PointMan

PointMan[™] KRAS (codon 12/13/61) DNA Enrichment Kit

Instructions for use of PointMan KRAS DNA enrichment kit



KRAS User Manual EN 11.13

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Section 1 Kit Contents

Kit Contents: PointMan KRAS Kit

The PointMan KRAS kit contains 3 separate primer mixes and control primers. Please read tube contents carefully before preparing individual reactions

PointMan KRAS 12/13 primer mix (24 reactions BLACK) PointMan KRAS 61 primer mix (24 reactions BLACK)

Control primer mix KRAS 12/13 (24 reactions WHITE) Control primer mix KRAS 61 (24 reactions WHITE)

PointMan KRAS Mastermix with SYBRgreen (225 reactions GREEN)

Sequencing primer forward KRAS 12/13 (100 reactions YELLOW) Sequencing primer reverse KRAS 12/13 (100 reactions YELLOW) Sequencing primer forward KRAS 61 (100 reactions YELLOW) Sequencing primer reverse KRAS 61 (100 reactions YELLOW)

1% Positive control KRAS 12/13 (8 reactions RED) 1% Positive control KRAS 61 (8 reactions RED) RNase/DNase free water (CLEAR)

Section 2 Shipping and Storage

The PointMan KRAS kit is shipped frozen and must still be frozen upon arrival. If the PointMan KRAS kit is not frozen upon arrival, the outer packing has been opened during transit, the shipment does not contain a packing note, instruction booklet or the reagents please contact the EKF Molecular technical services department (info@ekfmolecular.com). The PointMan KRAS kit should be stored at -15°C to -25°C and protected from sunlight.

When stored under the recommended storage conditions in the original packaging, the PointMan KRAS kit is stable for 6 months from the date of purchase. Repeated thawing and freezing should be avoided. We recommend a maximum of 7 freeze-thaw cycles.

Section 3 Product Use Limitations

EKF Molecular Diagnostics sell products solely for use in laboratory research to gain information for use by the purchaser (the "Permitted Use"). By purchasing a PointMan enrichment kit, purchasers undertake that they are purchasing for the Permitted Use only and that purchasers will not use products for any other use, including without limitation: diagnostics for medical purposes or commercial purposes and will not resell any products. Products are covered by patent applications owned by or licenced to EKF Molecular Diagnostics (please see www. ekfmolecular.com). By purchasing any PointMan enrichment kit, purchasers acquire a non-exclusive licence to use the product for the Permitted Use only. EKF Molecular Diagnostics do not warrant that any use of a product will not infringe any patent or any other intellectual property rights whatsoever of any third party. Polymerase Chain Reaction (PCR) is covered by several patents owned by Hoffman-Roche Inc and Hoffman-LaRoche, Ltd. Purchase of EKF Molecular Diagnostics kits does not include or provide any licence with respect to any patents owned by Hoffman-LaRoche inc

Section 4 Product Warranty and Guarantee

We warrant to you that any product purchased from us will on delivery: conform in all material respects to its specification; be of satisfactory quality (within the meaning of the Sale of Goods Act 1979, as amended) and fit for the purpose held out by us; be free from material defects in design, material and workmanship and; comply with all applicable statutory and regulatory requirements.

You may reject any product which does not comply with the above; in the case of a defect that is apparent on normal visual inspection, within five business days of delivery and in the case of a latent defect, within a reasonable time of the latent defect having become apparent. If you reject a product described as above you may either require us to replace the rejected product or require us to repay the price of the rejected goods in full, following return of the goods.

Section 5 Quality Control

As part of our routine quality assurance program, all EKF Molecular Diagnostics products are manufactured to ISO9001 standards and monitored to ensure the highest levels of performance and reliability.

Section 6 Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. All chemicals and biological material must be considered as potentially hazardous. Specimens are potentially infectious and must be treated accordingly.

Discard sample and assay waste in accordance with your local safety regulations.

Section 7 Introduction to PointMan

There is an unmet clinical need for the accurate detection and genotyping of rare somatic mutations. The PointMan system is a novel method for selective amplification of genotype specific sequences. Unlike allele specific priming, PointMan does not, in the first instance, achieve results by attempting to selectively prime and amplify mutant sequences. Such methods may be error prone, need extensive optimisation and have limited resolution. By contrast, PointMan uses allele specific primers for priming on the wild-type (WT) sequence and this then drives those WT templates into terminally extended products that are not available for exponential amplification. The variant sequences are uninhibited and therefore free to undergo exponential amplification and enrichment.

The system uses 4 primers in total (fig.1). The internal primers confer the selective power of the assay and are called the enriching primers. The external primers are unmodified and are simply used to PCR amplify across the region of interest (amplifying primers).

