SABiosciences™

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User Manual

RT² Nano PreAMP cDNA Synthesis Kit

Synthesis and Pre-amplification of First Strand cDNA from RNA of Small Samples for Gene Expression Profiling with RT²Profiler PCR Arrays

NOTE: For Scientists using FFPE Samples, Please Refer to the User Manual for the RT² FFPE PreAMP cDNA Synthesis Kit at:

http://sabiosciences.com/support_manual.php

See Purchaser Notification for limited use license and warranty information (page 3).

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SABiosciences Corporation 6951 Executive Way, Frederick, MD 21703 USA

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Synthesis and Pre-amplification of First Strand cDNA from RNA of Small Samples for Gene Expression Profiling with RT² Profiler PCR Arrays

User Manual

(For Catalog Number: C-06)

NOTE: For Scientists using FFPE Samples, Please Refer to the User Manual for the RT² FFPE PreAMP cDNA Synthesis Kit at:

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Ordering and Technical Service Contact Information:

- Tel: 1-888-503-3187 (US) 301-682-9200 (outside US)
- Fax: 1-888-465-9859 (US) 301-682-7300 (outside US)
- On-line Order: **www.SABiosciences.com**
- E-MAIL: **order@SABiosciences.com** (to place an order)
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- Your contact information (name, phone, email address)
- Product name, catalog number and quantity
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I. Background and Introduction

Recent advances in tissue preparation and RNA isolation procedures allow researchers to work with minute amount of RNA such as those from small cell population, laser capture microdissection (LCM) samples, fluorescence-activated cell sorting (FACS) samples or fine needle aspiration biopsies (FNAB). However, the nano-scale RNA obtained from these small samples is often too low of an amount for reliable gene expression analysis by even such sensitive techniques as real-time reverse transcription (RT) PCR.

RT² Nano PreAMP cDNA Synthesis Kit and Primer Mixes are a breakthrough technology enabling expression analysis of hundreds of pathway- or disease- focused genes using as little as 1 ng total RNA. Our RT² Nano PreAMP technology utilizes multiplex PCR-based pre-amplification to provide amplification of gene-specific cDNA target templates with minimal bias. This kit is intended for synthesis followed by pre-amplification of first strand cDNA from limited amount of total RNA samples for gene expression analysis with our RT²Profiler™ PCR Arrays. You can prepare enough cDNA from each RNA sample for gene expression analysis on as many as four different pathways. Two simple steps are involved in this kit:

- cDNA first strand synthesis
  This kit provides enough reagents for synthesizing first strand cDNA from 12 different samples. Our RT² First Strand cDNA synthesis system in this kit comes with a built-in external RNA control template that would be detected by the Reverse Transcription Control (RTC) tests in the RT² Profiler PCR Arrays. This allows the detection of any presence of inhibitors of reverse transcription, ensuring the efficiency of the first strand cDNA synthesis reactions.
- Pre-Amplification of cDNA for pathway-specific genes
  Each first strand cDNA synthesis reaction from 1ng to 100ng of total RNA can be amplified by 4 different sets of PCR Array-specific primer mixes, allowing gene expression analysis on as many as four different pathways. The Side Reaction Reducer eliminates the residual primers after pre-amplification, making the pre-amplified cDNA ready for PCR Array analysis.

To complete the PCR Array procedure, mix the amplified templates with one of our instrument-specific and ready-to-use RT² qPCR SYBR Green Master Mixes. Aliquot the mixture into each well of the same plate containing pre-dispensed gene-specific primer sets. Perform PCR, and finally, determine relative expression with your real-time instrument and the $\Delta\Delta C_t$ method.

Important Note: Each RT² Nano PreAMP cDNA Synthesis Primer Mix is PCR Array-specific and can only be used for the specified RT²Profiler PCR Arrays. The first strand synthesis components and the RT² Nano PreAMP reagents in this kit have been optimized hand-in-hand for SYBR Green real-time RT-PCR detection, further enhancing the sensitivity of our RT² Profiler PCR Arrays. The convenient and quick workflow of the RT² Nano PreAMP cDNA Synthesis Kit also makes this technology accessible for routine use in every research laboratory.

Benefits of RT² Nano PreAMP cDNA Synthesis Kit:

- **Robust Performance on Small Samples:** Demonstrated to provide high performance pre-amplification without bias
- **Easy Workflow and Designed for Routine Use:** Simple and quick procedures with minimal hands-on time to pre-amplify target templates under two hours
- **Superior Sensitivity:** Maximally enhances the sensitivity of RT²Profiler PCR Array to analyze limited amounts of cDNA

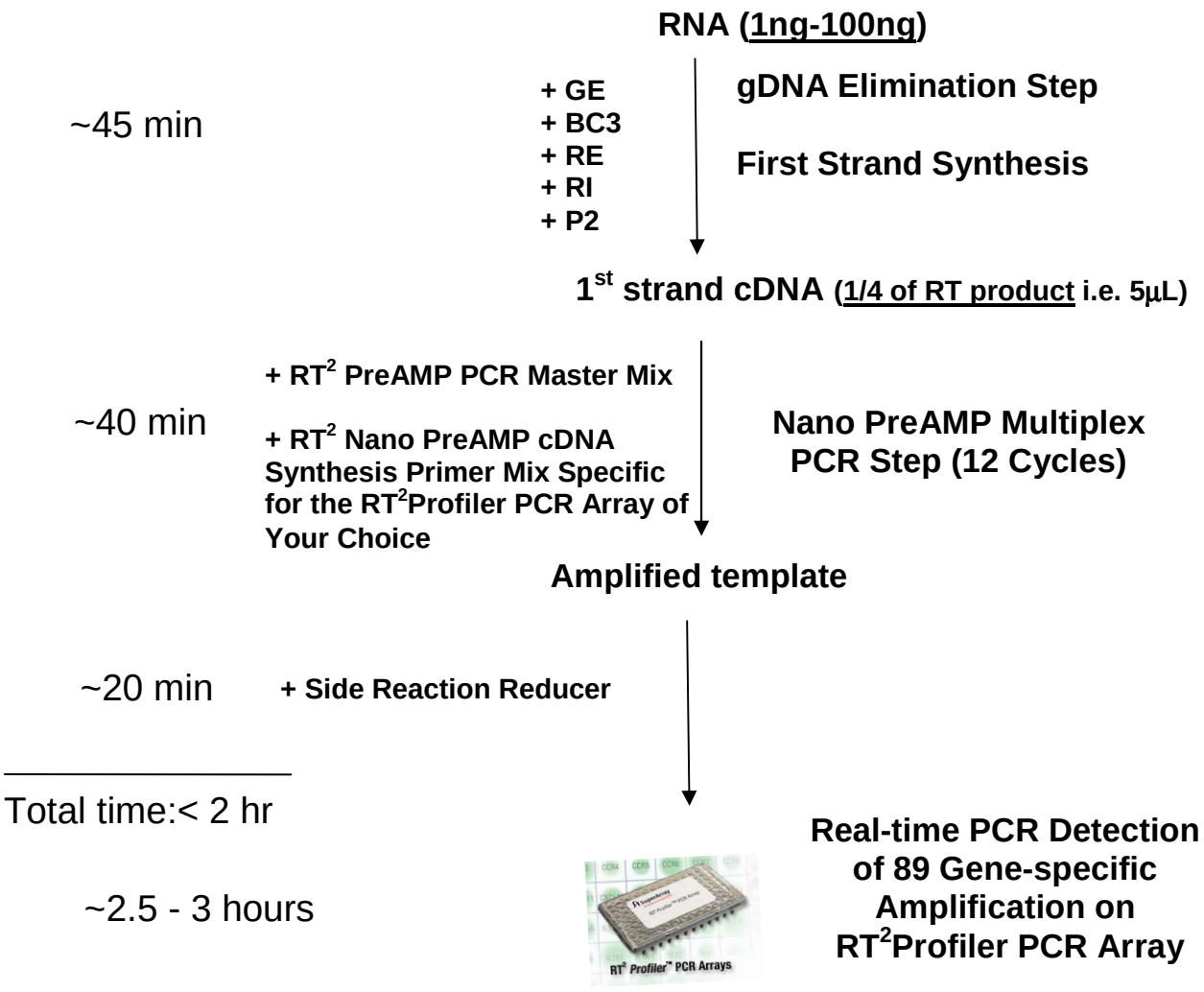

Figure 1: Workflow of RT² Nano PreAMP cDNA Synthesis Procedures

II. Materials Provided:

This kit includes enough of the following reagents for 12 RNA samples:

A. RT² First Strand cDNA Synthesis Components (enough for 12 20-µl RT reactions)

- **One (1) tube of GE (5X gDNA Elimination Buffer)**
- **One (1) tube of BC3 (5X Reverse Transcription Buffer 3)**
- **One (1) tube of RE (cDNA Synthesis Enzyme Mix)**
- **One (1) tube of RI (RNase Inhibitor)**
- **One (1) tube of P2 (Primer and External Control Mix)**
- **One (1) tube of RNase-free H_2O**

B. RT² Nano PreAMP components

- **One (1) tube of RT² PreAMP PCR Master Mix (PA-030) containing 600µl of 2X solution and enough for four 25-µl standard reactions per sample for 12 samples (48 reactions)**
- **One (1) tube containing 96µl Side Reaction Reducer (SR1) enough for four standard reactions per sample for 12 samples (48 reactions)**

Storage Conditions:

All components included in this kit are shipped on dry ice or blue ice packs and must be stored at -20℃ upon receipt. When stored properly at -20℃, their quality is guaranteed for 6 months.

Note: *Ensure that you do not contaminate the 2X RT² PreAMP PCR Master Mix and avoid repeated freezing and thawing by dividing into aliquots containing the amount of cocktail necessary for the number of reactions you are preparing each day. The rest of the cocktail should be kept in storage away from any sources of template DNA.*

III. Additional Materials Required:

A. RNA Isolation Kit: See Page 13 for specific recommendations.

B. RT^2 Nano PreAMP cDNA Synthesis Primer Mix (PB(H/M/R/Q/D)-XXXX): MANDATORY for a Successful and Unbiased Pre-Amplification of your samples—each primer mix is specific to a cataloged RT^{2}Profiler PCR Array. Check the label to verify that you have the correct pathway-specific RT^2 Nano PreAMP cDNA Synthesis Primer Mix for the PCR Arrays you are performing. Please also verify that the lot number of your Primer Mix is compatible with that of the RT^2 Profiler PCR Arrays you are going to use. We strongly recommend the Primer Mix and the PCR Arrays to be purchased together whenever possible. If the Primer Mix and the PCR Arrays have been purchased at different times, please check with our Customer service (1-888-503-3187) to ensure their compatibility.

C. SABiosciences RT^2 Profiler PCR Array: The PCR Arrays are available in six different plate formats, each tailored to a specific subset of real-time PCR instruments and associated blocks. Formats A, C, D, and F are 96-well plates, while Formats E and G are 384-well plates.

Format	For Real-Time Instruments	Plate
A	ABI "**standard**" blocks: 5700, 7000, 7300, 7500, 7700, 7900HT (96-block) Bio-Rad: iCycler®, iQ5, MyiQ, MyiQ2, Chromo4 (MJ Research) Eppendorf: MasterCycler® ep RealPlex® 2, 2s, 4, 4s Stratagene: Mx3005p®, Mx3000p® Takara: TP-800	96-well
C	ABI: 7500 **FAST** block, 7900HT **FAST** block, StepOnePlus™	96-well
D	Bio-Rad: CFX96™, Opticon® and Opticon 2 (MJ Research) Stratagene: Mx4000®	96-well
E	ABI: 7900HT (384-well block) Bio-Rad: CFX384™	384-well
F	Roche: LightCycler 480 96-well block	96-well
G	Roche: LightCycler 480 384-well block	384-well
H	Fluidigm BioMark	96x96 Chip

NOTE: *The format of the PCR Array is indicated by the last letter of the catalog number. Be sure that you have the correct PCR Array format for your instrument before starting the experiment.*

The 96-well PCR Arrays (Formats A, C, D, and F) are shipped in sets of two (2) or twelve (12), while the 384-well PCR Arrays (Formats E and G) are shipped in sets of four (4). Each PCR Array shipment includes the arrays and either twelve (12) optical thin-wall 8-cap strips (Formats A and D) or one (1) optical adhesive film (Formats C, E, F, and G) per array.

D. SABiosciences RT² qPCR Master Mix

MANDATORY for a Complete and Successful Experiment

Be sure to pick the correct one for the instrumentation in your laboratory.

1. 96-Well & 384-Well (96 x 4 Format) PCR Arrays

RT² SYBR Green / ROX qPCR Master Mix: Specifically designed for:

- **ABI** 5700,7000, 7300, 7500 (Standard & FAST), 7700, 7900HT 96-well block (Standard & FAST) and 384-well block, StepOnePlus
- **Eppendorf** Mastercycler ep realplex 2/2S/4/4S
- **Stratagene** Mx3000p, Mx3005p, Mx4000
- **TaKaRa** TP-800

Catalog Number	Size
PA-012	For 2 96-well RT² Profiler PCR Arrays
PA-012-12	For 12 96-well RT² Profiler PCR Arrays
PA-012-24	For 24 96-well RT² Profiler PCR Arrays
PA-012-8	For 4 384-well RT² Profiler PCR Arrays

RT² SYBR Green / Fluorescein qPCR Master Mix: Specifically designed for:

- **Bio-Rad** iCycler, iQ5, MyiQ, MyiQ2

Catalog Number	Size
PA-011	For 2 96-well RT² Profiler PCR Arrays
PA-011-12	For 12 96-well RT² Profiler PCR Arrays
PA-011-24	For 24 96-well RT² Profiler PCR Arrays
PA-011-8	For 4 384-well RT² Profiler PCR Arrays

RT² SYBR Green qPCR Master Mix: Specifically designed for instrumentation that does not require a reference dye:

- **Bio-Rad** CFX96, CFX384, Chromo4, Opticon 2
- **Roche** LightCycler 480 (96-well & 384-well)
- **All Others**

Catalog Number	Size
PA-010	For 2 96-well RT² Profiler PCR Arrays
PA-010-12	For 12 96-well RT² Profiler PCR Arrays
PA-010-24	For 24 96-well RT² Profiler PCR Arrays
PA-010-8	For 4 384-well RT² Profiler PCR Arrays

E. Equipment:

1. A conventional programmable thermal cycler with 0.2mL tube heat block, heated lid and 10-100 µL reaction capacity.
2. For recommendations on specific real-time instrumentation (thermal cyclers with fluorescent detection), see the list of master mixes and plate formats above.

NOTE: *The PCR Arrays can only be used in 96-well and 384-well real-time PCR instruments. PCR Arrays can not be used in the Cepheid SmartCycler®, the Roche LightCycler® 2.0, or the Corbett Research Rotorgene.*

3. 0.2 mL individual or 8-tube strip PCR tubes with caps
4. Calibrated P2, P20, P200 and P1000 Single Channel Pipettors
5. Calibrated Multi-Channel Pipettor
6. RNase / DNase-free pipette tips and tubes

IV. Complementary Products:

XpressRef™ Universal Total RNA:
Universal RNA to control PCR conditions is available from the following species:

Human XpressRef™ Universal Total RNA	Cat. No. GA-004
Mouse XpressRef™ Universal Total RNA	Cat. No. GA-005
Rat XpressRef™ Universal Total RNA	Cat. No. GA-006

V. Quick Protocol:

NOTE: For Scientists using FFPE Samples, Please Refer to the User Manual for the RT² FFPE PreAMP cDNA Synthesis Kit at: http://sabiosciences.com/support_manual.php

1. Before starting the experiment, make sure both the pathway and the lot number of your RT² Nano PreAMP cDNA Synthesis Primer Mix are compatible with those of your RT²Profiler PCR Array (Refer to page 4 and page 11 for details).

   - Ensure that you do not contaminate the 2X RT² PreAMP PCR Master Mix by dividing into aliquots containing the amount of cocktail necessary for the number of reactions you are preparing each day. The rest of the cocktail should be kept in storage away from any sources of template DNA.

2. Perform first strand synthesis as follows:
   a. Add 2 µl of GE to 8 µl of RNA (1-100ng).
   b. Incubate at 42℃ for 5 min and immediately chill on ice.
   c. Mix a master mix for the RT reaction as below:

	For 1 reaction:
BC3	4 ul
RE	1 ul
RI	1 ul
P2	1 ul
RNase-free H_2O	3 ul

   d. Add 10 µl of the RT master mix to 10 µl GE-treated RNA.
   e. Incubate at 42℃ for 30 min and heat at 95℃ for 5 min.
   f. Chill on ice or store at -20℃ until use.

3. For the normal standard reaction, mix the following components in a PCR tube:

12.5 µl	2X RT² PreAMP PCR Master Mix (PA-030)
7.5 µl	RT² FFPE PreAMP Primer Mix for the RT²Profiler™ PCR Array of your choice
5.0 µl	Template: undiluted cDNA from a 20-µl first strand synthesis reaction
25.0 µl	final volume

4. Perform 12 cycles of PCR in a thermal cycler:
   NOTE: *The 10 min step at 95 ℃ is required to activate t he HotStart Taq DNA polymerase.*

   95℃, 10 min ; 12 cycles of (95℃, 15 sec; and 60℃, 2 min); 4℃ forever

5. Add 2 µl of the Side Reaction Reducer (SR1) to each pre-amplified reaction, and incubate at 37 ℃ for 15 min followed by heat inactivation at 95 ℃ for 5 min.

6. Dilute the 27-µl pre-amplified templates to 111 µl by adding 84 µl of dd H_2O. Use immediately and keep on ice prior to loading onto the RT²Profiler™ PCR Array or store at -20℃ until use.

7. For use in the RT²Profiler™ PCR Array, mix well the following components in 15-mL conical tube:

1350 µl	2X SABiosciences RT² qPCR SYBR Green Master Mix (Note: Use the appropriate master mix specific for your real-time PCR instrument)
102 µl	diluted PreAMP PCR reaction (from Step 5)
1248 µl	ddH_2O
2700 µl	final volume

8. Add 25 µl of the above Experimental Cocktail to each well of the PCR Array, preferably from a reservoir with an eight-channel pipettor (or a twelve-channel pipettor but only using eight tips).

9. Run the following real-time thermal cycler program:
   NOTE: *The 10 min step at 95°C is required to activate th e HotStart Taq DNA polymerase.*

   95°C, 10 min ; 40 cycles of (95°C, 15 sec; and 60°C, 60 sec)

10. Program the real-time thermal cycler to detect and record the SYBR® Green I signal from every reaction during the annealing step of each cycle.

VI. Detailed Protocol:

NOTE: For Scientists using FFPE Samples, Please Refer to the User Manual for the RT² FFPE PreAMP cDNA Synthesis Kit at: http://sabiosciences.com/support_manual.php

Please read through this entire protocol before beginning your experiment.
RNA samples are very sensitive to RNase digestion; therefore, wear gloves and maintain an RNase-free work area while performing this protocol.

Important Note: Please note that each RT² Nano PreAMP cDNA Synthesis Primer Mix is specific to a catalogued RT²Profiler PCR Array and can only be used for the intended PCR Array. Please also verify that the lot number of your Primer Mix is compatible with that of the RT² Profiler PCR Arrays you are going to use. Otherwise, the Nano PreAMP process may not work optimally. If the Primer Mix and the PCR Arrays have been purchased at different times, please check with our Customer service (1-888-503-3187) to ensure their compatibility.

NOTE: Master Mix and First Strand Synthesis Considerations

The performance of our RT² Nano PreAMP cDNA Synthesis Kit is only guaranteed with our RT² Nano PreAMP cDNA Synthesis Primer Mix and our RT² Profiler PCR Arrays and RT² qPCR SYBR Green Master Mixes. In addition, the RT² PreAMP PCR Master Mix in this kit is specially formulated for multiplex PCR-based pre-amplification with our RT² Nano PreAMP cDNA Synthesis Primer Mix for optimal results without introducing bias.

The chemically-modified and tightly controlled HotStart enzyme and other proprietary chemical components in our RT² qPCR Master Mixes uniquely provide more accurate SYBR Green results by preventing the amplification of primer dimers and other non-specific products. They also help ensure high amplification efficiencies even for those genes that are the most difficult to amplify. When we test other sources of enzymes with our PCR Arrays, we frequently see primer dimers and other non-specific products that confound SYBR Green-based real-time PCR detection. Because each instrument uses a different reference dye to normalize their optics, be sure that you use the correct master mix for the instrumentation in your laboratory.

Our RT² First Strand cDNA Synthesis components in this kit include a proprietary buffer to eliminate any residual genomic DNA contamination in your RNA samples before it can be

amplified into secondary products that would otherwise cause false positive signals. The Reverse Transcription Controls (RTC) on our RT^2 Profiler PCR Array can only be evaluated with the built-in external RNA control of our RT^2 First Strand cDNA Synthesis components. These controls do not yield results when used with other sources of reverse transcriptases or first strand synthesis kits. The buffer components and the magnesium concentration in our RT^2 First Strand cDNA Synthesis components are also more compatible with our RT^2 PreAMP PCR and RT^2 qPCR master mixes than other enzymes or kits, providing the PCR Arrays with maximum levels of sensitivity with ng of total RNA.

NOTE: Preparing a Workspace Free of DNA Contamination

For accurate and reproducible PCR Array results, it is very important to avoid contamination of the assay with foreign DNA. Any DNA contamination will artificially inflate the SYBR Green signal yielding skewed gene expression profiles and false positive signals. The most common sources of DNA contamination are the products of previous experiments spread into the air of your working environment. Please follow the recommendations below on how to set up and maintain a working environment free of DNA contamination.

1. Wear gloves throughout the procedure. Use only fresh PCR-grade reagents (H_20) and lab ware (tips and tubes).

2. Physically separate the workspaces used for PCR setup and post-PCR processing or non-PCR operations. Decontaminate your PCR workspace and lab ware (pipettor barrels, tube racks, etc.) before each new use with UV light to render any contaminating DNA ineffective in PCR through the formation of thymidine dimers or with 10% bleach to chemically inactivate and degrade any DNA.

3. Close all tubes containing PCR products once you are finished adding or removing volumes. Before discarding any lab ware (tips or tubes) containing PCR products or other DNA, treat with 10% bleach.

4. Do not remove the PCR Array plate from its protective sealed bag until immediately ready to use. Do not leave lab ware (tubes and tip boxes) exposed to the air for long periods of time.

5. Do not open any previously run and stored PCR Array plate. Removing the thin-wall 8-cap strips or the adhesive film from PCR Arrays releases PCR product DNA into the air where it will contaminate and confound the results of future real-time PCR experiments.

A. RNA Preparation, Quality and Amount Considerations

High quality RNA is ESSENTIAL for obtaining good real-time PCR results.

The most important prerequisite for any gene expression analysis experiment is consistent, high-quality RNA from every experimental sample. Therefore, the sample handling and RNA isolation procedures are critical to the success of the experiment. Residual traces of proteins, salts or other contaminants will either degrade the RNA or decrease the efficiency of (if not block completely) the enzyme activities necessary for optimal reverse transcription, Nano PreAMP PCR and real-time PCR performance.

1. **Recommended RNA Preparation Methods:**

   Laser Capture Microdissected (LCM) Samples: Capture LCM samples on the Arcturus Capsure® caps and purify the RNA using the Arcturus PicoPure® RNA isolation kit following the manufacturer's instructions. Special care needs to be taken during the sample preparation and microdissection process to minimize RNA degradation. For example, LCM sessions longer than 30 minutes per slide lead to precipitous loss in recovery of intact RNA (1). Thus, shorten the entire staining and dissection procedure as much as possible.

   Fine Needle Aspiration Biopsies (FNAB) or Other Small Biological Samples: Use SABiosciences' RT² qPCR-Grade RNA Isolation Kit (Catalog # PA-001) or other similar commercial product for most small samples with a small number of cells such as fine needle aspiration biopsies, manual dissection, or FACS. In addition, the Arcturus PicoPure® RNA isolation kit can also be used for non-LCM samples such as cell samples in suspension. For any isolation kit that you use, it is important that you perform the recommended DNase treatment step.

   Total RNA Isolated Using a Phenol-Based Method: If you have already prepared total RNA from any biological source material using a phenol-based method (such as TRIzol®, RNAzol, etc.), you must clean up the RNA with SABiosciences' RT² qPCR-Grade RNA Isolation Kit (Catalog # PA-001) or the Qiagen RNeasy® Mini Kit (Catalog # 74104). You must perform the recommended on-column DNase treatment step.

2. RNA Quality Control:

For the optimal performance of RT^2 Nano PreAMP cDNA Synthesis procedures and best results from the PCR Array, all RNA samples should be suspended in the RNase-free water provided with the RNA Isolation kit. DO NOT use DEPC-treated water!

If you have at least 2 ng of RNA, you may electrophorese a fraction of each RNA sample (200 pg) on an Agilent BioAnalyzer® using an RNA 6000 Pico LabChip®. Verify that there is a sharp distinction at the small side of both the 18S and 28S ribosomal RNA (rRNA) peaks. Any smearing or shoulder to the rRNA peaks indicates that degradation has occurred in the RNA sample.

Due to the extremely low yield of RNA from small samples, it is often impossible to confirm the quality of total RNA using any analytical means. Therefore, it is essential to choose the proper RNA isolation method for your biological sample as described above, and follow the manufacturers' protocol very carefully.

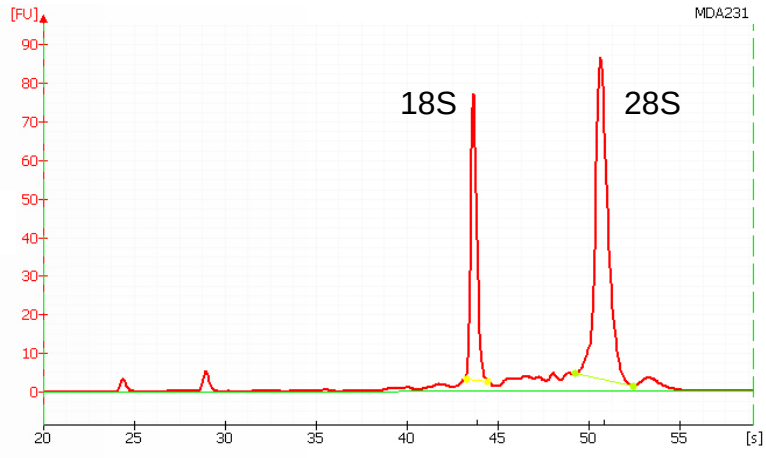

Figure 2: Good Ribosomal RNA Band Integrity Is Important for Optimal PCR Array Results.
The figure displays an Agilent BioAnalyzer electropherogram of a high-quality total RNA preparation showing sharp peaks without shoulders (especially to the left of each peak) for the 18S and 28S ribosomal RNA (left to right).

3. Genomic DNA Contamination:

Eliminating genomic DNA contamination is essential for obtaining optimal real-time gene expression profiling results using the PCR Array. The Genomic DNA Contamination (GDC) Control in each PCR Array specifically tests for genomic DNA contamination in each sample during each run. A GDC threshold cycle (C_t) value with the inclusion of pre-amplification step less than 30 indicates the presence of a detectable amount of genomic DNA contamination that should be addressed.

We highly recommend performing the on-column DNase treatment step in the RT² qPCR-Grade RNA Isolation Kit (PA-001) or the Qiagen RNeasy® Mini Kit (Catalog # 74104) followed by using the gDNA Elimination Buffer (GE) in this kit to remove any and all residual contamination from your RNA samples.

4. Amount Considerations:

Quantifying total RNA isolated from very small samples using UV spectrophotometry is often impossible. Instead, estimate the total RNA amount by assuming that each eukaryotic cell contains an average of ~10 pg of total RNA or less. Please note that the yield of RNA varies widely from cell type to cell type.

The RT² Nano PreAMP cDNA Synthesis Kit generates sufficient templates for gene expression analysis in RT²Profiler PCR Arrays with as little as 1 ng or as much as 100 ng total RNA input into each first strand cDNA synthesis reaction prior to pre-amplification step. Each RT reaction allows the user to perform as many as four RT² Nano PreAMP PCR reactions and four RT²Profiler PCR Arrays. However, the optimal amount of starting material depends on the relative abundance of the transcripts of interest. Lower abundance transcripts require more RNA; higher abundance transcripts require less RNA. Greater amounts of input total RNA yield a greater number of positive calls; that is, genes expressed in the linear dynamic range of the method. Lower amounts of input total RNA yield a smaller number of positive calls and increase false negative calls.

The RT² First Strand cDNA Synthesis components in this kit is optimized for the subsequent RT² Nano PreAMP procedures and maximizes the number of positive calls at low amounts (1 ng) of total RNA. For successful results and maximum positive call rates, we recommend that first-time users try starting with anywhere from 10ng to 50ng of total RNA. It is also important to use a consistent amount of total RNA for all samples in a single experiment to be characterized and compared.

B. First Strand cDNA Synthesis

NOTE: *The use of RT^2 First Strand cDNA Synthesis Components in this kit is critical for successful pre-amplification of templates for the RT^{2}Profiler PCR Arrays and detecting the Reverse Transcription Controls (RTC) in the PCR Arrays. For more information on the importance of these components, refer to the notes found on Pages 11 and 15.*

NOTE: *RNA samples must meet the standards of integrity and purity from protein, organics, and genomic DNA contamination described on the previous three pages.*

1. **Clean up RNA by eliminating genomic DNA contamination:**
   a. For each RNA sample, combine the following in a sterile PCR tube:

Genomic DNA Elimination Mixture**	1 reaction
Total RNA	1 ng to 100 ng
GE** (5X gDNA Elimination Buffer)	2 µl
RNase-free **H_2O**	Varied
Final Volume	10 µl

****The RT^2 First Stand Kit (C-03) is not compatible with the chemicals in Ambion's DNA-*free*™ kits. If your RNA sample has been treated with Ambion's DNA-*free*™ reagents, please call SABiosciences Technical Support at 1-888-503-3187.**

NOTE: *Use the similar amount of total RNA in this reaction for every sample. First-time RT^2 Nano PreAMP users are recommended to start with 10ng to 50ng of total RNA. Lower amounts of total RNA than 1 ng will dramatically affect the performance of the pre-amplification process.*

   b. Mix the contents gently with a pipettor followed by brief centrifugation.
   c. Incubate at 42℃ for 5 min & Chill on ice immedi ately for at least one minute.

2. **Prepare the RT Cocktail:**

RT Cocktail	1 reaction**
BC3 (5X Reverse Transcription Buffer 3)	4 µl
P2 (Primer & External Control Mix)	1 µl
RE (cDNA Enzyme Synthesis Mix)	1 µl
RI (RNase Inhibitor)	1 µl
RNase-free **H_2O**	3 µl
Final Volume	10 µl

**Scale up the volume for each of the RT Cocktail reagents accordingly when multiple reactions are carried out to minimize pipetting variations.

3. **First Strand cDNA Synthesis Reaction:**
   a. Add 10 µl of **RT Cocktail from Step B.2** to each 10-µl **Genomic DNA Elimination Mixture from Step B.1**.
   b. Mix well but gently with a pipettor. Spin the tubes briefly to remove any air bubbles and collect all the liquid to the bottom.
   c. Incubate at 42℃ for ***exactly*** 30 min and then ***immediately*** stop the reaction by heating at 95℃ for 5 minutes.
   d. Hold the finished First Strand cDNA Synthesis Reaction on ice until the next step (Step C) or store overnight at -20℃.

C. Pre-Amplification of cDNA Target Templates:

NOTE: *The use of SABiosciences' RT² Nano PreAMP cDNA Synthesis Primer Mix (Cat. No. PB(H/M/R/Q/D)-XXXX) is critical for a successful multiplex PCR pre-amplification of your cDNA and for obtaining the best results from the PCR Array.* Be sure to check the label to verify that you have the correct **pathway** and **lot number** of the RT² Nano PreAMP cDNA Synthesis Primer Mix for the PCR Arrays you are performing. (See pg 4 and pg 11 for details)

NOTE: *The accuracy and precision of your pipetting determine the consistency of your results. Be sure that all of your micro-pipettors are calibrated before beginning this procedure. Also, make sure to not introduce any bubbles into the wells of the PCR tubes.*

1. **Thaw the RT² PreAMP PCR Master Mix (PA-030) and RT² Nano PreAMP cDNA Synthesis Primer Mix at room temperature. If precipitates are observed, warm the reagents at 42℃ for 1 min and vortex briefly t o dissolve. Repeat the process if necessary.**

2. **Prepare the Nano PreAMP PCR Cocktail:**

RT² Nano PreAMP PCR Cocktail	1 reaction**
RT² PreAMP PCR Master Mix (PA-030) (2X Solution)	12.5 µl
RT² Nano PreAMP cDNA Synthesis Primer Mix (each specific for a RT²Profiler PCR Array)	7.5 µl
Final Volume	20 µl

**Scale up the volume for each of the Nano PreAMP PCR Cocktail reagents accordingly when multiple reactions are carried out to minimize pipetting variations.

3. **Nano PreAMP PCR Reaction:**

   a. Pipet 5 µl of the **First Strand cDNA Synthesis Reaction from Step B.3** into a 0.2-ml PCR tube**. Then** add 20 µl of the **Nano PreAMP PCR Cocktail (from Step C.2).**
   b. Mix well but gently with a pipettor. Spin the tubes briefly to remove any air bubbles and collect all the liquid to the bottom.
   c. Perform 12 cycles of PCR in a thermal cycler:

NOTE: *The 10-min step at 95℃ is required to activate th e HotStart Taq DNA polymerase.*

95℃, 10 min ; 12 cycles of (95℃, 15 sec; and 60℃, 2 min); 4℃ forever

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4. **Treatment with the Side Reaction Reducer**
   a. Take out the tubes from the thermal cycler and put on ice. Add 2 µl of the **Side Reaction Reducer (SR1)** to each pre-amplified reaction. Mix well but gently with a pipettor. Spin the tubes briefly to remove any air bubbles and collect all the liquid to the bottom.
   b. Incubate at 37℃ for 15 min followed by heat ina ctivation at 95℃ for 5 min.
   c. Add immediately 84 µl of RNase-DNase free H_2O to each 27-µl of Nano PreAMP PCR reaction. Mix well.
   d. Hold the finished Nano PreAMP PCR reaction on ice until the next step (Step D) or store overnight at -20℃.

D. Performing Real-Time PCR with RT^{2}Profiler PCR Arrays:

NOTE: *The use of SABiosciences' RT^2 qPCR Master Mixes is critical for obtaining the most accurate results from the PCR Array. Be sure to use the correct master mix for your instrument before continuing with this protocol (See Page 8).*

NOTE: *An incorrectly chosen PCR Array plate format will not properly fit into your real-time PCR instrument, and its use will damage the instrument. Be sure you have the correct PCR Array format for your instrument before continuing with this protocol (See Page 7).*

NOTE: *The accuracy and precision of your pipetting determine the consistency of your results. Be sure that all of your micro-pipettors are calibrated before beginning this procedure. Also, make sure to not introduce any bubbles into the wells of the PCR Array.*

1. **Experimental Cocktail Preparation:**

Plate Format:	**96-well A, C, D, & F**	**384-well E & G**
2X SABiosciences RT^2 qPCR SYBR Green Master Mix	1350 µl	550 µl
Diluted RT^2 Nano PreAMP PCR Reaction	102 µl	102 µl
RNase/DNase free H_2O	1248 µl	448 µl
Total Volume	2700 µl	1100 µl

NOTE: *This recipe provides an excess volume of* ONLY *~ 290 µl (for the 96-well format.) Very carefully add the cocktail to the PCR Array precisely to insure that each well receives the required volume.*

2. **For the rest of PCR Array protocol, please follow the instructions from Step C.2 to Step C.5 ("Performing Real-Time PCR with RT^{2}Profiler PCR Arrays") of the RT^2 Profiler PCR Array User Manual.**

E. Data Analysis: $\Delta\Delta C_t$ Method

Upon completion of PCR Array Run, Please access the free PCR Array Data Analysis Web Portal at:

http://www.SABiosciences.com/pcrarraydataanalysis.php.

Note: Please check the box in the "Readout" page for RT² Nano PreAMP cDNA Synthesis Kit.

The PCR Array Data Analysis Web Portal automatically performs the following calculations and interpretation of the control wells upon including threshold cycle data from a real-time instrument. The PCR Array Data Analysis Web Portal presents the results in a tabular format, a scatter plot, a three-dimensional profile, and a volcano plot (when replicates are included).

1. Any C_t value equal to or greater than 35 is considered a negative call. Due to the inclusion of the pre-amplification step, for the assays showing C_t values greater than 35, the expression level of these genes will be too low to be reliably quantified. Consider removing these data points from the rest of the results.

2. Examine the threshold cycle values of the control wells.

   a. Genomic DNA Control (GDC):

      i. Calculate C_t^{GDC}.
      ii. If the value is greater than 30, then the level of genomic DNA contamination is too low to affect gene expression profiling results. No action is needed.
      iii. If the value is less than 30, then genomic DNA contamination is evident. See the Troubleshooting and FAQ section.

   b. Reverse Transcription Control (RTC):
      Any impurities in your RNA sample that affect the reverse transcription of the RT² First Strand cDNA Synthesis' built-in external RNA control in this kit also affect the reverse transcription of your messages of interest.

      i. Calculate ΔC_t = AVG C_t^{RTC} – AVG C_t^{PPC}.
      ii. If this value is less than 7, then no inhibition is apparent.
      iii. If this value is greater than 7, then evidence of impurities that inhibited the reverse transcription phase of the procedure is evident. See the Troubleshooting and FAQ section.

c. Positive PCR Control (PPC):
Any impurities in your RNA sample that affect the PCR amplification of the positive control also affect the PCR amplification for your messages of interest.

i. The average C_t^{PPC} value should be 20 ± 2 on each PCR Array and should not vary by more than two cycles between PCR Arrays being compared.
ii. Larger differences in average C_t^{PPC} values between samples indicate the presence of different amounts of PCR amplification inhibitors in each sample and that all of the RNA samples require further purification.
iii. An average value of C_t^{PPC} that is consistently greater than 22 for all of your samples may indicate a problem with the cycling conditions or may simply be indicative of the relative sensitivity of your instrument. See the Troubleshooting and FAQ section.

