RayBio® Mouse Angiogenesis Antibody Array 1 (G-Series)

Patent Pending Technology

User Manual (Revised June 3, 2014)

RayBio® Mouse Angiogenesis Antibody Array G-Series 1
Cat# AAM-ANG-G1-4

RayBio® Mouse Angiogenesis Antibody Array G-Series 1
Cat# AAM-ANG-G1-8

RayBio® Mouse Angiogenesis Antibody Array
G-Series Testing Service
Cat# AAM-SERV-G

Please read manual carefully
before starting experiment

RayBiotech, Inc.
the protein array pioneer company

We provide you with excellent Protein Array systems and services

Tel: (Toll Free) 1-888-494-8555 or +1-770-729-2992; Fax: +1-770-206-2393;
Website: www.raybiotech.com Email: info@raybiotech.com

RayBiotech, Inc., the Protein Array Pioneer Company, strives to research and develop new products to meet demands of the biomedical community. RayBiotech's patent-pending technology allows detection of up to 1,000 cytokines, chemokines and other proteins in a single experiment. Our format is simple, sensitive, reliable, reproducible and cost-effective.

Our product offerings include:

1. Protein (antigen) Arrays
2. RayBio® Cytokine Antibody Arrays
   - C Series (Membrane, chemiluminescence detection)
   - G-Series (Glass chip, fluorescence detection)
3. Pathway- and disease-focused antibody arrays
   - Angiogenesis Antibody Arrays
   - Apoptosis Antibody Arrays
   - Atherosclerosis Antibody Arrays
   - Chemokine Antibody Arrays
   - Growth Factor Antibody Arrays
   - Inflammation Antibody Arrays
   - MMP Antibody Arrays
   - Obesity Antibody Arrays
4. Quantibody® Multiplex ELISA Arrays
5. RayBio® L-Series Biotin Label-based Antibody Arrays
6. RayBio® E-Series Competition-based Antibody Arrays
7. RayBio® Phosphorylation Antibody Arrays
   - Receptor Tyrosine Kinases
   - EGFR and ErbB family (site-specific phosphorylation)
8. Over 1,300 different ELISA kits
9. EIA (Competitive ELISA) kits
10. Cell-based Phosphorylation Assay
11. Over 20,000 different antibodies
12. Recombinant proteins
13. Peptide
14. Recombinant antibodies

Protocol for RayBio® Mouse Angiogenesis Antibody Array G-Series 1

TABLE OF CONTENTS

I. Introduction

New techniques such as cDNA microarrays have enabled us to analyze global gene expression[1-3]. However, almost all cell functions are executed by proteins, which cannot be studied simply through DNA and RNA techniques. Experimental analysis clearly shows disparity can exist between the relative expression levels of mRNA and their corresponding proteins[4]. Therefore, analysis of the proteomic profile is critical.

The conventional approach to analyzing multiple protein expression levels has been to use 2-D SDS-PAGE coupled with mass spectrometry[5,6]. However, these methods are slow, expensive, labor-intensive and require specialized equipment[7]. Thus, effective study of multiple protein expression levels can be complicated, costly and time-consuming. Moreover, these traditional methods of proteomics are not sensitive enough to detect most cytokines (typically at pg/ml concentrations).

Cytokines, broadly defined as secreted cell-cell signaling proteins distinct from classic hormones or neurotransmitters, play important roles in inflammation, innate immunity, apoptosis, angiogenesis, cell growth and differentiation[7]. They are involved in most diseases, including cancer, obesity and inflammatory and cardiac diseases.

Simultaneous detection of multiple cytokines undoubtedly provides a powerful tool to study cytokines. Regulation of cellular processes by cytokines is a complex, dynamic process, often involving multiple proteins. Positive and negative feedback loops, pleiotropic effects and redundant functions, spatial and temporal expression of or synergistic interactions between multiple cytokines, even regulation via release of soluble forms of membrane-bound receptors, all are common mechanisms modulating the effects of cytokine signaling[8-14]. As such, unraveling the role of individual cytokines in physiologic or pathologic processes generally requires consideration and detection of multiple cytokines rather than of a single cytokine.

RayBio® G-Series Cytokine Antibody Arrays have several advantages over detection of cytokines using single-target ELISA:

1. More Data, Less Sample: Antibody arrays provide high-content screening using about the same sample volume as for ELISA.
2. Global View of Cytokine Expression: Antibody array screening improves the chances for discovering key factors, disease mechanisms or biomarkers related to cytokine signaling.
3. Greater Sensitivity: As little as 4 pg/ml of MCP-1 can be detected using the G-Series array format. In contrast, our similar MCP-1 ELISA assay has a sensitivity of 40 pg/ml of MCP-1.
4. Increased Range of Detection: ELISA assays typically detect a concentration range of 100- to 1000-fold, however, RayBiotech arrays can detect IL-2 at concentrations of 25 to 250,000 pg/ml, a range of 10,000-fold.
5. Better Precision: As determined by densitometry, the inter-array Coefficient of Variation (CV) of spot signal intensities is 5-10%, comparing favorably with ELISA testing (CV = 10-15%).

The RayBio® G-Series Cytokine Antibody Array is a glass chip that is a highly sensitive approach to simultaneously detect multiple cytokine expression levels from diverse sample types. The experimental procedure is simple and can be performed in any laboratory. The signals from G-Series arrays are detected using a laser scanner.