Section 7 Introduction to PointMan

The enriching primers have the 3' terminal residue situated over the SNP and this residue is homologous with the WT sequence. A second important feature of the enriching primers is that they also contain a PCR blocking residue. Whilst this does not prevent binding of the primer and extension on the genomic DNA template, it does prevent the enriching primers from participating in the second and all subsequent rounds of PCR amplification (fig.1). The single stranded template made by the enriching primer on the WT sequences blocks the extension of the full length amplicon from the amplifying primer. In this way, exponential amplification on WT sequences is terminated.

On variant sequences, the enriching primer does not extend as it is displaced by the enzyme during the extension step. This allows for full exponential amplification across the variant region (fig.1).

The PointMan KRAS kit will exponentially amplify deletions and alteration in the sequences highlighted in red below. For a full list of the mutations enriched please visit: www.ekfmolecular.com

- Codon 12 3' GTT GGA GCT GGT GGC GTA GGC AAG AGT 5'
- Codon 13 3 GTT GGA GCT GGT GGC GTA GGC AAG AGT 5'
- Codon 61 3'GAC ACA GCA GGT CAA GAG GAG TAC AGT 5'



Figure 1: PointMan amplifies the SNP sequence but blocks amplification of the WT sequence

<u>Step 1 & 2 Wild Type</u> Enriching primers (\longrightarrow) containing blocking moiety (\times) complementary to the wild-type sequence bind with high avidity to sample DNA

<u>Step 1</u> Enriching primers (\longrightarrow) with blocking moiety (\times) complimentary to the wild-type sequence and a mismatch at the 3' end to the variant (mutant) sequence (the base of interest is shown as: wild-type – blue oval \bigcirc ; variant - red oval \bigcirc). Amplifying primers (\longrightarrow) complimentary to sequences flanking the region of interest.

Step 2 Wild-Type. Enriching primers anneal to wild-type with high avidity due to match at 3' end; Extension from enriching primers occurs, further increasing avidity. Amplifying primers anneal and extend, but extension is terminated by presence of the extending enriching primer.

Variant. Enriching primers do not efficiently anneal and extend due to mismatch at the 3' end and stringency of PCR conditions. Amplifying primers therefore extend right through the region of interest, without hindrance.

Steps 3 & 4 Wild-Type. In the subsequent cycle, only the product of extension from the enriching primers provides priming sites for further replication, but extension is terminated by the blocking moieties, and therefore the product does not itself contain any priming sites. Therefore no further replication is possible in subsequent cycles, and the reaction is driven into linear amplification based only on the original sample material.

Variant. In subsequent cycles, stemming from the full replicons unencumbered by blocking moieties, exponential amplification proceeds in the usual way.

Note that there is no risk of false positives from mispriming: no extraneous source of the variant sequence is introduced in the form of primers (unlike allele-specific PCR and its variants).

Section 8 Equipment and Reagents to Be Supplied by User

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

- DNA sample (see below)
- Dedicated pipettes (adjustable) for PCR master mix preparation*
- Dedicated pipettes (adjustable) for dispensing of sample DNA*
- Sterile pipette tips with filters
- Benchtop centrifuge with rotor for 1.5 ml reaction tubes
- Real-time PCR instrument
- Sterile microcentrifuge tubes for preparing master mixes

Section 9 Suitable Sample Material

PointMan will work with any high quality source of extracted human DNA and is therefore compatible with a wide range of extraction protocols. DNA samples should be quality checked and quantified using a UV spectrophotometer or similar approach.

General precautions

The user should always pay attention to the following:

Use sterile pipette tips with filters and make sure that pipettes have been calibrated according to the manufacturer's instructions.

Store and extract positive materials (samples and positive controls) separately from all other reagents and add them to the reaction mix in a spatially separated facility or within a laminar flow cabinet.

Thaw all components thoroughly on ice before starting an assay.

When thawed, mix the components by inverting each tube 10 times and centrifuge briefly. Do not vortex the mastermix as this may inactivate the Taq DNA polymerase.

Use extreme caution to prevent contamination of PCRs with synthetic control material. We recommend using separate, dedicated pipettes for setting up reaction mixes and adding DNA template. Preparation and dispensing of reaction mixes must be carried out in a separate area to the addition of sample.

Reagents for the PointMan KRAS kit have been optimally diluted. EKF Molecular Diagnostics do not recommend further dilution of reagents as this may result in a loss of performance. We do not recommend using reaction volumes of less than 15 μ l as this may increase the risk of false negatives.

All reagents in the PointMan KRAS Kit are formulated specifically for use with the stated tests. All reagents supplied in the PointMan KRAS Kit are intended to be used solely with the other reagents in the same Kit. Substitutions to the reagents in the kit must not be made if optimal performance is to be maintained.

Only use the Taq DNA polymerase that is provided in the kit. Do not substitute with Taq DNA polymerase from other kits of the same or any other type, or with Taq DNA polymerase from another supplier.

* Ensure that all instruments have been checked and calibrated according to the manufacturer's recommendations.

Section 10 Principles of the Test

PointMan Primer Mix (Black)

This primer mix contains the 4 primers required for a PointMan reaction. When used in conjunction with the PointMan[™] Mastermix and user supplied DNA sample, this reaction will selectively enrich the PCR reaction for variant sequences, if present. Following PointMan reaction completion, the enriched product can be sequenced to confirm presence at the variant sequence in the DNA sample.

Control Primer Mix (White)

The PointMan reaction blocks amplification of the wild type (WT) sequence, thus it follows that clinical samples which contain only the WT sequence may not produce any amplification product. Experimentally, it has been demonstrated that if the sample contains less than 1000 copies of WT sequence, no PCR product will accumulate. For this reason it is essential to confirm that the sample has been extracted correctly and is competent to support PointMan PCR amplification. The control primer mix contains only the amplifying primers without the enriching primers. This mix will therefore produce a post reaction product on both WT and variant sequences. The control primer mix is used to confirm that a biological sample is competent for PCR and therefore valid for PointMan analysis. A positive signal from the control assay primers is essential to enable validation of negative results.

PointMan Mastermix with SYBRgreen (Green)

The PointMan Mastermix is specifically formulated to provide optimal blocking of amplification on WT sequences whist allowing efficient amplification of variant sequences. Consistent with the PointMan process outlined above, the enzyme is exonuclease deficient. The mastermix also contains SYBRgreen. This allows the monitoring of product accumulation in real time and also permits melting curve analysis to ensure that a single amplicon, corresponding to BRAF, has been amplified. Analysis of the SYBRgreen data following PointMan enrichment enables the user to validate that PointMan[™] has produced a PCR product prior to sample sequencing. The PointMan[™] Mastermix cannot be substituted for another enzyme and buffer system as this will greatly affect the sensitivity of the enrichment.

PointMan Sequencing Primers (Yellow)

To confirm the presence of variant sequences following PointMan[™] enrichment, samples can be sequenced using the sequence primers provided to prime the reaction. The sequencing primer is unlabelled and is supplied lyophilised.

Section 11 Bench-side Protocol

To minimise the risk of contamination with foreign DNA, EKF Molecular recommend that all pipetting be performed in a PCR clean environment. Ideally this would be a designated PCR cabinet. Filter tips are recommended for all pipetting steps.

1. Pulse-spin each tube in a centrifuge before opening.

This ensures all products are situated in the base of the tube and prevents reagent loss upon opening the tube.

2. Resuspend all components according to the table below using RNase/DNase free water supplied.

COMPONENT	VOLUME
PointMan primer mix	72 µl
Control primer mix	72 µl
Forward sequencing primer*	110 µl
Reverse sequencing primer*	110 µl
1% positive control **	100 µl

* 3.2 pmol per µl when resuspended

** Contamination risk - contains high copy number of positive control template

Preparation of a PointMan reaction

Please note that the PointMan KRAS enrichment kit contains 3 separate primer mixes and control primers. Please read tube contents carefully before preparing individual reactions.

1. Make up reaction mixes

For each sample make up a reaction mix for both the PointMan test and the control well. Remember to include additional reactions for the positive and negative controls.

COMPONENT	1 REACTION
PointMan primer mix (BLACK)	2 µl
PointMan Mastermix with SYBRgreen (GREEN)	10 µl
RNAse/DNAse free water (CLEAR)	3 µl
Final volume	15 µl
Control primer mix (WHITE)	2 µl
PointMan Mastermix with SYBRgreen (GREEN)	10 µl
RNAse/DNAse free water (WHITE)	3 µl
Final volume	15 µl

2. Pipette 15 μl of relevant reaction mix into each well according to your PointMan experimental plate set up. e.g.

	SAMPLE 1	SAMPLE 2	SAMPLE 3	SAMPLE 4	SAMPLE 5	POSITIVE CTRL	NTC
PointMan							
Control primer		\bullet					•

3. Add 5 μl of DNA sample to each reaction

Ideally 1-10 ng of high quality DNA should be used. For no template controls (NTC)/ negative control wells replace sample with RNAse/DNAse free water. For the 1% positive control well, use 5 μ l of control template.

Section 12 Amplification Protocol

PointMan uses a carefully optimised 4 step cycling parameter. Please follow this protocol exactly using a block based real time PCR machine.

COMPONENT	STEP	TIME	TEMP
	Enzyme Activation (if required)	2 mins	95 °C
	Denaturation	10 s	95 °C
	Primer Annealing	20 s	50 °C
SUCICLES	Enriching Primer Displacement	1 s	70 °C
	Extension*	30 s	60 °C
MELT CURVE	60-90 °C in 1 °C intervals *		

* Flourogenic data should be collected during these steps through the SYBRgreen/FAM channel

Section 13 Interpretation of Results

Real time PCR traces

Analysis of the real time PCR data should give amplification plots in the region of CT=25-40.

Please note that variant traces for codon 12/13 will appear before the WT traces due to exponential replication of variant sequence.



Melting curve analysis

The melting curve for codon 12/13 should give a single peak with a melting temperature of 80° C as shown below.



Section 13 Interpretation of Results



Melting curve analysis

The melting curve for codon 61 should give a single peak with a melting temperature of 80.5° C as shown above.



Section 14 Table 1: Data Interpretation

CONTROL PRIMERS	POINTMAN PRIMERS MIX	INTERPRETATION
Amplicon produced	Amplicon produced with melting Tm 80 °C	Positive for a variant sequence. Confirm by sequence analysis
Amplicon produced	NO Amplicon produced	Reaction successful but no variant sequences amplified.
NO Amplicon produced	NO Amplicon produced	The sample has failed. Either there was insufficient amplifiable DNA or PCR inhibitors have prevented a successful test

Mutation Details

Numerous mutations and deletions in KRAS exons 12, 13 and 61 are reported, the most common are listed below. For a full list of the possible mutations enriched by this component of the kit, please visit www.ekfmolecular.com

Cosmic IDs are taken from the catalogue of Somatic Mutations in Cancer (www.sanger.ac.uk/genetics/CGP/cosmic).

Section 14 Table 1: Data Interpretation

Table 2: List of KRAS mutations and COSMIC IDs.

Mutation	Sequence	Base Change	Cosmic ID
Gly12Asp	3'GTAGTTGGAGCTGATGGCGTAGGCAAGAGT5'	35 G>A	521
Gly12Ala	3'GTAGTTGGAGCTG <mark>C</mark> TGGCGTAGGCAAGAGT5'	35 G>C	522
Gly12Val	3'GTAGTTGGAGCTGTTGGCGTAGGCAAGAGT5'	35 G>T	520
Gly12Ser	3'GTAGTTGGAGCTAGTGGCGTAGGCAAGAGT5'	34 G>A	517
Gly12Arg	3'GTAGTTGGAGCTCGTGGCGTAGGCAAGAGT5'	34 G>C	518
Gly12Cys	3'GTAGTTGGAGCTTGTGGCGTAGGCAAGAGT5'	34 G>T	516
Gly13Asp	3'GTAGTTGGAGCTGGTG <mark>A</mark> CGTAGGCAAGAGT5'	38 G>A	532
Gln61Lys	3'GAC ACA GCA GG <mark>A A</mark> AA GAG GAG TAC agt5'	180_181 TC>AA	87298
Gln61Lys	3'GAC ACA GCA GG <mark>C A</mark> AA GAG GAG TAC AGT5'	180_181 TC>CA	12729
Gln61Lys	3'GAC ACA GCA GGT AAA GAG GAG TAC AGT5'	181 C>A	549
Gln61Glu	3'GAC ACA GCA GGT GAA GAG GAG TAC AGT5'	181 C>G	550
Gln61Arg	3'GAC ACA GCA GGT C <mark>GT</mark> GAG GAG TAC AGT5'	180_181 AA>GT	1168052
Gln61Lys	3'GAC ACA GCA GGT CTA GAG GAG TAC AGT5'	182 A>T	553
Gln61Arg	3'GAC ACA GCA GGT CGA GAG GAG TAC AGT5'	182 A>G	552
Gln61Pro	3'GAC ACA GCA GGT CCA GAG GAG TAC AGT5'	182 A>T	551
Gln61His	3'GAC ACA GCA GGT CA <mark>C</mark> GAG GAG TAC AGT5'	183 A>C	554
Gln61His	3'GAC ACA GCA GGT CAT GAG GAG TAC AGT5'	183 A>T	555

Section 15 Notices and Disclaimers

These products are sold exclusively for research and development (R&D) use by the purchaser. PointMan enrichment kits may not be used for human or veterinary in vitro diagnostic (IVD) applications and they may not be re-sold, distributed or re-packaged without express written authorization from EKF Molecular Diagnostics.

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The purchase of EKF Molecular Diagnostics reagents cannot be construed as an authorisation or implicit license to practice PCR under any patents held by Hoffmann-LaRoche Inc or others.

PointMan[™] Mastermix containing GoTaq[®] Hot Start Polymerase manufactured by Promega Corporation for distribution by EKF Molecular Diagnostics. Licensed to Promega under U.S. Patent Nos. 5,338,671 and 5,587,287 and their corresponding foreign patents.

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