. 14. Centrifuge the empty HiBind® DNA Mini Column at maximum speed for 2 minutes to dry the column. **Note:** This step is critical for removal of trace ethanol that may interfere with downstream applications. 15. Transfer the HiBind® DNA Mini Column into a clean 1.5 mL microcentrifuge tube (not provided). 16. Add 30-50 µL Elution Buffer, TE Buffer, or sterile deionized water directly to the center of column matrix. 17. Let sit at room temperature for 2 minutes. 18. Centrifuge at maximum speed for 1 minute. Note: This represents approximately 80-90% of bound DNA. An optional second elution will yield any residual DNA, though at a lower concentration. 19. Store DNA at -20°C.

E.Z.N.A.® Cycle Pure Kit Vacuum Protocol

E.Z.N.A.® Cycle Pure Kit - Vacuum Protocol

Materials and Equipment to be Supplied by User:

- Vacuum Manifold
- Microcentrifuge capable of at least 13,000 x q
- Nuclease-free 1.5 mL microcentrifuge tubes
- 100% ethanol
- Optional: Sterile deionized water or TE Buffer
- For Fragments < 200 BP, 100% Isopropanol

Before Starting:

- Prepare DNA Wash Buffer according to the "Preparing Reagents" section on Page 4.
- 1. Perform agarose gel/ethidium bromide electrophoresis to analyze PCR product.
- 2. Determine the volume of your PCR reaction.
- 3. Transfer the sample into a clean 1.5 mL microcentrifuge tube.
- Add 4-5 volumes CP Buffer. For PCR products smaller than 200 bp, add 5 volumes CP Buffer and 0.4 V 100 % Isopropanol

Note: Volume refers to the size of your PCR reaction. For example, if your PCR reaction is 100 μ L and is smaller than 200 bp, you would use 500 μ L CP Buffer abd 40 μ L Isopropanol.

- Vortex to mix thoroughly. Briefly centrifuge to collect any drops from the inside of the lid.
- 6. Prepare the vacuum manifold according to manufacturer's instructions and connect the HiBind® DNA Mini Column to the manifold.
- 7. Transfer the entire sample to the HiBind® DNA Mini Column.

E.Z.N.A.® Cycle Pure Kit Vacuum Protocol

- 8. Switch on vacuum source to draw the sample through the column.
- 9. Turn off the vacuum.
- 10. Add 700 µL DNA Wash Buffer.

Note: DNA Wash Buffer must be diluted with ethanol before use. Please see the Preparing Reagents section on Page 4 for instructions.

- 11. Switch on vacuum source to draw the DNA Wash Buffer through the column.
- 12. Turn off the vacuum.
- 13. Repeat Steps 10-12 for a second DNA Wash Buffer wash step.
- 14. Transfer the HiBind® DNA Mini Column into a 2 mL Collection Tube (provided).
- 15. Centrifuge the empty HiBind® DNA Mini Column at maximum speed for 2 minutes to dry the column.

Note: This step is critical for removal of trace ethanol that may interfere with downstream applications.

- 16. Transfer the HiBind® DNA Mini Column into a clean 1.5 mL microcentrifuge tube (not provided).
- 17. Add 30-50 μL Elution Buffer, TE Buffer, or sterile deionized water directly to the center of column matrix.
- 18. Let sit at room temperature for 2 minutes.
- 19. Centrifuge at maximum speed for 1 minute.

Note: This represents approximately 80-90% of bound DNA. An optional second elution will yield any residual DNA, though at a lower concentration.

Troubleshooting Guide

Please use this guide to troubleshot any problems that may arise. For further assistance, please contact the technical support staff , toll free, at **1-800-832-8896**.

Possible Problems and Suggestions

Low DNA Yields			
Not enough CP Buffer added to sample	Add more CP Buffer as indicated. For DNA fragments <200 bp in size, add 5 volumes CP Buffer 0.4 V Isopropanol		
Water pH is too low (< 7.5)	Check the pH of the water, adjust the pH of the water to 8.0 using Tris-HCl (2M, pH 8.5)		
No DNA eluted			
DNA Wash Buffer was not diluted with 100% ethanol	Prepare DNA Wash Buffer as instructed on the bottle, or refer to page 3		
Optical densities do not agree with DNA yield on agarose gel			
Trace contaminants eluted from column will increase A260	Make sure to wash column as instructed in Steps 10-13 of either protocol, rely on agarose gel/ethidium bromide electrophoresis for quantification		
DNA sample floats out of well while loading agarose gel			
Ethanol not completely removed from column	Centrifuge as instructed in Step 14 of the centrifugation protocol and Step 15 of the vacuum protocol to completely dry the HiBind® matrix		

Ordering Information

The following components are available for purchase separately. (Call Toll Number (800-832-8896)

Product	Part Number
DNase/RNase-free microcentrifuge tubes, 1.5 mL, 500/pk, 10 pk/cs	SSI-1210-00
DNase/RNase-free microcentrifuge tubes, 2.0 mL, 500/pk, 10 pk/cs	SSI-1310-00
Vacuum Manifold	VAC-08
CP Buffer (200 mL)	PDR042
Elution Buffer (100 mL)	PDR048
DNA Wash Buffer (100 mL)	PS010
2 mL Collection Tubes	SS1-1370-00

Notes: