RayBio[®] Human/Mouse/Rat Visfatin Enzyme Immunoassay Kit

Catalog #: EIA-VIS, EIAM-VIS, EIAR-VIS

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Caution: Extraordinarily useful information enclosed



ISO 13485 Certified

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Please read the entire manual carefully before starting your experiment

I. Introduction

Obesity, which is characterized by excessive accumulation of adipose tissue in the body, has become one of the greatest public health challenges. Obesity is not only associated with health problems linked to increased weight-dependent pressure overload on lung, joints and bones, but also a important risk factor for life-threatening diseases such as cardiovascular diseases, type 2 diabetes and certain cancers.

Visfatin, an adipokine that is highly enriched in the visceral fat, has increased expression level in plasma during the development of obesity, implying its important role in obesity.

Visfatin (also known as PBEF and Nampt), a 52 kDa cytoplasmic protein composed of 473-amino acids, is expressed primarily in bone marrow, muscle and liver tissue but can also be detected in placenta, lung, kidney and heart tissue.

In addition to its role in obesity, visfatin also has other biological functions. Visfatin is a nicotinamide phosphoribosyltransferase (Nampt) enzyme that catalyzes first step in the biosynthesis of NAD from nicotinamide. Visfatin has also been reported to be a cytokine (PBEF) that promotes B cell maturation and inhibits neutrophil apoptosis.

II. General Description

The RayBio[®] Visfatin Enzyme Immunoassay (EIA) Kit is an in vitro quantitative assay for detecting Visfatin peptide based on the competitive enzyme immunoassay principle.

In this assay, a biotinylated Visfatin peptide is spiked into the samples and standards. The samples and standards are then added to the plate, where the biotinylated Visfatin peptide competes with endogenous (unlabeled) Visfatin for binding to the anti-Visfatin antibody. After a wash step, any bound biotinylated Visfatin then interacts with horseradish peroxidase (HRP)-streptavidin, which catalyzes a color development reaction. The intensity of the colorimetric signal is directly proportional to the amount of captured biotinylated Visfatin peptide and inversely proportional to the amount of endogenous Visfatin in the standard or samples. A standard curve of known concentration of Visfatin peptide can be established and the concentration of Visfatin peptide in the samples can be calculated accordingly.

III. How It Works



IV. Storage

The entire kit may be stored at -20°C to -80°C for up to 6 months from the date of shipment. For extended storage, it is recommended to store at -80°C. **Avoid repeated freeze-thaw cycles.** For prepared reagent storage, see table below.

V. Reagents

Component	Size / Description	Storage / Stability After Preparation	
Visfatin Microplate (Item A)	96 wells (12 strips x 8 wells) coated with secondary antibody.	1 month at 4°C*	
Wash Buffer Concentrate (20X) (Item B)	25 ml of 20X concentrated solution.	1 month at 4°C	
Standard Visfatin Peptide (Item C)	2 vials of Lyophilized Visfatin Peptide. 1 vial is enough to run each standard in duplicate.	Do not store and reuse	
Anti-Visfatin Polyclonal Antibody (Item N)	2 vials of Lyophilized anti-Visfatin.	Do not store and reuse	
5X Assay Diluent B (Item E)	15 ml of 5X concentrated buffer. Diluent for both standards and samples including serum, plasma, cell culture media or other sample types.	1 month at 4°C	
Biotinylated Visfatin Peptide (Item F)	2 vials of Lyophilized Biotinylated Visfatin Peptide, 1 vial is enough to assay the whole plate.	Do not store and reuse	
HRP-Streptavidin Concentrate (Item G)	600 μl 100X concentrated HRP-conjugated streptavidin.	Do not store and reuse	
Positive Control (Item M)	1 vial of Lyophilized Positive Control.	Do not store and reuse	
TMB One-Step Substrate Reagent (Item H)	12 ml of 3,3,5,5'-tetramethylbenzidine (TMB) in buffer solution.	N/A	
Stop Solution (Item I)	8 ml of 0.2 M sulfuric acid.	N/A	

*Return unused wells to the pouch containing desiccant pack, reseal along entire edge.

VI. Additional Materials Required

- 1. Microplate reader capable of measuring absorbance at 450 nm
- 2. Precision pipettes to deliver 2 µl to 1 ml volumes
- 3. Adjustable 1-25 ml pipettes for reagent preparation
- 4. 100 ml and 1 liter graduated cylinders
- 5. Absorbent paper
- 6. Distilled or deionized water
- 7. SigmaPlot software (or other software which can perform four-parameter logistic regression models)
- 8. Tubes to prepare standard or sample dilutions
- 9. Orbital shaker
- 10. Aluminum foil
- 11. Plastic wrap

VII. Reagent Preparation

Keep kit reagents on ice during reagent preparation steps.

A. Preparation of Plate and Anti-Visfatin Antibody

- 1. Equilibrate plate to room temperature before opening the sealed pouch.
- 2. Label removable 8-well strips as appropriate for your experiment.
- 3. 5X Assay Diluent B (Item E) should be diluted 5-fold with deionized or distilled water.
- Briefly centrifuge the anti-Visfatin antibody vial (Item N) and reconsititute with 55 µl of 1X Assay Diluent B to prepare the antibody concentrate. Pipette up and down to mix gently.
- 5. The antibody concentrate should then be diluted 100-fold with 1X Assay Diluent B. This is your anti-Visfatin antibody working solution, which will be used in step 2 of Assay Procedure (Section VIII).

Note: The following steps may be done during the antibody incubation procedure (step 2 of Assay Procedure)

B. Preparation of Biotinylated Visfatin (Item F)

- 5. Briefly centrifuge the vial of Biotinylated Visfatin (Item F) and reconstitute with 20 μ I of ddH₂O before use.
- See the image below for proper preparation of Item F. Transfer the entire contents of the Item F vial into a tube containing 10 ml of 1X Assay Diluent B. This is your Working Stock of Item F. Pipette up and down to mix gently. *The final concentration of biotinylated Visfatin will be* **20 ng/ml**.
 - a. Second Dilution of Item F for Standards: Add 2 ml of Working Stock Item F to 2 ml of 1X Assay Diluent B. The final concentration of biotinylated Visfatin will be 10 ng/ml.
 - b. Second Dilution of Item F for Positive Control: Add 100 µl of Working Stock Item F to 100 µl of the prepared Positive Control (Item M). (See section D for Positive Control preparation) The final concentration of biotinylated Visfatin will be **10 ng/ml**.
 - c. Second Dilution of Item F for samples: Add 125 µl of Working Stock Item F to 125 µl of prepared sample (see section E for sample preparation). This is a 2-fold dilution of your sample. The final concentration of biotinylated Visfatin will be **10 ng/ml**.



C. Preparation of Standards

 Label 6 microtubes with the following concentrations: 1000 ng/ml, 100 ng/ml, 10ng/ml, 1 ng/ml, 100 pg/ml and 0 pg/ml. Pipette 450 µl of biotinylated Visfatin Item F working solution (prepared in step 6a) into each tube, except the 1,000 ng/ml (leave this one empty).

It is very important to make sure the concentration of biotinylated Visfatin is 10 ng/ml in all standards.

- 8. Briefly centrifuge the vial of Visfatin Standard (Item C). Reconstitute with 10 μl of ddH₂O and briefly vortex if desired. Pipette 8 μl of Item C and 792 μl of 10 ng/ml biotinylated Visfatin working solution (prepared in step 6a) into the tube labeled 1000 ng/ml. Mix thoroughly. This solution serves as the first standard (1,000 ng/ml Visfatin standard, 10 ng/ml biotinylated Visfatin).
- 9. To make the 100 ng/ml standard, pipette 50 µl of the 1000 ng/ml Visfatin standard into the tube labeled 100 ng/ml. Mix thoroughly.
- 10. Repeat this step with each successive concentration, preparing a dilution series as shown in the illustration below. Each time, use 450 µl of biotinylated Visfatin and 50 µl of the prior concentration until the 100 pg/ml is reached. Mix each tube thoroughly before the next transfer.



D. Positive Control Preparation

- 11. Briefly centrifuge the Positive Control vial (Item M) and reconstitute with 100 μ l of ddH₂O.
- 12. Refer to step 6b. This is a 2-fold dilution of the Positive Control. The final concentration of biotinylated Visfatin should still be 10 ng/ml.

The Positive Control is a cell culture media sample that serves as a system control to verify that the kit components are working. The resulting OD will not be used in any calculations; if no positive competition is observed please contact RayBiotech Technical Support. The Positive Control may be diluted further if desired, but be sure the final concentration of biotinylated Visfatin is 10 ng/ml.

E. Sample Preparation

 If you wish to perform a 2-fold dilution of your sample, proceed to step 6c. If you wish to perform a higher dilution of your sample, dilute your sample with 1X Assay Diluent B before performing step 6c.

EXAMPLE (to make a 4-fold dilution of sample):

- a. Dilute sample 2-fold (62.5 µl of sample + 62.5 µl of 1X Assay Diluent B.).
- b. Perform step 6c (125 μl of working solution Item F + 125 μl of sample prepared above).

The total volume is 250 µl, enough for duplicate wells on the microplate.

It is very important to make sure the final concentration of the biotinylated Visfatin is **10 ng/ml**.

Note: Optimal sample dilution factors should be determined empirically, however you may reference below for recommended dilution factors for serum: Human=4X Mouse=4X Rat=2X.

If you have any questions regarding the recommendended dilutions you may contact technical support at 888-494-8555 or techsupport@raybiotech.com.

F. Preparation of Wash Buffer and HRP

- 14. If Item B (20X Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved.
- 15. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.
- 16. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use.
- 17. Dilute the HRP-Streptavidin concentrate 100-fold with 1X Assay Diluent B.

VIII. Assay Procedure

- 1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
- Add 100 µl of Anti-Visfatin Antibody (Item N) (See Reagent Preparation step 3) to each well. Incubate for 1.5 hours at room temperature with gentle shaking (1-2 cycle/sec). You may also incubate overnight at 4°C.
- 3. Discard the solution and wash wells 4 times with 1X Wash Solution Buffer (200-300 µl each). Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
- 4. Add 100 µl of each standard (see Reagent Preparation Section C), Positive Control (see Reagent Preparation Section D) and sample (see Reagent Preparation Section E) in appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1-2 cycles/sec) overnight or at 4°C.
- 5. Discard the solution and wash 4 times as directed in Step 3.

- Add 100 µl of prepared HRP-Streptavidin solution (see Reagent Preparation step 7) to each well. Incubate for 45 minutes at room temperature with gentle shaking. It is recommended that incubation time should not be shorter or longer than 45 minutes.
- 7. Discard the solution and wash 4 times as directed in Step 3.
- Add 100 µl of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1-2 cycles/sec).
- 9. Add 50 µl of Stop Solution (Item I) to each well. Read at 450 nm immediately.

IX. Assay Procedure Summary

- 1. Prepare all reagents, samples and standards as instructed.
- Add 100 µl anti-Visfatin to each well. Incubate 1.5 hours at room temperature or overnight at 4°C.
- 3. Add 100 µl standard or sample to each well. Incubate 2.5 hours at room temperature or overnight at 4°C.
- Add 100 µl prepared Streptavidin solution. Incubate 45 minutes at room temperature.
- 5. Add 100 µl TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.
- 6. Add 50 µl Stop Solution to each well. Read at 450 nm immediately.

X. Calculation of Results

Calculate the mean absorbance for each set of duplicate stands, controls, and samples and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit curve through the standard points.

Percentage absorbance = (B-blank OD)/ B_0 -blank OD) where B = OD of sample or standard and B₀ = OD of zero standard (total binding)

A. Typical Data

These standard curves are for demonstration only. A standard curve must be run with each assay.



B. Sensitivity

The minimum detectable concentrations of Visfatin is 0.778 ng/ml or 6.82 pM.

C. Detection Range

0.1-1,000 ng/ml

D. Reproducibility

Intra-Assay: CV<10% Inter-Assay: CV<15%

E. Assay Diagram

Recommended Plate Layout:

Blank	Blank	SA1	SA1	SA9	SA9	SA17	SA17	SA25	SA25	SA33	SA33
Total Binding	Total Binding	SA2	SA2	SA10	SA10	SA18	SA18	SA26	SA26	SA34	SA34
Standard1	Standard1	SA3	SA3	SA11	SA11	SA19	SA19	SA27	SA27	SA35	SA35
Standard2	Standard2	SA4	SA4	SA12	SA12	SA20	SA20	SA28	SA28	SA36	SA36
Standard3	Standard3	SA5	SA5	SA13	SA13	SA21	SA21	SA29	SA29	SA37	SA37
Standard4	Standard4	SA6	SA6	SA14	SA14	SA22	SA22	SA30	SA30	SA38	SA38
Standard5	Standard5	SA7	SA7	SA15	SA15	SA23	SA23	SA31	SA31	SA39	SA39
Pos Control	Pos Control	SA8	SA8	SA16	SA16	SA24	SA24	SA32	SA32	SA40	SA40

Key:

Blank = Buffer Only Total Binding = Biotin- Visfatin only Standard 1 = 1000 ng/ml Standard 2 = 100 ng/ml Standard 3 = 10 ng/ml Standard 4 = 1 ng/ml Standard 5 = 100 pg/ml Pos Control = Biotin with Item M

XI. Specificity

Cross Reactivity: This EIA kit shows no cross-reactivity with any of the cytokines tested: Ghrelin, Nesfatin, NPY and APC.

XIV. Publications Citing This Product

 Elsamanoudy A., et al. Endothelial Dysfunction and Insulin Resistance as Pathophysiologic Mechanisms in a Rat Model of Preeclampsia. American Journal of Biochemistry and Biotechnology 6 (3): 172-180, 2010. DOI : 10.3844/ajbbsp.2010.172.180 Species: Rat

Sample Type: Plasma

 Abdin A., et al. Effect of propranolol on IL-10, visfatin, Hsp70, iNOS, TLR2, and survivin in amelioration of tumor progression and survival in Solid Ehrlich Carcinoma-bearing mice. Pharmacological Reports 2014, Available online 8 August 2014. DOI: 10.1016/j.pharep.2014.07.010

Species: Mouse

Sample Type: Tissue Lysate

3. Guzel S., et al. Visfatin, Leptin, and TNF-?: Interrelated Adipokines in Insulin-Resistant Clinical and Subclinical Hypothyroidism. Endocrine Research. Aug2013, Vol. 38 Issue 3, p184-194. DOI:10.3109/07435800.2012.760588

Species: Human Sample Type: Serum

 Mabrouk R., et al. Serum Visfatin, Resistin and IL-18 in A Group of Egyptian Obese Diabetic and Non Diabetic Individuals. THE EGYPTIAN JOURNAL OF IMMUNOLOGY Vol. 20 (1), 2013

Species: Human

Sample Type: Serum

 Yanni D., et al. The role of leptin, soluble leptin receptor, adiponectin and visfatin in insulin sensitivity in preterm born children in prepubertal ages. Cytokine 64 (2013) 448Ã,–453. http://dx.doi.org/10.1016/j.cyto.2013.04.034

Species: Human Sample Type: Serum

 Gumus U., et al. Plasma visfatin levels in adolescents with polycystic ovary syndrome: A prospective case-control study. Journal of Pediatric and Adolescent Gynecology. Sept 2014, epub ahead of print. DOI: 10.1016/j.jpag.2014.08.007

Species: Human

Sample Type: Plasma

For additional publications citing this product, please contact technical support at 888-494-8555 or techsupport@raybiotech.com.

XIII. Troubleshooting Guide

Problem	Cause	Solution				
Poor standard curve	 Inaccurate pipetting Improper standard dilution 	 Check pipettes Briefly centrifuge Item C and dissolve the powder thoroughly by gently mixing 				
Low signal	 Improper preparation of standard and/or biotinylated antibody Too brief incubation times Inadequate reagent volumes or improper dilution 	 Briefly spin down vials before opening. Dissolve the powder thoroughly. Ensure sufficient incubation time; assay procedure step 2 may be done overnight Check pipettes and ensure correct preparation 				
Large CV	Inaccurate pipettingAir bubbles in wells	Check pipettesRemove bubbles in wells				
High background	 Plate is insufficiently washed Contaminated wash buffer 	 Review the manual for proper wash. If using a plate washer, ensure that all ports are unobstructed. Make fresh wash buffer 				
Low sensitivity	 Improper storage of the ELISA kit Stop solution 	 Follow storage recomendations in sections IV and V. Keep substrate solution protected from light. Add stop solution to each well before reading plate 				

Over 2,000 ELISA kits available, visit www.RayBiotech.com/ELISA-Kits.html for details.

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