

# Catchment Manual

*Tie Shan Ping*  
(TSP) monitoring  
site

Chongqing municipality

## **IMPACTS project**

Integrated Monitoring Program on Acidification of  
Chinese Terrestrial Systems

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## 1. INTRODUCTION

One important aspect of intensive monitoring is to follow the changes in water chemistry as it passes through the various compartments of the watershed. In order to achieve this the water must be sampled on a routine basis from a set of stations within the catchment.

At the Tie Shan Ping (TSP) monitoring site the following sampling equipment are installed:

- 1 weather station for recording the main meteorological parameters
- 1 bulk precipitation collector and 1 wet only precipitation collector
- 1 three-stage filterpack system for sampling main air components (i.e. SO<sub>2</sub>, NO<sub>3</sub><sup>-</sup> + HNO<sub>3</sub>, NH<sub>4</sub><sup>+</sup> + NH<sub>3</sub>)
- 1 sequential air sampler for NO<sub>2</sub> sampling utilising the iodide method
- 1 UV monitor for ozone measurements
- 16 forest canopy throughfall collectors and 21 ground vegetation throughfall collectors
- 4 percolation lysimeters and 23 suction lysimeters
- 16 litterfall collectors
- 16 soil temperature sensors and 16 soil moisture sensors
- 48 nitrogen mineralization tubes
- 1 V-notch weir

In addition a two-stage filterpack sampler with impactor is installed for measuring particulate matter PM<sub>2.5</sub>, PM<sub>10</sub> for campaign use.

The weather station and precipitation collectors are set up just outside the watershed. Most equipment for measuring throughfall, litterfall and soil water is placed in four macro-plots, i.e. well defined 10m x 10m squares in the catchment. These four plots are called *intensive macro-plots*. Each of the four intensive macro-plots have 4 litterfall samplers, 4 forest canopy throughfall samplers and 3 ground vegetation throughfall samplers. In addition, there are 4-5 soil water lysimeters, 12 nitrogen mineralization tubes, 4 tensiometers and 4 soil temperature sensors. The lysimeters are soil water sampling devices of two designs (for details see below). The tensiometers and temperature sensors are situated at similar depth as the soil lysimeters. At all intensive macro-plots nitrogen mineralisation rates will be studied. *Soil water-only plots*, having lysimeters and ground vegetation throughfall collectors only, are installed at 3 additional places, together spanning the variation in soil types found within the catchment. All lysimeters are installed at different soil depths, according to genetic horizons.

## 2. INSTRUMENTATION

### 2.1 Bulk precipitation collector

The bulk precipitation collector (Figure 1) is delivered by NILU Products, Norway. (<http://www.nilu.no/products/sampler/sampler.html>). The collector consists of a precipitation collector, steel ring, 2.5 litre bottle, screw cap, bug sieve, O-ring, basket and ground spike. The material used for the collectors is high-density polyethylene and the mounting stand is made of stainless steel, and the height above ground is adjustable between 1.7m to 2.6m. Diameter of collecting surface is 200mm; hence, 1 litre rainwater is equivalent to 31.42mm precipitation.

### 2.2 Wet only precipitation collector

The wet only collector (Figure 1) is from the Department of Meteorology at Stockholm University, MISU, Sweden. It consists of a precipitation collector (5L bottle), a lid and a funnel. The lid is opened when the sensor is activated by water. A heating system dries the sensor when it stops raining; hence, the lid closes again. The collector is driven by a solar panel. The sensor needs to be cleaned now and then with a tissue wetted with alcohol. 1 litre rainwater is equivalent to 26mm precipitation; hence, the diameter of the collecting surface is approximately 215mm; hence, 1 litre rainwater is equivalent to 27.54mm precipitation.



**Figure 1.** Bulk and wet only precipitation collectors

### 2.3 Weather station

It records:

- Wind speed
- Wind direction
- Temperature
- Relative humidity

The data logger records averages for every 5 minutes; however only averages of half an hour should be reported to CRAES.

## 2.4 Ozone

A UV absorption monitor is used for ozone measurements. The data logger records averages for every 5 minutes; however hourly averages should be reported to CRAES. The instrument needs frequent maintenance and calibration following the procedures from the manufacture. In general, maintenance should be done at least every third month including inlet filter exchange, leak test, checking the pressure transducer and test the performance of the scrubber. The instrument needs to be frequently calibrated (at least every third month) by doing zero and span checks. The monitor also needs to be calibrated against a transfer standard, at least every year. This transfer standard should be traceable to a NIST Standard Reference photometer.

## 2.5 Sequential NO<sub>2</sub> sampler (NILU SS2000)

The sequential NO<sub>2</sub> sampler is equipped for 8 samplers and the sampling flow is 0.5 l min<sup>-1</sup> (corresponding to about 0.7 m<sup>3</sup> in 24h). The airflow through the sampler is measured with a dry gas meter in line between the solenoid valves and the pump.

### *Air inlet*

An inverted funnel made of PTFE teflon, polypropylene, borosilicate or polyethylene should be used in order to prevent entrance of precipitation at the sampling point.

### *Tubing*

The sampling tube connection between the air inlet and the absorption system should be as short as possible, and made of PTFE teflon, polypropylene, borosilicate glass or polyethylene.

### *Filterholder with prefilter*

A filterholder with a filter should be used in front of the absorption system in order to remove particulate matter. The filter must be inert to NO<sub>2</sub>. A teflon membrane filter with a pore size 1–2 µm or a Whatman 40 cellulose filter or equivalent may be used. The filterholder and the connections to the sampling line must be air-tight. The prefilter can be used for one week.

### *Absorption system*

A 4 mm thick sintered glass filter 25 mm i.d. with a porosity of 40–60 µm enclosed in a glass bulb as shown in Fig. 2 is used as a substrate for the impregnation. The NO<sub>2</sub> gas will be absorbed on the glass sinters, which are treated with sodium iodide and other chemicals. The glass bulbs should be connected to the sampling line using short pieces of silicon tubing. Note that the sinter should be nearest to the tube leading to the solenoid valve. During transport the silicon tubing must be closed by pieces of glass, plastic rods or appropriate caps made of PP, PE or PTFE teflon.