3. Calculate the ΔC_t for each pathway-focused gene in each plate.

$$\Delta C_t = C_t^{GOI} - C_t^{AVG\ HKG}$$

NOTE: Choosing the right normalization factor
The expression level of the housekeeping genes chosen for normalization in the $\Delta\Delta C_t$ method must not be influenced by your experimental conditions. If one or more such genes have been previously identified by independent means and if the PCR Array reproduces those results, use the average of their C_t values in the equation above. If an appropriate housekeeping gene has not been previously identified, use the average C_t value of all housekeeping genes. Or, if your RNA samples were of sufficiently high concentration for reliable quantification such that equal amounts of RNA from all samples were used, simply use zero (0) in the place of the average of HK genes' C_t for each group to be compared, and rely on the consistency in the quantity and quality of your original input total RNA across your groups to effectively normalize your results.

4. When biological and/or technical replicates are performed, calculate the average ΔC_t value of each gene (each well) across those replicate arrays for each treatment group.

5. Calculate the $\Delta\Delta C_t$ for each gene across two PCR Arrays (or groups).
$\Delta\Delta C_t = \Delta C_t$ (group 2) - ΔC_t (group 1)
Where group 1 is the control and group 2 is the experimental

6. Calculate the fold-change for each gene from group 1 to group 2 as 2 ^ ($-\Delta\Delta C_t$).

OPTIONAL: *If the fold-change is greater than 1, then the result may be reported as a fold up-regulation. If the fold-change is less than 1, then the negative inverse of the result may be reported as a fold down-regulation. The fold-change ratios may also be reported as is.*

NOTE: Detailed Mathematical Explanation of $\Delta\Delta C_t$ Data Analysis Method

Due to the inverse proportional relationship between the threshold cycle (C_t) and the original gene expression level, and the doubling of the amount of product with every cycle, the original expression level (L) for each gene of interest is expressed as:

$$L = 2^{-C_t}$$

To normalize the expression level of a gene of interest (GOI) to a housekeeping gene (HKG), the expression levels of the two genes are divided:

$$\frac{2^{-C_t(GOI)}}{2^{-C_t(HKG)}} = 2^{-[C_t(GOI) - C_t(HKG)]} = 2^{-\Delta C_t}$$

To determine fold change in gene expression, the normalized expression of the GOI in the experimental sample is divided by the normalized expression of the same GOI in the control sample:

$$\frac{2^{-\Delta C_t(expt)}}{2^{-\Delta C_t(control)}} = 2^{-\Delta\Delta C_t} \qquad \text{Where } \Delta\Delta C_t \text{ is equal to } \Delta C_t(expt) - \Delta C_t(control)$$

The complete calculation is as follows:

$$\frac{\frac{2^{-\Delta C_t(GOI)}\,expt}{2^{-\Delta C_t(HKG)}\,expt}}{\frac{2^{-\Delta C_t(GOI)}\,control}{2^{-\Delta C_t(HKG)}\,control}} = \frac{2^{-[C_t(GOI) - C_t(HKG)]}\,expt}{2^{-[C_t(GOI) - C_t(HKG)]}\,control} = \frac{2^{-\Delta C_t}\,expt}{2^{-\Delta C_t}\,control} = 2^{-\Delta\Delta C_t}$$

VII. Alternative PreAMP Protocol for Individual RT^2 qPCR Primer Assays

1. **Perform First Strand cDNA Synthesis as in Part VI Step B.**
2. **Prepare the PreAMP Primer Mix:**
   To prepare 1 mL of the PreAMP primer mix for your selected gene(s) (up to ten genes) using SABiosciences' RT^2 qPCR Primer Assays, pipet 40 µl of each primer set into a 1.5 mL microcentrifuge tube and add DNase/RNase-free H_2O according to the table below. Vortex to mix well and spin down the tube briefly.

	RT^2 qPCR Primer Assay (10 µM)	**DNase/RNase-free H_2O**
1 gene	1 gene x 40 µl each = 40 µl	960 µl
2 genes	2 genes x 40 µl each = 80 µl	920 µl
3 genes	3 genes x 40 µl each = 120 µl	880 µl
4 genes	4 genes x 40 µl each = 160 µl	840 µl
5 genes	5 genes x 40 µl each = 200 µl	800 µl
6 genes	6 genes x 40 µl each = 240 µl	760 µl
7 genes	7 genes x 40 µl each = 280 µl	720 µl
8 genes	8 genes x 40 µl each = 320 µl	680 µl
9 genes	9 genes x 40 µl each = 360 µl	640 µl
10 genes	10 genes x 40 µl each = 400 µl	600 µl

**Note: One mL of the primer mix is sufficient for ~130 PreAMP reactions.

3. **Thaw the RT^2 PreAMP PCR Master Mix (PA-030) at room temperature. If precipitates are observed, warm the reagents at 42°C for 1 min and vortex briefly to dissolve. Repeat the process if necessary.**

4. **Prepare the PreAMP PCR Cocktail:**

RT^2 PreAMP PCR Cocktail	1 reaction**
RT^2 PreAMP PCR Master Mix (PA-030) (2X Solution)	12.5 µl
PreAMP Primer Mix	7.5 µl
Final Volume	20 µl

**Scale up the volume for each of the PreAMP PCR Cocktail reagents accordingly when multiple reactions are carried out to minimize pipetting variations.

5. **PreAMP PCR Reaction:**

   a. Pipet 5 µl of the **First Strand cDNA Synthesis Reaction** into a 0.2-ml PCR tube**. Then** add 20 µl of the **PreAMP PCR Cocktail (from Step 4).**
   b. Mix well but gently with a pipettor. Spin the tubes briefly to remove any air bubbles and collect all the liquid to the bottom.
   c. Perform 8 cycles of PCR in a thermal cycler:

NOTE: *The 10-min step at 95°C is required to activate th e HotStart Taq DNA polymerase.*

95°C, 10 min ; 8 cycles of (95°C, 15 sec; and 60°C, 2 min); 4°C forever

6. Treatment with the Side Reaction Reducer

a. Take out the tubes from the thermal cycler and put on ice. Add 2 µl of the **Side Reaction Reducer (SR1)** to each pre-amplified reaction. Mix well but gently with a pipettor. Spin the tubes briefly to remove any air bubbles and collect all the liquid to the bottom.
b. Incubate at 37℃ for 15 min followed by heat ina ctivation at 95℃ for 5 min.
c. Add immediately 28 µl of RNase-DNase free H_2O to each 27-µl of PreAMP PCR reaction. Mix well.
d. Hold the finished PreAMP PCR reaction on ice until the next step (Step 7) or store overnight at -20℃.