Larger, multi-array G-Series Cytokine Antibody Array Kits can detect hundreds of cytokines in a single experiment. For example, the Mouse G1000 arrays can detect up to 96 cytokines, and the Mouse G2000 can detect up to 144 cytokines.

RayBiotech, The Protein Array Pioneer Company, introduced the first protein arrays to the market in 2001 and continues to lead in the development of innovative protein array technologies. For a list of publications demonstrating the usefulness of this easy-to-use array format, see Section VIII.

1. Tang X, Marciano DL, Leeman SE, Amar S. LPS induces the interaction of a transcription factor, LPS-induced TNF-a factor, and STAT6(B) with effects on multiple cytokines. *PNAS*. 2005;102(14): 5132-5137.
2. Xu Y, Kulkosky J, Acheampong E, et al.. HIV-1-mediated apoptosis of neuronal cells: Proximal molecular mechanisms of HIV-1-induced encephalopathy. *PNAS*. 2004;101(18): 7070-7075.
3. El-Hage N, Gurwell JA, Singh IN, et. al. Synergistic increases in intracellular Ca(2+), and the release of MCP-1, RANTES, and IL-6 by astrocytes treated with opiates and HIV-1 Tat. *Glia*. 2005;50(2): 91-106.
4. Oh HS, Moharita A, Potian JG, et al. Bone Marrow Stroma Influences Transforming Growth Factor-β Production in Breast Cancer Cells to Regulate c-myc Activation of the Preprotachykinin-I Gene in Breast Cancer Cells. *Cancer Res*. 2004;64: 6327-6336.
5. Osorio Y, Ghiasi H. Recombinant Herpes Simplex Virus Type 1 (HSV-1) Codelivering Interleukin-12p35 as a Molecular Adjuvant Enhances the Protective Immune Response against Ocular HSV-1 Challenge. *J Virol.* 2005;79(6): 3297-3308.
6. Mullick A, Elias M, Picard S, Bourget L, et al. Dysregulated Inflammatory Response to *Candida albicans* in a C5-Deficient Mouse Strain. *Infect Immunity*. 2004;72(10): 5868-5876.
7. Maruvada P, Wang W, Wagner PD, Srivastava S. Biomarkers in molecular medicine: cancer detection and diagnosis. *Biotechniques*. 2005;38: S9-S15.
8. Bartek J, Hodny Z, Lukas J. Cytokine loops driving senescence. *Nature Cell Biol*. 2008;10: 887-889.
9. Minami K, Yanagawa Y, Iwabuchi K, et al. Negative feedback regulation of T helper type 1 (Th1)/Th2 cytokine balance via dendritic cell and natural killer T cell interactions. *Blood*. 2005;106: 1685-1693.
10. Ozaki K, Leonard WJ. Cytokine and Cytokine Receptor Pleiotrophy and Redundancy. *J Biol Chem*. 2002;227: 29355-29358.
11. Ragab AA, Nalepka JL, Bi Y, Greenfield EM. Cytokines synergistically induce osteoclast differentiation: support by immortalized or normal calvarial cells. *Am J Physiol Cell Physiol*. 2002;283(3): C679-C687.
12. Devalaraja MN, Richmond A. Multiple chemotactic factors: fine control or redundancy. *Trends Pharmacol Sci*. 1999;20(4): 151-156.
13. Heaney ML, Golde DE. Soluble Cytokine Receptors. *Blood*. 1996;87(3): 847-857.

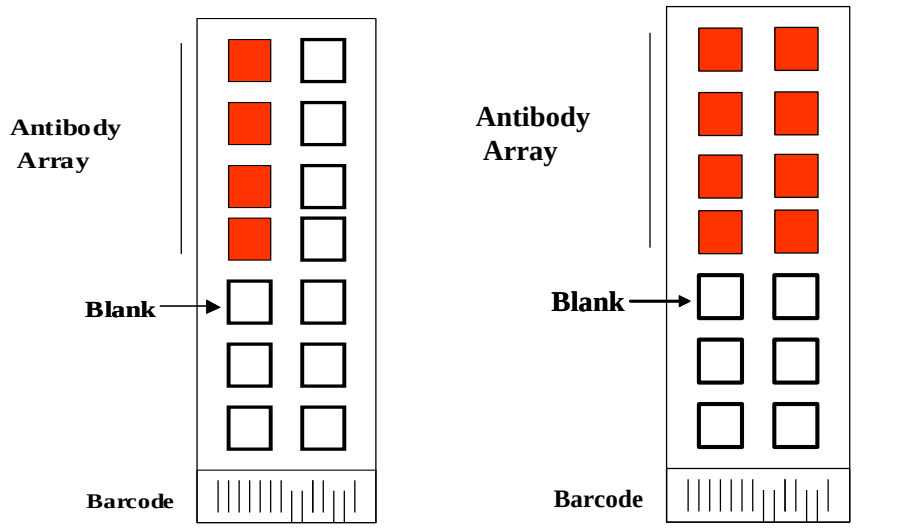
II. Product Information

A. Storage Recommendations:

For best results, we recommend storing the entire kit at -20°C or -80°C upon arrival and using the kit within 6 months of receipt. RayBiotech warranties this product for 6 months if stored in this manner.

Once thawed, store glass chips and 1X Blocking Buffer at -20°C or -80°C and all other component at 4°C. After thawing, the entire kit should be used within 3 months. RayBio® Antibody Array kits are robust and will retain full activity even if accidentally stored at room temperature (RT) for up to 24 hours.

B. RayBio® G-Series Glass Chip Layout



4 arrays in one glass chip **8 arrays in one glass chip**

C. Materials Provided

Item	Description	AAM-ANG-G1-4	AAM-ANG-G1-8
AAM-ANG-G1	RayBio® Mouse Angiogenesis G1 Microarray Glass Chip*	1 chip with 4 Sub-arrays*	1 chip with 8 Sub-arrays*
0103002-ANG-MG1	Biotin-Conjugated Anti-Cytokines	1 ea	2 ea
0103004-H	1,500X HiLyte Plus™ 555 Streptavidin-Fluor†	1 ea	1 ea
0103004-B	1X Blocking Buffer	10 ml	20 ml
0103004-W‡	20X Wash Buffer I ‡	30 ml	30 ml
0103004-W‡	20X Wash Buffer II ‡	30 ml	30 ml
0103004-L	2X Cell Lysis Buffer (optional)	10 ml	10 ml
Other Kit Components: Manual, Adhesive Plastic Strips, 30 ml Centrifuge Tube			

* Kit contains 1 pre-assembled glass chip with either 4 or 8 printed sub-arrays per chip (in sealed plastic envelope)
[*NOTE: In some cases, 2 chips x 4 sub-arrays/chip may be substituted in kits containing 8 sub-arrays]*

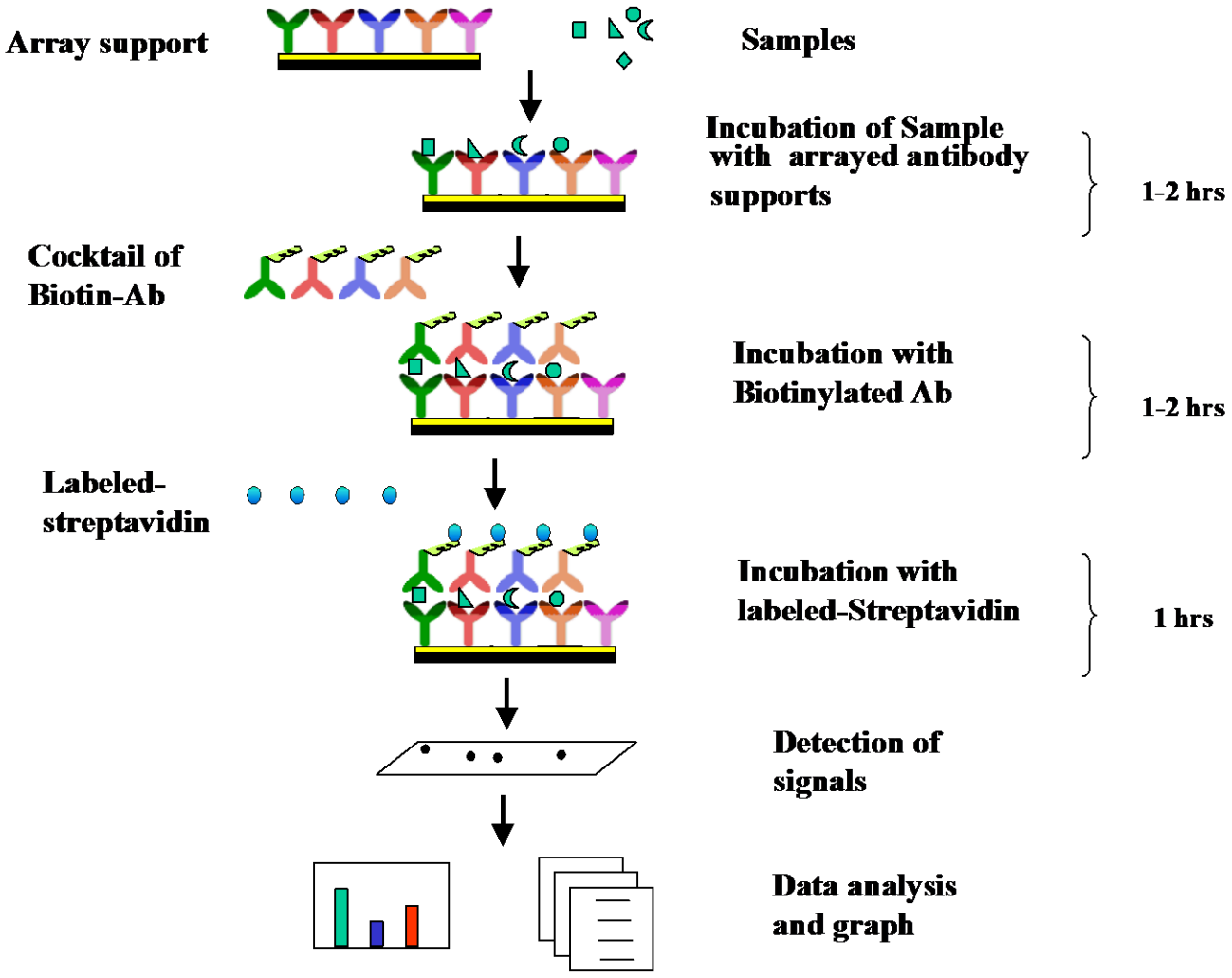
† This fluor is patent-pending technology from Anaspec, Inc.