**Figure 2.** Sintered glass filter in a glass bulb

*Measurement range and sampling efficiency*

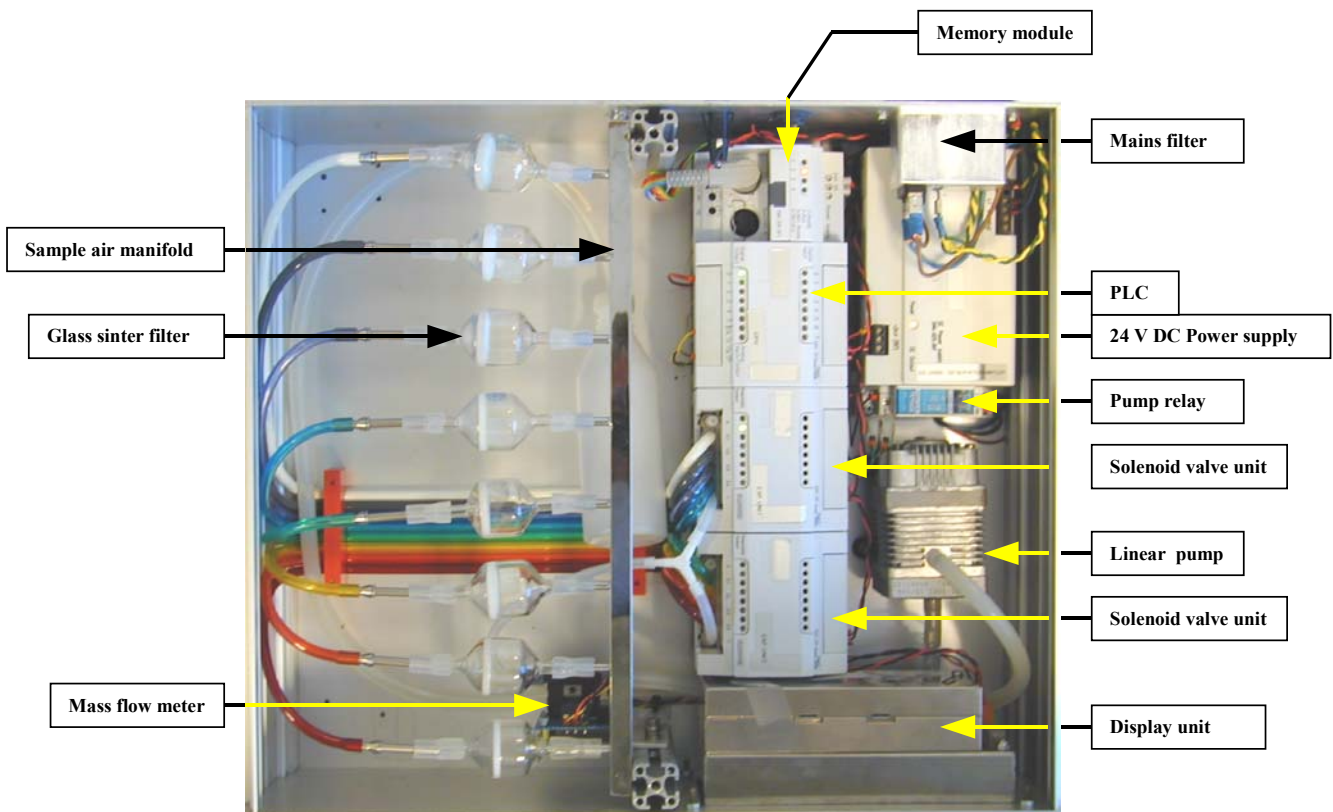
With a sample time of 24 hours, a sample volume of 0,7m<sup>3</sup> and an extraction volume of 4 ml you can detect between 0,1 and 10 µg of NO<sub>2</sub>/Nm<sup>3</sup> in ambient air. Exposed samples are stable for several weeks, which allows transfer to a remote laboratory for analysis. The sampling efficiency is greater than 98 % with a flow rate of 0,5 lpm at 15 % RH. The efficiency is better than 98 % at 60 % RH even up to 4 lpm.

*Flow control.*

The present flow control is passive. A glass capillary tube sets the flow. The mass flow sensor records and stores the accumulated volume for each sample. The flow meter is calibrated at 25 °C and 1013,25 mbar. The flow meter should be checked with a reference flow meter like BIOS.

*Pump.*

A linear motor, free piston, pump draws in the sample air. The only moving part in the pump is the piston. The piston is pushed to one side by the internal spring. The motor is driven by alternating current through an armature coil. The half cycle of the current is rectified with a diode. The active half cycle will push the piston to one side compressing the spring. The air is drawn through the inlet valve and into the cylinder. In the next half cycle the coil is idle and the spring will force the piston back while the air is expelled through the outlet valve. This pump has a free flow capacity of approximately 6 lpm of air. An electronic pump control is under development and will be introduced later. The pump will then even work when the sampler is running from the back-up battery



**Figure 3:** Instrument layout (lid removed)

*Instrument layout.*

The instrument is laid out in two sections. Facing the instrument on a bench, the left hand half is the sampling side while the right hand side contains the electronics, the display and the pump. The sample air manifold lies along the centre line of the case. The lid is in two halves. You would usually only access the sample side.

*Display and operator panel commands.*

The operator panel with display has its own real time clock with battery back up. The PLC will use this clock rather than its own to keep track of time. This is to enable the operator to adjust the system time without the need of a programming tool. The display has 4 lines of 20 characters to display the system status.

The display will show the active sample number, present flow, time and accumulated sampling time for the active sample when the sampler is operating. This is the main display. The operator can check the accumulated volumes for each exposed filter by means of the <RIGHT> arrow. The “LEFT” arrow will bring you back to the main display. The display will also revert back to main window after 15 seconds without interference from the operator. “UP” and “DOWN” arrows let you display text, which may lie outside your field of view.



**Figure 4:** Operator panel layout.

**SET correct DATE and TIME :**

Push <enter> and hold until the display below appears. Select “TIM” with the arrow keys. Push <enter> again.

The cursor will appear on the day field. Adjust the day with <UP> or <DOWN> arrow. Hit <enter> cursor will move to month field. The field is adjusted like the previous field. Hit <enter> again. You advance to next field by pushing <enter>. Hit <enter> again if the field should remain as it is. Finally you hit “clear” to exit the date adjust mode. The new date will be effective after exit.

**SET STRT, set start time**

which will recur every 24 hours by pushing <SET STRT> i.e. the #5 key pad on the panel. Push <ins> i.e. #0 key pad. Enter hours for instance 07 with the key pads. Hit <enter>. Push <SET STRT> again and then <ins>. Move to the minutes with the <RIGHT> arrow. Enter the minutes for instance 15 with the key pads. Then hit

<enter>. The display should now read **Start time: 07:15**. The display will automatically revert to the main display.

**NON STOP:**

This button will toggle between continuous sampling and single sequence sampling. Single sequence will stop after the eight filter is sampled. In continuous mode the sampler will retain pertinent data for the active filter when you temporarily stop the sampling to change filters. You simply push the <CONT> button to resume sampling on the active filter when the filters are changed. Selected mode will show up as continuous with the green LED lit and single sequence when the LED is off.

**STRT:**

Starts the sampling. This button toggles between start and stop. So when you want to stop sampling for some reason like changing filters, you actually hit <STRT>. Please observe that the sample filter is incrementing with one every second time you hit <STRT>. Start/stop status is indicated with <STRT> LED lit when the sampler is running.

**CONT:**

Is used to resume sampling without advancing to next filter.

**prn:**

is used to print out all data from the display. An optional printer will be introduced in a later version of the instrument. You can, however, transmit the data to a computer by means of a terminal emulating program and an interface cable to the display unit.

2.6 Filterpack sampler (NILU-EK)

The NILU-EK (<http://www.nilu.no/products/seq-samplers/seq-s-ek.html>) air sampler collects aerosol and gas in ambient air in a filter pack, which consists of an aerosol prefilter, and one or more impregnated filters mounted in a filter holder (<http://www.nilu.no/products/filter/filter.html>). The sampler consists of two funnels with filter holders, a control unit, a pump, a dry gas meter as well as a flow controller.

The pump draws air through the filters during a preset time period. The EK air sampler has two parallel sampling lines, each consisting of a funnel with a filter pack which are activated sequentially. The start/stop of each sampling line is set with the timer. A solenoid valve is used to change from one line to the other at the preset time. The volume of air sampled is read on an air volume counter connected to a dry gas meter. Figure 5 gives a schematic overview of the equipment.

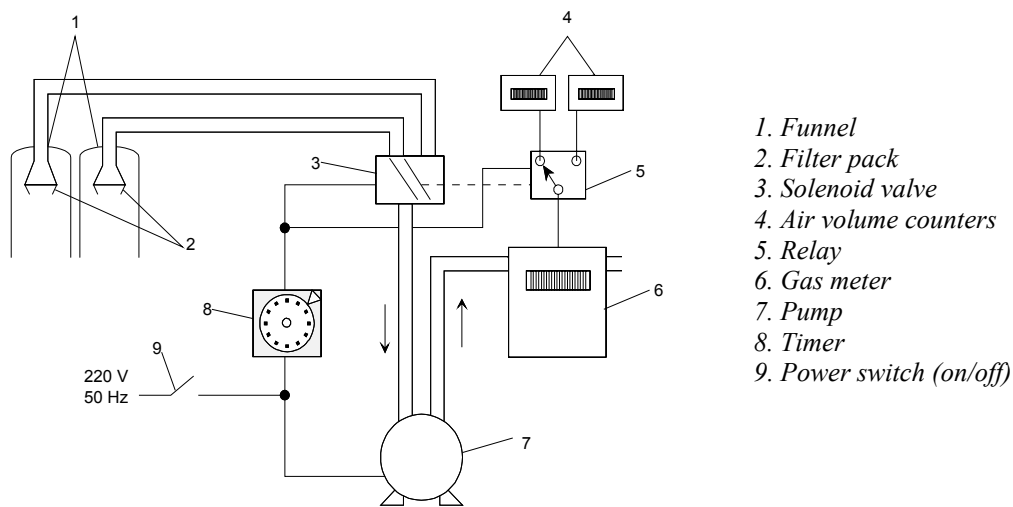
Air is drawn through the filters in the filter pack (2) when the pump (7) is operating. The position of the solenoid valve (3) determines which of the two filter packs (2) is exposed. The funnel (1) protects the filter pack against direct sunlight, rain and snow, and prevents large particles to reach the aerosol prefilter. The air volume passing through the filter is measured with the gas meter (6). For every 10 litres of air the gas meter gives a pulse to that air volume counter (4) which corresponds to the open position of the solenoid valve (3). The solenoid valve (3) and the relay (5) is controlled by the timer (8). The sampling period is preset with the timer (8), and at the end of one sampling period the solenoid valve will change sampling line and the relay



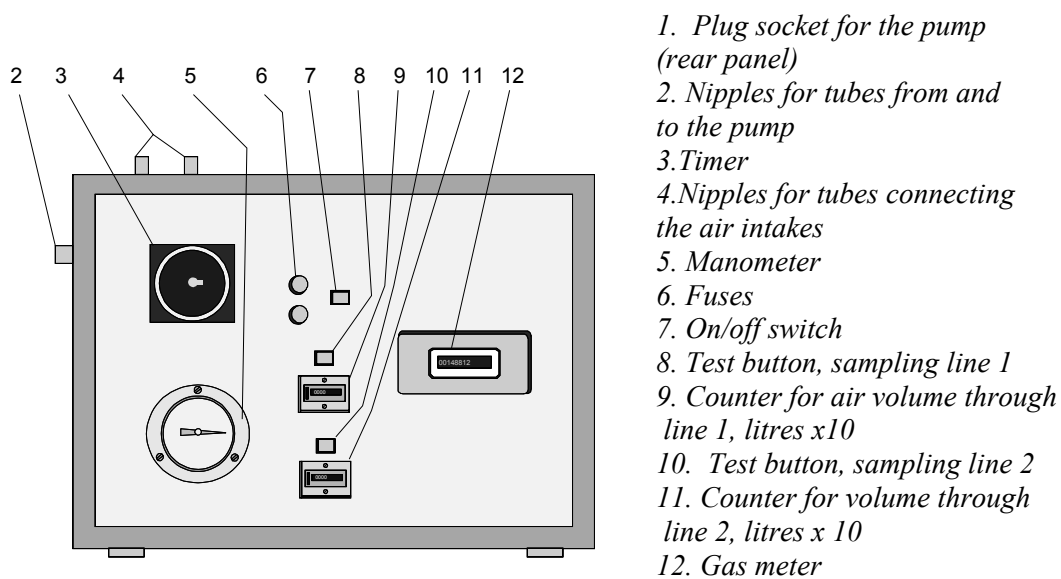
will switch to the second air volume counter. The timer will initiate a switch from one sampling line to the next at the end of every sampling period and hence, the exposed filter pack must be replaced by an unexposed one and the corresponding air volume read in the counter, before the next change of the sampling line.

Figure 6 shows the front of the control unit. The manometer (5) shows the pressure behind the filter pack in exposure. The measured pressure will reveal leakages and other problems with the sampling line. The actual pressure will, however, depend on the filter type and the aerosol mass collected on the aerosol prefilter.

The control unit is equipped with test buttons (8) and (10) for each of the two sampling lines. The light on the button tells that the corresponding sample line is activated.



**Figure 5.** Sampling principle



**Figure 6.** Control unit, front panel

By pushing the button without light, air will be sucked through the corresponding sample line as long as the button is pressed. The sampling line is immediately deactivated when the test button is released. The test buttons are used to check the pressure and the counter. The counters (9) and (11) give the air volume in units 10 litres through the two sampling lines. These counters can be reset. The gas meter (12) gives the total air volume and can not be reset.

*Making the sampler ready for use*

The control unit and the pump should be placed in a room with electrical connections which must be grounded if the room contains water pipes. The air intake, i.e. the funnel and filter pack, is mounted on a rack which is fixed to the outdoor wall when ambient air is measured. The opening of the funnel must point downwards, and the tubes from the funnels should be led through the wall to the control unit. The distance from the ground to the funnel is usually 2-5 m.