7. Perform Real-time RT² qPCR Primer Assays

NOTE: *The use of SABiosciences' RT² qPCR Master Mixes is critical for obtaining the most accurate results from the PCR Array. Be sure to use the correct master mix for your instrument before continuing with this protocol (See Page 8).*

NOTE: *The accuracy and precision of your pipetting determine the consistency of your results. Be sure that all of your micro-pipettors are calibrated before beginning this procedure. Also, make sure to not introduce any bubbles into the wells of the PCR Array.*
Experimental Samples:

a. To insure the consistency of your results and that each experimental sample yields a reliably detectable C_t value, we recommend setting up duplicate or triplicate reactions for each template.
b. For every experimental sample, prepare one set of reactions for every gene of interest and for a single housekeeping gene or a set of housekeeping genes to normalize your raw data. Choose housekeeping gene(s) known to not change their expression under your experimental conditions.
c. Positive and Negative Controls:
   - Prepare a positive control reaction using template known to represent the genes of interest such as template generated from SABiosciences' XpressRef™ Universal Total RNA.
   - To control for DNA contamination introduced during reaction setup, prepare a negative control reaction replacing template with water, the so-called no template control (NTC).
   - To control for genomic DNA contamination, perform one assay for each gene of interest and each housekeeping gene using an equivalent volume of product from the No Reverse Transcription (NRT) reaction performed for each RNA sample.

d. **Experimental qPCR Cocktail Preparation:**

	per 25-µl reaction
2X SABiosciences RT^2 qPCR SYBR Green Master Mix	12.5 µl
RNase/DNase free H_2O	10.5 µl
RT^2 qPCR Primer Assay (10uM)	1 µl
Diluted RT^2 PreAMP PCR Reaction	1 µl
Total Volume	25 µl

**Scale up the volume for each of the qPCR Cocktail reagents accordingly when multiple reactions are carried out to minimize pipetting variations.

e. Load 25 µl of each of the above reaction to PCR tubes or a PCR plate appropriate for your real-time thermal cycler.

f. **For the rest of qPCR assay protocol and data analysis, please follow the instructions from Step C.2 to Step C.4 ("Real-Time qPCR Primer Assay") of the RT^2 qPCR Primer Assays User Manual.**

VIII. Troubleshooting and FAQs

A. Troubleshooting:

1. High Ct Values for Many Genes including HouseKeeping Genes:

If the Ct values for all the housekeeping genes are high, this may suggest that either the total RNA input is less than the recommended lower limit of 1ng or the quality of RNA is poor and affects the success of pre-amplification. Double check the concentration and quality of your RNA samples following the suggestions outlined in RNA Quality Control and Amount Considerations at the beginning of the protocol in this User Manual.

2. Evidence of Genomic DNA Contamination:

You must perform the recommended DNase treatment step included in the protocol of SABiosciences' RT² qPCR-Grade RNA Isolation Kit (PA-001), Qiagen's RNeasy® Mini Kit (Catalog # 74104), or any other RNA isolation kit you choose to use. You must also then use the RT² First Strand cDNA Synthesis components in this kit with its genomic DNA elimination step.

If the genomic DNA contamination proves difficult to remove, fold-changes in gene expression may still be obtained. However, it will then be very important to validate any results for individual genes by a separate more rigorous real-time PCR analysis that includes a "minus RT" control.

Apparent genomic DNA contamination may also indicate evidence of more general DNA contamination of other reagents, tips, and tubes. See the Note about Preparing a Workspace Free of DNA Contamination at the beginning of the protocol in this User Manual.

2. Evidence of Poor Reverse Transcription Efficiency:

If possible, double-check the A260:A280 and A260:A230 ratios of your RNA samples in RNase-free Tris pH 8.0 buffer. If necessary, re-purify your RNA samples with a spin-column based clean up method, such as SABiosciences' RT² qPCR-Grade RNA Isolation Kit (PA-001).

3. Improving Poor PCR Amplification Efficiency:

Different instruments have different levels of sensitivity. If an average C_t^{PPC} value of 20 ± 2 is difficult to obtain for your instrument, the observed average C_t^{PPC} value should be acceptable as long as it does not vary by more than two cycles between PCR Arrays being compared.
Be sure that the initial heat activation step at 95 ℃ had been lengthened to 10 minutes from the shorter time in the default program. Be sure that all other cycle parameters also have been correctly entered according to the recommendations in the User Manuals of this kit and the RT²Profiler PCR Array. Also, double check the quality of your RNA as described in "Evidence of Poor Reverse Transcription Efficiency" above.

B. Frequently Asked Questions:

1. Shall I use C_t values that are equal to or greater than 35?

Any C_t values $\geq$35 is considered a negative call. Due to the inclusion of the pre-amplification step, for the assays showing C_t values greater than 35, the expression level of these genes will be too low to be reliably quantified. Consider removing these data points from the rest of the results.

2. What is the range of RNA amount that I can use with this kit?

This kit can be used with RNA input as little as 1ng and as much as 100ng of total RNA. For successful results and maximum positive call rates, we recommend that first-time users try starting with anywhere from 10ng to 50ng of total RNA. For best results, use a consistent amount of total RNA for all samples in a single experiment to be characterized and compared.

3. The concentration of my RNA samples is too low to be quantified by UV spectrophotometry. How do I decide on how much RNA to use for RT^2 Nano PreAMP cDNA Synthesis procedures?

When quantifying total RNA isolated from very small samples where using UV spectrophotometry is not possible, try estimating the total RNA amount by assuming that each eukaryotic cell contains an average of ~10 pg of total RNA or less. However, please note that the yield of RNA varies widely from cell type to cell type.

4. Can I use the resulting amplified product from this kit for microarrays and GEArrays?

No. The templates amplified from this kit are intended for analysis on individual RT^{2}Profiler PCR Arrays only.

5. Can I use the resulting products amplified with one Nano PreAMP cDNA Synthesis Primer Mix for all PCR Arrays?

No. Each RT^2 Nano PreAMP cDNA Synthesis Primer Mix is specific to a catalogued RT^{2}Profiler PCR Array and can only amplify cDNA templates for the intended PCR Array**.** To analyze on another PCR Array, a separate Nano PreAMP PCR reaction has to be performed using the correct RT^2 Nano PreAMP cDNA synthesis primer mix specific to the latter PCR Array.

References:

1. Katie Kinnecom and Joel S Pacther (2005) Selective capture of endothelial and perivascular cells from brain microvessels using laser capture microdissection. Brain Research Protocols. 16, 1-9.

If you have additional questions, please check our website (www.SABiosciences.com**) for a more complete listing of Frequently Asked Questions (FAQs), or call our Technical Support Representatives at 1-888-503-3187 or 301-682-9200.**

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