‡ Wash Buffers and Detection Buffers are sold as sets

D. Additional Materials Required

- Small plastic boxes or containers
- Pipettors, pipette tips and other common lab consumables
- Orbital shaker or oscillating rocker
- Aluminum foil
- Gene microarray scanner or similar laser fluorescence scanner (see pages 9 & 15)

E. How It Works



III. Helpful Tips and General Considerations

A. Preparation and Storage of Samples

1. General Considerations:

- Freeze samples as soon as possible after collection.
- Avoid multiple freeze-thaw cycles. If possible, sub-aliquot your samples prior to initial storage.
- Spin samples hard (5-10 minutes at 10K to 15K RPM) immediately prior to incubation of samples with array.
- Optimal sample concentrations may need to be determined empirically based on the signal intensities of spots and background signals obtained.

- Most samples will not need to be concentrated. If concentration is required, we recommend using a spin-column concentrator with a chilled centrifuge.

2. Recommended Sample Volumes and Dilution Factors

NOTE: All sample dilutions should be made using 1X Blocking Buffer. Final sample volume = 50-100 µl per sub-array

- **Cell Cultured Media**: Neat (no dilution needed)
- **Serum & Plasma**: 5-fold to 10-fold dilution
- **Most other Body Fluids**: Neat or 2-fold to 5-fold dilution
- **Cell and Tissue Lysates**: Minimum 5-fold to 10 fold to equal concentrations of total protein in each lysate sample.
  - You must determine the total protein concentration of each lysate/homogenate. We recommend using the BCA method (available from Pierce); it is insensitive to detergents commonly found in lysis buffers.
  - Minimum Recommended Dilution of Lysates (prior to sample incubation): 5-fold to 10 fold with 1X Blocking Buffer. Dilute all lysate samples to the same final concentration of total lysate protein in 1X Blocking Buffer to 100 µl final volume.
  - To start, we recommend using 10-100 µg of total protein in 100 µl of 1X Blocking Buffer (final volume) per sub-array.
  - Optimal amounts of total lysate protein may range from 5-500 µg per sub-array. Based upon background and spots intensities, you may increase or decrease the amount of protein used in subsequent experiments.
- **Other Liquid Sample Types**: Most often Neat or 2-fold to 5-fold. However, optimal dilutions should be determined empirically.

3. Sample Preparation

For tips on sample preparation, please visit our Website:
http://www.raybiotech.com/Tech-Support/SampleTips.pdf

B. Handling Glass Chips

- Do not remove glass chip from assembly until Step 16.
- Hold the slides by edges only; do not touch the surface.
- Handle all buffers and slides with powder-free gloves.
- Dry glass chip completely before proceeding to Step 3.
- Handle and dry glass chip in clean environment.
- Avoid breaking glass chip when removing the chamber assembly.

C. Incubations and Washes

- Cover incubation chamber with adhesive film (included in kit) to prevent evaporation, particularly during incubation or wash steps >2 h or with liquid volumes <100 µl per well.
- Perform all incubation and wash steps under gentle rotation or rocking motion (~0.5 to 1 cycle/s).
- Wash steps in Wash Buffer II and all incubation steps may be performed overnight at 4°C.
  - Overnight sample incubations are the most effective at increasing sample spot intensities.
- Avoid cross-contamination of samples to neighboring wells
- To remove Wash Buffers and other reagents from chamber wells, you may invert the Glass Chip Assembly to decant, and aspirate the remaining liquid.
- In Wash Steps 6, 12 and 15, you may gently flush wells several times using a wash bottle filled with Wash Buffer I.

D. Scanning and Data Extraction Tips:

For tips on scanning and data extraction, please visit our Website: http://www.raybiotech.com/Tech-Support/ScanningTips.pdf

For a list of recommended scanners, please visit our Website: http://www.raybiotech.com/files/Tech-Support/Laser_Scanners_for_Glass_Slide_Arrays.pdf

See also page 18 of this manual.

IV. Protocol

A. Preparation and Storage of Reagents

NOTE: During this protocol, prepare reagents immediately prior to use and keep working dilutions of all reagents on ice at all times.

1. Blocking Buffer is supplied at 1X concentration. No dilution is required.

2. Wash Buffers I and II are supplied at 20X concentration.
   a). For each glass chip (4 or 8 sub-arrays/chip), dilute 6 ml of 20X concentrate with deionized H_20 to a final volume of 120 ml each of Wash Buffer I & Wash Buffer II.
   b). Wash buffer reagents at working dilution (1X) can be stored at 4°C for up to 1 month. Stock solutions at 20X can be stored 4°C for up to 3 months.

3. Biotin-conjugated Anti-Cytokines are supplied at high concentration in a small liquid bead (typically ~2-5 µl).
   a). Spin down the tube prior to reconstitution, as the concentrated liquid bead may have moved to the top of the tube during handling.
   b). Prepare stock reagent by adding 300 µl 1X Blocking Buffer to Biotin-Conjugated Anti-Cytokines. Mix well.
   c). 1X Biotin-Conjugated Anti-Cytokines may be stored for 2-3 days at 4°C.

4. Streptavidin-Fluor is supplied at 1500x concentration.
   a). Mix the tube containing 1500X Streptavidin-Fluor well before use, as precipitants may form during storage.
   b). Add 100 µl of 1X Blocking Buffer to tube containing 1500X Streptavidin-Fluor. Mix well.
   c). Quantitatively transfer all of Streptavidin-Fluor reagent from the original tube to a larger one, and dilute with 1X Blocking Buffer to a final volume of 1500 µl (ie, 1.5 ml).
   d). Wrap tube containing Streptavidin-Fluor with aluminum foil.
   e). This working dilution can be stored for 3-5 days at 4°C.