The air intake and exhaust openings in the pump should be connected by 2 tubes to the corresponding nipples on the left hand side of the control unit. The tube from the nipple with the indication "ut" must be connected with the air intake of the pump. Make certain that there is some space between the pump and the tubes, otherwise the vibrations in the pump may eventually cause holes in the tubes. The electrical cable from the pump is to be connected to the plug socket in the side of the control unit. The tubes from the two air intakes must be connected to the nipples marked "filterholder" on the top of the control unit (Filter pack #1 to be combined with system #1).

The filter holder should be mounted/dismounted in the laboratory. During the inspection the filter pack which is not active (and exposed during the previous sampling period) should be replaced with an unexposed one.

*User manual for Grässlin digital timer for NILU's filterpack sampler*



**Figure 7.** Display of Grässlin's digital timer

**Setting time and week-day**



The timer retains its program and works for 140 hours without power. The timer must, however, be reset and reprogrammed if subjected to long transport or storage time. The resetting is described below:

1. Turn off the filterpack sampler with the red switch at top of the front panel.
2. A new or empty timer will display four large zeros. Seven digits 1 through 7 will flash on the top line of the display. These numbers represent the weekdays, 1=Monday, 2= Tuesday and so on. Correct day and time will be displayed if the timer is active. **Please note that the timer will revert to the main display after 15**

**seconds of inactivity.**

3. Firmly push the clock key at the left and hold, while you push the hour key (h), to the right and repeat until the correct hour is displayed.
4. Keep the clock key depressed and push the minute key (m) at the right until correct time is displayed in the window. The numbers will advance at a higher speed if you keep the hour or minute key depressed instead of successive hits.
5. Keep the clock key depressed and push the day key (Day) at the lower right of the timer until correct weekday is shown.
6. When you release the clock key, the weekday digits will stop flashing and the correct day and time will be shown with a flashing colon. This indicates that the timer is working.

**Programming weekly sampling interval.**

1. Push the programming key (Prog.) to the left to commence programming.
  2. Push the function key with the hand symbol at the top until the actuate sign appears in the lower left of the display. 
  3. Push the hours key (h) until correct start time e.g. "08"
  4. Push the minute key (m) until correct start minute e.g. "00"
  5. Push the day key (Day) until first day that will **not** be a sampling day, e.g. "2"
  6. Confirm that this day is no sampling day by pressing Sel
  7. Continue with 5 and 6 until only all 6 days are removed and only the day of the sampling day is left in the window, e.g. "1"
  8. Push the programming key (Prog.) again.
  9. Select disengage by pushing the hand key till the sign at the right appears. 
  10. Select stop time with the hour key (h) until e.g. "08" is shown.
  11. Push the minute key (m) until correct stop minute "01" is shown. The timer activates the filter-selecting relay for only for one minute. This relay has a toggle function, which means that the selected sample line will be changed every time the relay is activated.
  12. Select the day for sampling doing the same procedure as in number 5,6 and 7
- Exit programming mode by pushing the clock key.

**Programming irregular sampling intervals.**

One assumes that the sampling shall commence at the same time on any chosen day and that the timer already is programmed to 24 hours intervals.

1. Push Prog.-key once.
2. Select the first day when no new sample is to be taken with the Day-key. Push Sel. key and the day number will begin to flash. Advance to next day with Day-key if this day is correct. A wrongly selected day will be unselected by pushing the Sel.-key a second time.
3. Repeat the procedure until you have selected all days when a new sample is to be initiated.
4. Exit programming mode by pushing the clock key.
5. Push the Prog.-key once to check the first programming line.
6. Next line will be shown next time you push the Prog.-key and so on until you get to the first empty line. The next time you push the Prog.-key again will show you how many unused programming lines there are left.

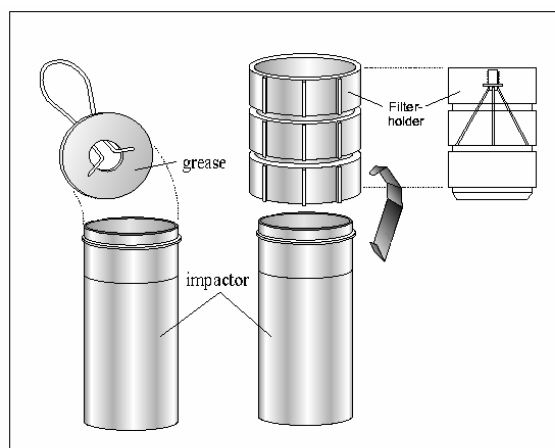
Exit from any of these lines by pushing the clock key.



### 2.7 Particle matter PM<sub>2.5</sub>/PM<sub>10</sub> sampler (NILU-EK)

The instrumentation is similar to the filterpack sampler described in 2.6. The particulate measurements do in addition have an impactor with a cut-off of 10 µm, figure 8. The coarse particles are collected on the front filter (from 2.5 – 10 µm) while the fine particles (less than 2.5 µm) penetrate through this and are collected on the second filter. This sampler will be used for episodic studies. An intensive sampling period of one month a year with daily sampling is suggested. Longer periods are also possible if wanted.

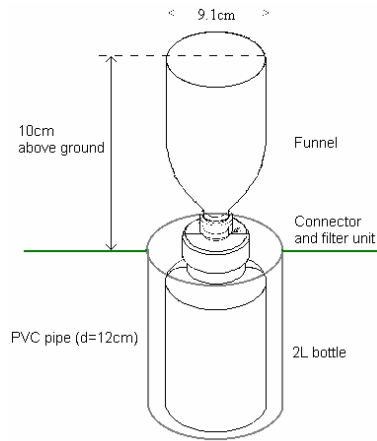
The cut off of 10 µm is valid with a constant flow rate of 10 litre/min, equivalent to 14.4 m<sup>3</sup> per day. The flat surface inside the impactor, see figure 8, needs to be greased with i.e. Vaseline.



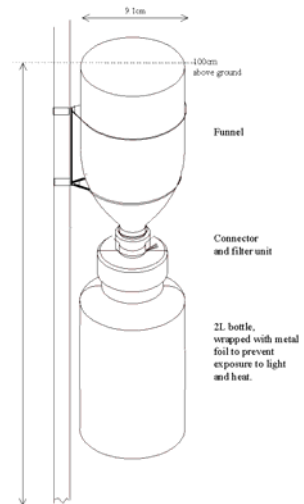
**Figure 8.** Impactor and filterholder

### 2.8 Throughfall collectors

Chinese Academy of Forestry (CAF) in China makes the ground vegetation throughfall collectors (FTF; Fig. 9) as well as the forest canopy throughfall collectors (CTF; Fig. 10) according to a design modified from the Norwegian Forest Research Institute. The collector is made of a 2L plastic bottle, a plastic funnel (d=9.1cm), a connector with a filter (nylon screen) and mounting equipment. For the ground vegetation throughfall collectors the funnel opening is located only 10cm above the ground. They are placed in a PVC pipe (Figure 9). 3 ground vegetation throughfall collectors are positioned subjectively underneath the ground vegetation at intensive macro-plots as well as soil water-only plots. The forest canopy throughfall collector is situated 1m above ground. Four forest canopy throughfall collectors are placed systematically in each intensive macroplot. At relatively flat intensive monitoring plots the forest canopy throughfall collectors are positioned on the diagonal of the 30m x 30m square, 2 m from each of the four corners. At the steep plots, the forest canopy throughfall collectors are positioned on the lower base-line of the 30m x 30m plot, at 1m, 10.3m, 19.7m and 29m from the left-hand corner.



**Figure 9.** Ground vegetation throughfall collector



**Figure 10.** Forest canopy throughfall collector

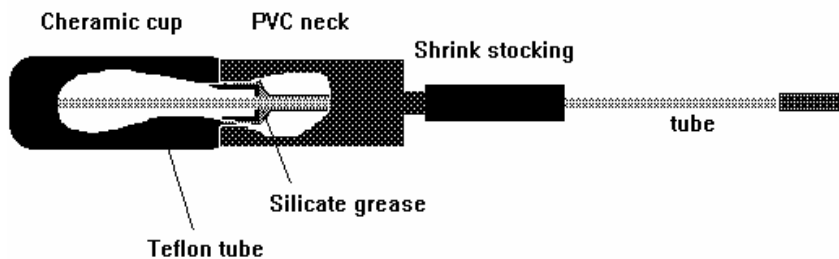
## 2.9 Lysimeters

The lysimeters are installed at the four intensive macro-plots and three soil water-only plots in the genetic soil horizons. There are two different lysimeters types; Percolation- and Ceramic cup lysimeters. The difference is whether suction ( $< 0.5$  atm.) is employed or not and in the material used.

- The Percolation lysimeter (made at UiO) are put together of 30cm x 30 cm plastic plate, nylon screen and Perspex tubing (Fig. 11).
- The Ceramic cup (Fig. 12) (Ceramic Cup, CERAMTEC, Germany. Tensiometer-celle P80 20:12x67mm item # 1.603112.00.00 (<http://www.ceramtec.de/>); Poresize: 1  $\mu$ m) is shaped as a bottle with a PVC-part pierced by the Teflon tubing. The Teflon tubing leads the soil solution from the bottom of the cup up to a pre-evacuated (approx. 0.5atm. vacuum pressure) bottle at the soil surface.



**Figure 11.** Percolation lysimeter (Note that the lysimeter drains freely through a tube connected to a 5-L collection container (poly-propylene), which is graded to allow the field recording of the collected volume.



**Figure 12.** Ceramic cup lysimeter

### 2.10 Soil temperature sensor

Thermocouples are thermistors with extension cords obtained from Geonor (<http://www.geonor.com/>) (Geonor AX24L). The sensors are used to measure soil temperature. The soil temperatures are read by a hand-held digital thermometer (Geonor C9003). The thermocouples are installed along with tensiometers (see below) at the four intensive macro-plots and are usually buried in the soil at the same depth as the lysimeters. Measurements are made in °C.

### 2.11 Soil tensiometer

Granular matrix soil water potential sensors (Watermark) are obtained from Eijkelkamp (<http://www.eijkelkamp.com/>). The tensiometers (Eijkelkamp 14.27.05) are placed in the undisturbed soil in close vicinity of the thermistors (see above). The sensors are used to measure the strength at which water is held by the soil; this is also called soil water potential. Measurements are made using a hand-held soil water potential meter (Eijkelkamp 14.27.01) after temperature correction. Readings are in centibar (=kPa). Water-saturated soil gives a reading of 0 centibar; the drier the soil, the higher the soil water potential and the measured value. The maximum value is 200 centibar.

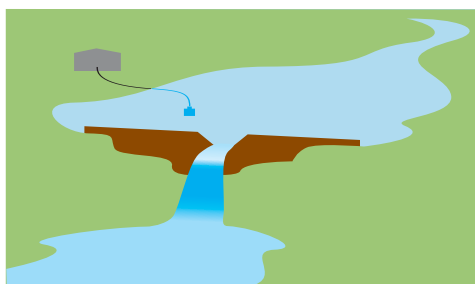
### 2.12 Limnigraph

The dam is located at the stream outlet from the catchment. The dam is used to measure the discharge, which must be determined in order to calculate catchment budgets. The dam consists of a V-notch weir, a discharge-logging device (limnigraph), and a small house for the logging device (Fig. 13). At TieShanPing a 90°, 50cm high V-notch profile in plastic is installed, with a capacity of measuring runoff from 0-0.244m<sup>3</sup> sec<sup>-1</sup> (878.4m<sup>3</sup> h<sup>-1</sup>).

The limnigraph consists of:

- Flowlogger are delivered by ISCO (<http://www.ISCO.com/>) (ISCO 4120, article no.: 68-4120-001). This probe uses an integral differential pressure transducer to measure the depth of the water and has a 7.5m sensor-cable (Fig. 8). Logging height: 0-3.05 m.
- Data collecting device consisting a Flowlogger recording unit and transformation to PC (ISCO, Model 581 RTD module, article no.: 60-9004-027); ISCO, SPA 939 (article no. 69-1780-008), which is an interface for 581 RTD and PC (battery driven (12V) with cable; and a flowlink version 4 (article no.: 60-2544-047) for programming all ISCO-loggers, emptying of data, generation of graphs, reports, summaries from stored data and exports to PC-programs like Excel for Win95/98 and NT.

The Chinese ISCO sales representative in China is: North America Instrument Consultants Co, No. 140 Feng Tai Lu Kou, Beijing 100071, China



**Figure 13.** V-notch weir and limnigraph



**Figure 14.** ISCO Flow logger

### 2.13 Temperature sensors

The electronic thermometer consists of two displays. One showing the temperature at the thermistor connected to the extension cord (i.e. outdoor), the other to the internal thermistor (i.e. indoor). The thermistor on the extension cord must be placed in open air and out of direct sunlight. The thermometer has a memory function that stores the maximum and minimum temperature recorded since last time the memory was cleared. The electronic thermometer is installed in the small house for the logging device. Measurements are made in °C.

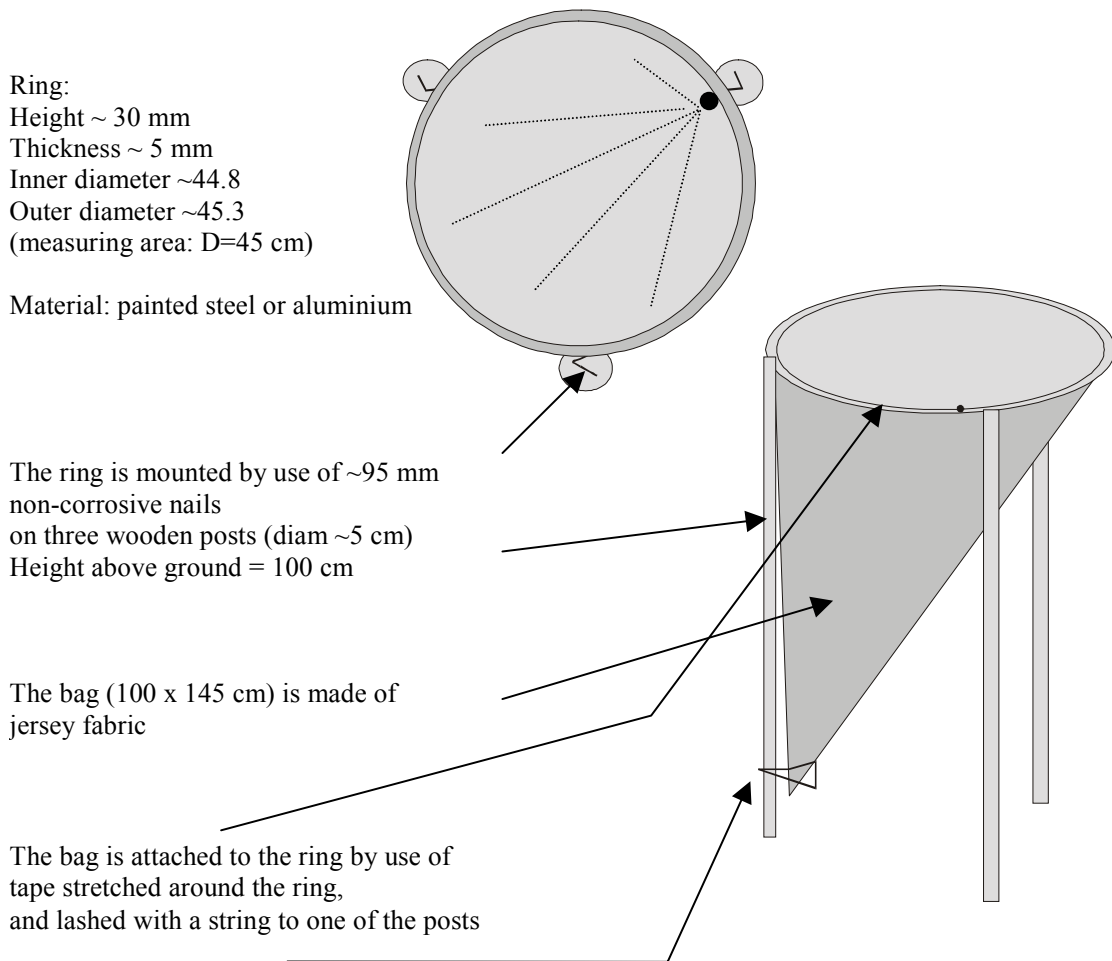
### 2.14 Macro-, core- and meso plot and tree marking

Within the catchment there are 10 forest *macro plots* (see Figure 16). Each forest macro plot consists of a 30x30m square, with a few exceptions where the measures of the plots were slightly adjusted due to terrain obstacles. The plot corners are marked with sticks with a painted band on top. Positioned in the centre of each macro plot there a 10x10m *core plot* comprising the ground vegetation meso plots which is marked similarly. Five 1-m<sup>2</sup> *meso plots* are randomly placed in each 10x10m-core plots; i.e. a total of 50 1-m<sup>2</sup> plots in the site. All corners of the 1-m<sup>2</sup> plots are permanently marked with subterranean eloxed aluminium tubes as well as by above-ground white plastic sticks. Yellow plastic numbers on the plastic stick marks the plot number of each 1-m<sup>2</sup> plot in the lower left corner of each plot. All 1-m<sup>2</sup> plots are drawn into sketch maps for each of the 10x10m plots. Inside the 30x30m plot, all trees having diameter at breast height above 5 cm outside bark, are marked with a tree number and a circular 2.5 cm point at breast height (where all diameter readings are to be taken). The exact position of corners, trees and equipment are measured using x and y coordinates.

### 2.15 Litterfall collector

Litterfall collectors are installed beside the canopy throughfall collectors (see 2.8). The litterfall collectors are made in China by local field responsible (under directions of central WPIII leader) according to a modified design from the Norwegian Forest Research Institute (NISK) (Fig. 15). Modifications are made in order to allow the application of local equipment for their construction. A litterfall collector consists of a squared 0.45m x 0.45m tree frame and bag made of brown or green Jersey fabric. The frame is 1m above the ground on wooden posts. One additional set of Jersey bags must be available to allow weekly changing of the bags.





**Figure 15** Litterfall sampler (a modification of this design from the Norwegian forest research institute)

### 2.16 Nitrogen mineralization tubes

Nitrogen mineralization tubes (15 cm long and 7 cm diameter pvc tubes) are hammered into the top soil after the L layer is carefully removed (note the F and H layer are not removed). In total 12 tubes are installed per intensive macro-plot. The tubes can be placed along two parallel 1 meter long lines (about 10 cm apart) with a distance of about 10 cm between the tubes. The tubes should be covered with a plastic cap to prevent precipitation water from entering. The caps should have small holes to allow gas exchange.

The 6 tubes situated along one of the lines are excavated immediately (i.e. the same day) after installation and brought back to the lab, while the remaining 6 stay in the soil for two months. **Important:** Make sure that the soil cores in the tubes remain intact during transport. In the lab each soil core is divided into three groups of horizons F+F, A+AB and B1 and their length is recorded. The soil samples are stored at  $-18^{\circ}\text{C}$  until extraction. Note that the samples should not be dried before extraction!

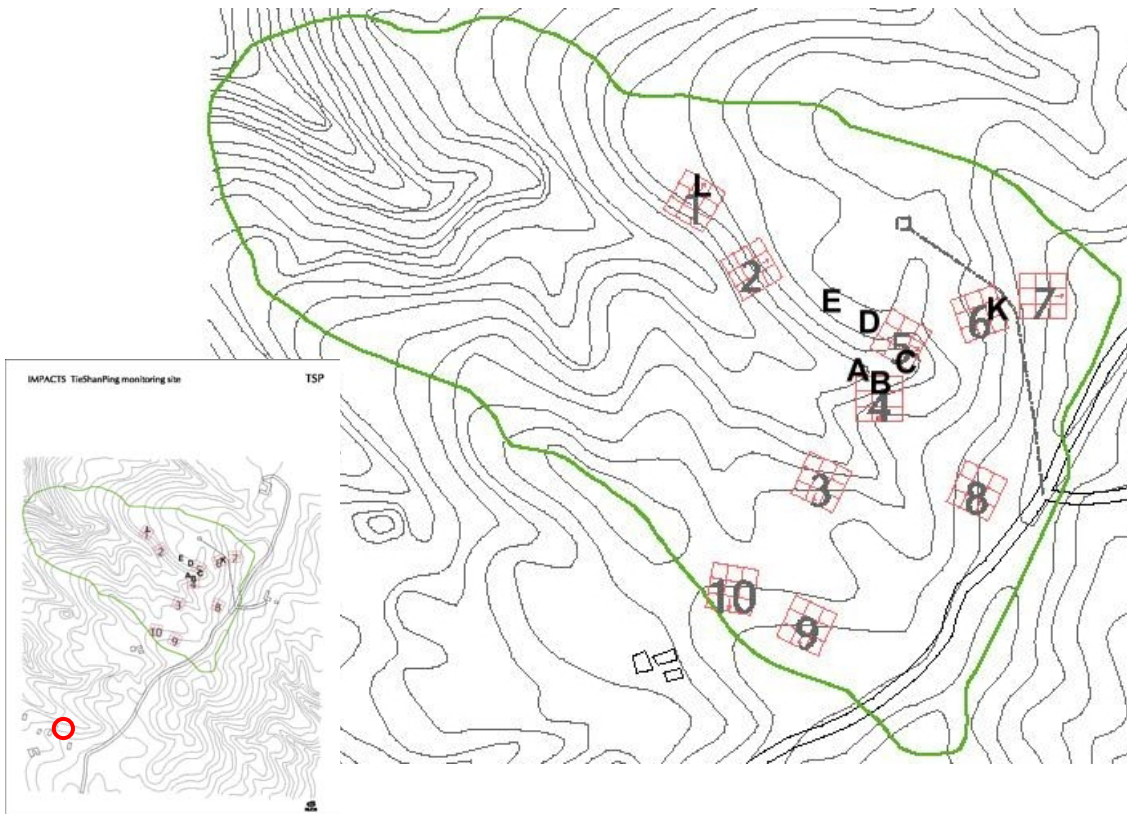
### 2.17 Instrument location

Within the catchment (Fig. 16) there are ten macro plots and four intensive macro plots. Each intensive macro-plot consist of:

- 4 forest canopy throughfall collectors

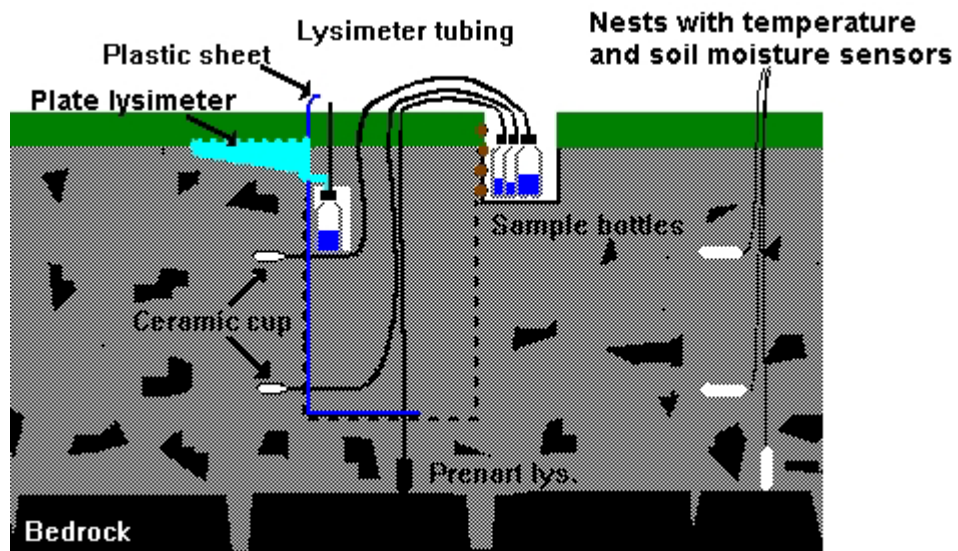
- 1 Soil water station consisting of (Fig. 17);
  - 3 ground vegetation throughfall collectors
  - 1 percolation lysimeter below the forest floor
  - 3-5 suction lysimeters situated at different depths in the soil profile
  - 4 soil temperature sensors
  - 4 tensiometers
- 4 litterfall collectors
- 12 pvc tubes (15 cm length,  $\varnothing=7$  cm) for N-mineralisation studies (not until after 2001)

Outside the intensive macroplots there are in addition three soil water-only plots, having 3 ground vegetation throughfall collectors and 3-5 suction lysimeters only. The different lysimeters are situated in the catchment according to Table 1.



**Figure 16.**

Map of the TSP catchment. Letters refer to soil water plots and numbers refer to macro-plots, both described in Table 1. Red circle indicate location of weather and air chemistry (WPI) station.



**Figure 17**

The field equipment in a soil water station. The nests of temperature and tensiometers are located to the side of the lysimeters. (Nitrogen mineralization tubes not indicated).

**Table 1**

The location of the lysimeters in the catchment.

<b>Plot</b>	<b>Location</b>	<b>Horizon</b>	<b>Depth limit cm</b>	<b>Lysimeter depth cm</b>	<b>Lys. Numb.</b>
<b>A</b>	In water saturated bottom of valley, below soilwater spring (old # 1)	A	20	15	A1
		B	-	30	A2
<b>B</b>	(old lys. plot # 2a & b)  Intensive MACROPLOT 4	L	+2	0	B0
		AB	3		
		B1	12	6	B1
		B2	27	15	B2
		B3		30	B3
<b>C</b>	(old lys. plot 3a & b and 8)  Intensive MACROPLOT 5	L	+2	0	C0
		AB	2		
		B1	12		
		B2	22	15	C1
		B3	-	30	C2
		B3	-	60	C3
		BC	-	75	C4
<b>D</b>	(old lys. plot 9 and 4)	A	2		
		B1	12		
		B2	22	15	D1
		B3	-	30	D2
		B3	-	60	D3
		BC	-	75	D4
<b>E</b>		B1	12		
		B2	22	15	E1
		B3	-	30	E2
<b>K</b>	New plot below the road into the catchment (old lys. plot 10)  Intensive MACROPLOT 6	L	+2	0	K0
		A	5	2	K1
		B1	20	8	K2
		B1	20	15	K3
		B2	30	30	K4
<b>L</b>	Along the path on the northern ridge over the dam (old lys. plot 11)  Intensive MACROPLOT 1	L	+2	0	L0
		A	2	1	L1
		B1	12	7	L2
		B2	32	22	L3
		B3	-	36	L4

### 3. PROCEDURES

Print out a copy of the appropriate weekly field log sheet on both sides of a waterproof sheet of paper to bring to the field and make sure by marking of on the sheet that all tasks are conducted. Note down the sampling date and start and stop time for the fieldwork.

#### 3.1 Precipitation

*Collection frequency:*

Bulk: daily samples to prevent a too high influence of dry deposition –composite into weekly samples for analysing; Wet only: weekly samples every monday morning (or another weekday) between 07.00 and 09.00.