B. Blocking and Incubations

NOTE: Please carefully read Section III of this manual before proceeding

NOTE: Prepare all reagents immediately prior to use as described above (Section IV.A) and before proceeding.

1) Remove the package containing the glass chip assembly from the freezer. Place unopened package on the benchtop and allow the glass chip assembly to equilibrate to room temperature (RT), approx. 15 min. Open package, remove the glass chip assembly and place in laminar flow hood to dry for 1-2 hours.

   NOTE: Be sure glass chip is completely dry before proceeding.

2) If necessary, assemble the glass chip into incubation chamber and frame as shown on page 12. (Note: if you slide is already assembled, you can proceed directly to Step 3).

3) Add 100 µl 1 X Blocking Buffer into each well and incubate at RT for 30 min to block slides.

   NOTE: Only add reagents or samples to wells printed with antibodies (see diagram on page 6)

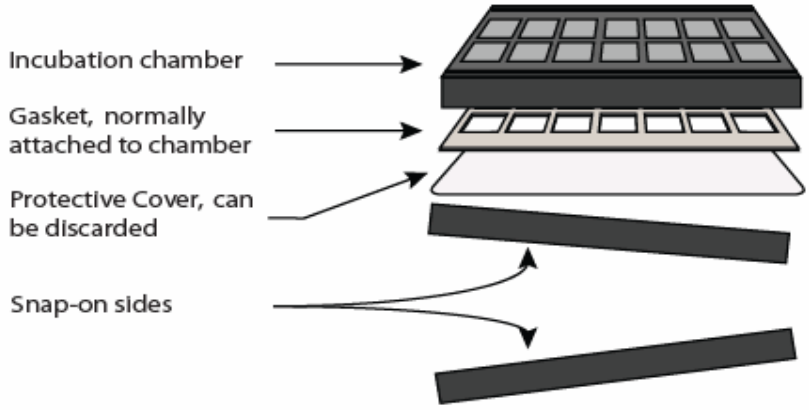
4) Decant Blocking Buffer; then aspirate remaining liquid from each well.

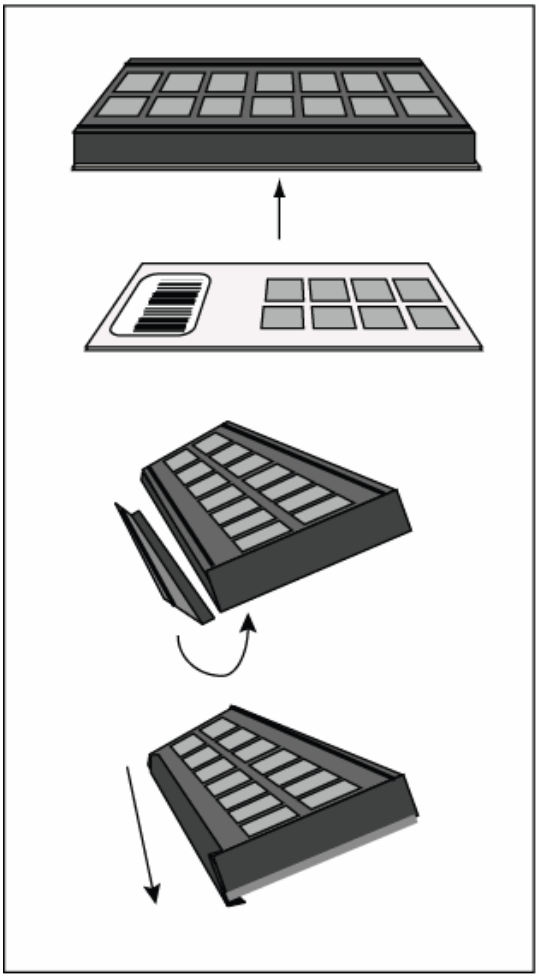
   NOTE: To aspirate liquid samples or reagents from wells, gently place the pipette tip only in the corners of the well. Do not scrape the pipette tip across the surface of the chip.

5) Add 50 to 100 µl of each sample to each sub-array. Cover the incubation chamber with Adhesive film (included in kit). Incubate arrays with sample at RT for 2 hours. Dilute sample using 1X Blocking Buffer if necessary.

Instructions for incubation chamber assembly

G Series and Quantibody Arrays





1 Carefully place slide at bottom of the chamber as shown. The slide will adhere somewhat to the bottom. Warning: the slide is fragile, so do not apply more than gentle force to the apparatus.

2 While gently holding chamber and slide, place side on chamber as shown, beginning with bottom flap first.

3 Then, press the top of the side into grove on chamber, and then apply even, gentle pressure from one end to the other. Repeat this procedure with the other side.

6) Remove adhesive film, and carefully aspirate samples from sub-arrays, touching only the corners with your pipette tip.

NOTE: Try to prevent solution from flowing into neighboring wells.

7) Wash 3 x 2 min with 150 µl 1X Wash Buffer I at RT. Be sure to completely remove sample and Wash Buffer each time and use fresh buffer for each wash. Decant final wash solution before proceeding to next step.

8) Obtain a clean container (eg, pipette tip box or slide staining jar) and place glass chip assembly into the container. Add enough 1X Wash Buffer I to submerge the entire glass chip with frame intact (approx. 30-50 ml) and remove all bubbles in wells. Wash 10 min at RT with gentle rocking or shaking.

9) Remove assembled glass chip and invert to decant liquid. Decant buffer from container and replenish with 1X Wash Buffer I. Submerge the entire glass chip assembly and wash 10 min at RT with gentle rocking or shaking.

10) Remove assembled glass chip and invert to decant liquid. Decant buffer from container and repeat Steps 8 & 9 with Wash Buffer II.

11) Remove assembled glass chip and invert to decant liquid, then carefully aspirate wash buffer from wells, touching only the corners with your pipette tip.

12) Add 70 µl of 1X Biotin-conjugated Anti-Cytokines to each sub-array. Cover incubation chamber with Adhesive film (included in kit). Incubate at RT for 2 hours with gentle rocking or shaking.

13) Carefully aspirate all of the Biotin-conjugated Anti-Cytokine reagent. Wash as described in Step 7 above, first with Wash Buffer I then with Wash Buffer II, making sure to completely remove buffer between washes and after final wash.

14) Add 70 µl of 1X Streptavidin-Fluor to each sub-array. Cover the incubation chamber with Adhesive film (included in kit), then cover entire assembly with aluminum foil to avoid exposure to light or incubate in dark room. Incubate at RT for 2 hours with gentle rocking or shaking.

15) Remove aluminum foil and adhesive film. Carefully aspirate the Streptavidin-Fluor reagent. Wash as described in Step 7 above, first with Wash Buffer I then with Wash Buffer II, making sure to completely remove buffer between washes and after final wash.

16) Remove the glass chip from the frame assembly. Place the whole chip in 30 ml centrifuge tube provided, or slide staining jar. Add enough Wash Buffer I to cover the whole slide (about 20 ml) and gently rock or shake at RT for 10 min.

17) Decant buffer and repeat wash as described in Step 16 (1 x 10 min with Wash Buffer I).

18) Decant buffer and repeat wash as described in Step 16, but this time using Wash Buffer II for only 2-3 minutes.

19) Decant buffer, remove the glass chip from the tube, then gently rinse the slide with de-ionized H_2O using a plastic wash bottle.

20) Remove water droplets by applying suction gently with a pipette tip.

NOTE: Be careful not to touch the array portions of the slide with your pipette tip, only touch the sides of the slide.

C. Obtaining Fluorescent Signal Intensities:

21) Allow glass chip to dry in a laminar flow hood for 20 minutes or until slide is completely dry. Place chip under an aluminum foil tent to protect it from light. Make sure the slides are absolutely dry before scanning or storage.

22) You may proceed immediately to scanning (Step 23), or you may store the slide at -20 °C in the centrifuge tube provided or at RT and to scan at a later time.

Note: Unlike most Cy3 fluors, the HiLyte Plus™ Fluor 555 used in this kit is very stable at RT and resistant to photobleaching on completed glass chips. However, please protect glass chips from strong light and temperatures above RT.

23) Scan the glass chip with a laser scanner (such as Innopsys' InnoScan®) using cy3 or "green" channel (excitation frequency = 532 nm). For tips on scanning, visit our Website: http://www.raybiotech.com/Tech-Support/ScanningTips.pdf

NOTE: If you do not have a laser scanner, for a nominal fee you can send your slide to us for scanning and data extraction, and we will return the results to you. Also, using alternate protocols, RayBio® G-Series arrays are compatible with Li-Cor's Odyssey and Gentel BioScience's APiX scanners. For more information, contact these vendors or RayBiotech.

V. Interpretation of Results:

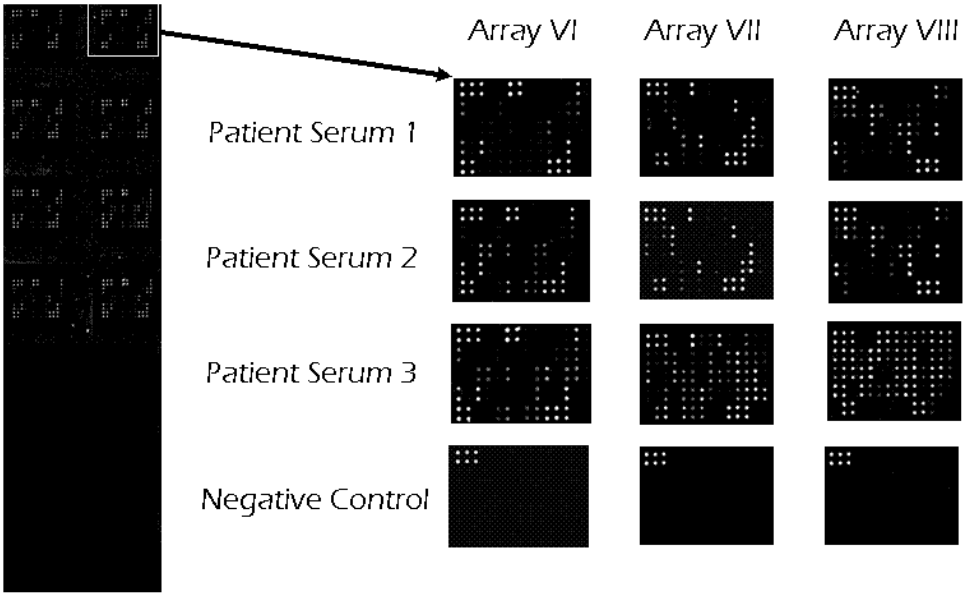
A. Explanation of Controls Spots

Positive Controls (POS1, POS2, POS3) are equal amounts of biotinylated IgGs printed directly onto the array. All other variables being equal, the Positive Control intensities will be the same for each sub-array This allows for normalization based upon the relative fluorescence signal responses to a known control, much as "housekeeping" genes or proteins are used to normalize results in PCR or Western blots, respectively.

Negative Control (NEG) spots are a protein-containing buffer (used to dilute antibodies printed on the array). Their signal intensities represent non-specific binding of Biotin-conjugated anti-Cytokines and/or Streptavidin-Fluor. Negative control signal intensities are usually very close to background signals in each sub-array.

B. Typical results from RayBio® G-Series Antibody Arrays

The following figure shows typical results obtained using RayBio® Antibody Array G-Series Arrays. The images were captured using a GenePix 4000B scanner.



In this example, sera from several patients were incubated with Human Cytokine Arrays 6, 7 & 8, (sold together as Human Cytokine Array G-Series 2000, AAH-CYT-G2000-4 or AAH-CTY-G2000-8) and processed using this standard protocol.

The 6 strong signals of the Positive Control spots in the upper-left corner are useful for proper orientation of the array image.

If scanned using optimal scan settings, 3 distinct Positive Control signal intensities will be seen: POS1>POS2>POS3. If all of these signals are of similar intensity, try increasing or decreasing laser power and/or signal gain settings.

Once you have obtained fluorescence intensity data, you should subtract the background and normalize to the Positive Control signals before proceeding to analysis.

C. Background Subtraction:

Most laser fluorescence scanner software have an option to automatically measure the local background around each spot. As with spot signal intensities, we recommend using MEDIAN background signals. If your resulting fluorescence signal intensity reports do not include these values (eg, a column labeled as "MED532-B532"), you may need to subtract the background manually or change the default settings on your scanner's data report menu.

D. Normalization of Array Data:

To normalize signal intensity data, one sub-array is defined as "reference" to which the other arrays are normalized. This choice can be arbitrary. For example, in our Analysis Tool Software, the array represented by data entered in the left-most column each worksheet is the default "reference array."

You can calculate the normalized values as follows:

$$X(Ny) = X(y) * P1/P(y)$$

Where:

P1 = mean signal intensity of POS spots on reference array
P(y) = mean signal intensity of POS spots on Array "y"
X(y) = mean signal intensity for spot "X" on Array "y"
X(Ny)= normalized signal intensity for spot "X" on Array "y"

The RayBio® Analysis Tool software is available for use with data obtained using RayBio® G-Series Arrays. You can copy and paste your signal intensity data (with and without background) into the Analysis Tool, and it will automatically normalize signal intensities to the Positive Controls.

To order the Analysis Tool, please contact us at +1-770-729-2992 or info@raybiotech.com for more information.

E. Threshold of significant difference in expression:

After subtracting background signals and normalization to Positive controls, comparison of signal intensities for antigen-specific antibody spots between and among array images can be used to determine relative differences in expression levels of each analyte (ie, protein detected) between samples or groups.

Any ≥1.5-fold increase or ≤0.65-fold decrease in signal intensity for a single analyte between samples or groups may be considered a measurable and significant difference in expression, provided that both sets of signals are well above background (Mean background + 2 standard deviations, accuracy ≈ 95%).

NOTE: In the absence of an external standard curve for each analyte, there is no means of assessing absolute or relative concentrations of different analytes in the same sample using immunoassays. If you wish to obtain quantitative data (ie, concentrations of the various analytes in your samples), try using our Quantibody® Multiplex ELISA arrays instead.

Data Extraction Tips:

- Ignore any comet tails
- Define the area for signal capture for all spots as 110-120 micron diameter, using the same area for every spot.
- Use median signal value, not the total or the mean
- Use local background correction (also median value).
- Exclude obvious outlier data in its calculations.
- Scan all slides at same PMT

VI. RayBio® Mouse Angiogenesis Antibody Array G-Series 1 Map:

Detects 24 Mouse Angiogenic Factors in one experiment

	A	B	C	D	E	F	G	H
1	POS1	POS2	POS3	NEG	NEG	CCL11	FASLG	bFGF
2	POS1	POS2	POS3	NEG	NEG	CCL11	FASLG	bFGF
3	CSF3	CSF2	IFN-γ	IGF2	IL1A	IL1B	IL12 p40/p70	IL12 p70
4	CSF3	CSF2	IFN-γ	IGF2	IL1A	IL1B	IL12 p40/p70	IL12 p70
5	IL13	IL6	IL9	LeptIn	MCP1	CSF1	MIG	PF4
6	IL13	IL6	IL9	LeptIn	MCP1	CSF1	MIG	PF4
7	TIMP1	TIMP2	TNF-α	THPO	VEGF-A	NEG	NEG	NEG
8	TIMP1	TIMP2	TNF-α	THPO	VEGF-A	NEG	NEG	NEG

Notes on Array Map:

CCL11 = Eotaxin, CSF3 = G-CSF, CSF2 = GM-CSF, THPO = Thrombopoietin

VII. Troubleshooting guide

Problem	Cause	Recommendation
No signal for any spots, including Positive Controls	Global detection failure	Adjust scanner settings or re-assemble chip into holder, wash slide 2 x 5 min with 150 µl Wash Buffer II and repeat Steps 12-19.
Similar signal intensities for POS1/2/3	Improper laser power and/or PMT setting	Repeat scan using higher and/or lower laser power or PMT settings
High background signals	Incomplete washes	Carefully follow wash protocols, and/or increase wash times
	Sample concentration is too high	Repeat using lower sample concentration
	Fluor and/or Anti-Cytokines are too concentrated	Review protocol for dilution of reagents
Uneven background and/or missing spots	Bubbles present on chip during incubations	Be sure to completely remove all bubbles from chip surface
	Evaporation during incubation steps	Cover chamber assembly during washes and incubations
	Pooling/precipitation of sample or reagent; Incomplete washes.	Cover chamber assembly and use a rocker or shaker during washes and incubations; carefully follow wash protocols.
	Sample is too concentrated	Repeat experiment using more dilute sample
Randomly scattered high-intensity spots	Dust or other particulates	Dry slides in laminar flow hood and/or use clean containers and powder-free gloves.

Weak or no signals antigen-specific pots + Low Background	Sample is too dilute	Repeat experiment using higher sample concentration
	Improper dilution of Anti-Cytokines or Streptavidin-Fluor	Re-assemble chip into holder, wash 2 x 5 min with 150 µl Wash Buffer II and repeat Steps 12-19. Spin down reagents before diluting and mix well.
	Other Tips	Rescan at higher laser power or signal gain setting
		Repeat using higher sample concentration and/or incubate wi sample O/N at 4°C
		Increase concentration of and/or length of incubation with Biotin-conjugated Anti-Cytokine (+ add'l large volume wash following Biotin-Ab incubation
		Review proper storage conditions for kit components

III. Selected References Citing RayBio® Mouse G-Series Arrays

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Customized RayBio® Cytokine Antibody Arrays. Select your cytokines of interest from the following list, and we will produce the customized array for you. For more information, please visit our website, www.raybiotech.com.

4-1BB
6Ckine
ACE
Activin A
ALK-1
Amphiregulin
ANGPTL3
Axl
B7-1 / CD80
BAFF R
basic FGF
BCMA
beta IG-H3
BLC
Betacellulin
Cardiotrophin-1
CCL28
CCL6
CD27
CD27 Ligand
CD30
CD30 Ligand
CD36
CD40
CD40 Ligand
CD48
CD6
Chemerin
Chordin
Clusterin
C5a
CRG-2
CTACK
CTLA-4
CXCL16
Cystatin C
DAN / NBL1
Decorin
Delta-like 4
Dkk-1
DPPIV
Dtk
E-Cadherin
EDAR
EGF
Endocan
Endoglin
Eotaxin
Eotaxin-2
Epigen
Epiregulin
E-Selectin
Fas Ligand
Fc gamma RIIA
Fc gamma RIIB
Fetuin A
Flt-3 Ligand
Fractalkine
Galectin-1
Galectin-3
Galectin-7
Gas 1
Gas 6
G-CSF / CSF3
GITR
GITR Ligand
GM-CSF / CSF2
gp130
Granzyme B
Gremlin-2
H60
HAI-1
HGF
HGF R
ICAM-1
IFN-gamma
IFN-gamma R1
IGFBP-2
IGFBP-3
IGFBP-5
IGFBP-6
IGF-I
IGF-II
IL-1 alpha
IL-1 beta
IL-1 R4 / ST2
IL-10
IL-11
IL-12 p40
IL-12 p70
IL-13
IL-15
IL-17
IL-17B
IL-17B R
IL-17E
IL-17F
IL-1ra
IL-2
IL-2 R alpha
IL-20
IL-21
IL-22
IL-23p19
IL-28A
IL-3
IL-3 R beta
IL-33
IL-4
IL-5
IL-6
IL-6 R
IL-7
IL-7 R alpha
IL-9
I-TAC
JAM-A
KC / CXCL1
Kremen-1
Leptin (OB)
Leptin R
Limitin
NGAL
LIX
LOX-1
L-Selectin
Lungkine
Lungkine
Lymphotactin
MAdCAM-1
MBL2
Marapsin
MCP-1 / CCL2
MCP-5 / CCL12
M-CSF / CSF1
MDC / CCL22
Meteorin
MFG-E8
MIG / CCL9
MIP-1α / CCL3
MIP-1β / CCL4
MIP-1γ / CCL9
MIP-2 / CXCL2
MIP-3α / CCL20
MIP-3β / CCL19
MMP-2
MMP-3
Neprilysin
NOPE
NOV / CCN3
Osteoactivin
Osteopontin
Osteoprotegerin
OX40 Ligand
P-Cadherin
Pentraxin 3
Periostin
PF-4 / CXCL4
PLGF-2
Progranulin
Prolactin
Pro-MMP-9
Prostasin
P-Selectin
RAGE / AGER
RANTES
Renin
Resistin
SCF / Kit Ligand
SDF-1α
Sonic Hedgehog
TACI
TARC / CCL17
TCA-3 / CCL1
TECK / CCL25
Testican 3
TGF beta 1
Thrombopoietin
Thymus CK-1
KIM-1 / TIM-1
TIMP-2
TNF alpha
sTNFRI
sTNFRI
TRAIL
TRANCE
TREM-1
TROY
Tryptase ε
TSLP
TWEAK
TWEAK R
VCAM-1
VEGF-A
VEGF R1 / FLT1
VEGF R2 / KDR
VEGF R3 / FLT4
VEGF-D

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