##### 3.1.1 In the field

1. If there is any chance for the operator to touch the inside of the collecting funnel, disposable polyethylene gloves should be put on.
2. Bring an empty clean precipitation bottle (2.5L for bulk and 5L for wet only) and screw cap to the precipitation samplers.
3. Exchange the collection bottle in the precipitation sample collector and put on a screw-stopper. Be careful not to touch the inner side of the bottle with the fingers.
4. For the wet only, open the lid adding a few drops of water on the precipitation sensor. Examine the collector funnel (both for bulk and wet only) for visible contamination such as insects, leaves or tree-needles, organic debris. If this is found, remove the contamination. Rinse the funnel twice using distilled water (~100 ml). *This rinsing must be done even if there has been no precipitation in that collection period.*
5. After the distilled water has drained off, put on the new collection bottle.

##### 3.1.2 In the field laboratory

1. Take the wet only collection bottle and bulk sample bottle indoors to the room assigned to function as the sampling laboratory. All the equipment used for measuring must be rinsed twice with distilled water before use.
2. Record the volume of the daily bulk sample: Measure the volume in a graduated cylinder. Use a large cylinder (0-250 mL) for large samples and a small (0-25 mL) cylinder for small samples. Bulk the daily bulk samples into weekly composite sample in a <20L container.
3. The bulk precipitation amount is measured within the nearest mL. The amount is written on the weekly field log sheet. If it hasn't been any precipitation, 0 shall be written on the scheme.
4. At end of week transfer a aliquot of the composite bulk sample (100mL) to a storage sample bottle
5. Measure the volume of the wet only sample in a graduated cylinder. Use a large cylinder (0-250 mL) for large samples and a small (0-25 mL) cylinder for small samples.
6. The wet only precipitation amount is measured within the nearest mL. The amount is written on the reporting schema together with date of the measuring interval. If it hasn't been any precipitation, 0 shall be written on the scheme.
7. Transfer a suitable aliquot of the wet only sample (100mL) to a storage sample bottle

8. The sample bottles are marked with the station name, date for sampling interval, and total precipitation amount. The bottles are put in plastic bags and stored in the fridge.
9. Pour out the remainder of the samples, rinse with distilled water and place the collecting bottles upside down in a clean place to dry. Also rinse the graduated cylinders.
10. Fill in the field sample registration form, and take time to record usual and unusual events, which may have influenced the sampling.
11. pH and conductivity should be performed as soon as possible
12. Follow the instructions given in "The local lab manual".

### 3.2 Changing of filterpack

*Collection frequency:*

Weekly samples.

Print out a copy of the weekly field log sheet on both sides of a waterproof sheet of paper to bring to the station and make sure by marking of on the sheet that all tasks are conducted. Note down the sampling date and start and stop time for the fieldwork.

1. Mark an unexposed filter pack with start date
2. Read the pressure behind the exposed filter pack and record the reading in the field log in the column for under pressure, stop.
3. Read the counter in the volume meter and record the volume in the field log
4. Remove (unscrew) the air intake or funnel covering the exposed filter pack and remove (unscrew) the filter pack
5. Dismount the covers from the new unexposed filter pack and mount them on the exposed filter pack
6. Mount the new unexposed filter pack and the air intake,
7. Read the pressure behind the unexposed filter pack and record the reading in the field log in the column for underpressure, start.
8. Reset if necessary the counter or volume meter of the new filter pack
9. Write the start and stop time of the new and exposed filter pack, respectively, in the field log
10. Activate or program the timer if necessary
11. Put the exposed old filter pack in a plastic bag, seal it and put it in the refrigerator

### 3.3 Particulate mass sampler

*Collection frequency:*

Daily samples during campaign studies.

Follow the same sampling procedure as for the filterpack described in 3.2. In addition it is important to control that the flow rate is constant at 10 litre/min. The upper part of the impactor inlet as well as the interior wall needs to be cleaned and greased every week. The filterholders with filters are transported to the laboratory for weighing and analysis.

### 3.4 Sequential NO<sub>2</sub> sampler

*Collection frequency:*

Daily samples. Change seven samples once a week.

Continue noting on the back side of the Daily air field log sheet from section 3.2.

1. Check that the filter being exposed and the present flow are correct. Please note the sample number and exposure time and check that this coincides with your settings.
2. Read and note all sample volumes in the field log. The volumes appear one sample after the other when you press the <RIGHT> arrow. Please observe that shortly after volume display the display will revert to the present active sample and you will have to press the <RIGHT> arrow the appropriate number of times to recall the sample you were reading. Active sample will only read flow and not volume.
3. Write down volume and exposure time in the field log
4. Hit <STRT> button to temporarily stop the sampling.
5. Check that the <STRT> and <NON STOP> light emitting diodes (LEDs) go off.
6. Remove the 7 exposed glass bulb and seal them with red protection plugs. Leave the glass bulb that is active.
7. Disconnect the filterholder (prefilter) from its fitting and replace it with a new one.
8. Mark 7 new glass bulbs with site name and date before you connect them to their correct position on the manifold.
9. Close off inlet by pinching the silicone tube between manifold and intake filter.
10. Hit <CONT>. The LEDs lights up and the pump starts.
11. Wait till the displayed flow drops to zero and hit <STRT> again to stop.
12. Hit <SHFT> button and check that all 8 sample flows go down to zero. The sampler will automatically shift through all 8 filters. You will probably reach zero flow just as it changes to next sample.
13. If not, trouble-shoot for leakages and test again.
14. Remove the pinch.
15. Hit <SHFT> again and check that all 8 sample flows reach normal value i.e. 0,5 litres per minute. The sampler will automatically shift through all 8 filters.
16. If not, trouble-shoot for blockage.
17. Mark in the field log that leaking has been tested.
18. Hit <CONT> to continue sampling on the filter that was active. Reconfirm that you are in NON-STOP mode if that is your intention. Selection is made with the <NON STOP> key.

You can get back to the main display by pushing the <LEFT> arrow.

17. The "Remarks" column in the field log is used for information of importance to the local operator, e.g. power failure. If possible, the period of "no current" shall be given.
18. The field log should be sent together with the exposed glass sinters to the lab.

### 3.5 Throughfall

#### *Collection frequency:*

Samplers are left in the field for 1-one week before the collectors are bulked. Every fourth week the samples from the ground vegetation and forest canopy throughfall collectors in each soil water plot and intensive macro plot, respectively, are mixed to 2 grand 4 weekly bulk plot sample.

#### 3.5.1 In the field

1. 7 days after start of sampling bring graded 1, 2 and 5L rinsed polyethylene or polypropylene transport vessels out to the respective plots ensuring that the marking on the bottles match the plot letter on the throughfall collector.
2. Before handling the throughfall collectors put on disposable polyethylene gloves. Collect the total volume of all four canopy throughfall collectors and three ground vegetation throughfall collectors per plot into the transport vessels. Depending on the total volume of throughfall in the throughfall collectors choose the 1, 2 or 5 L transport vessels. After the total volume of a throughfall collector has been collected in the transport vessel the collection bottle, funnel and sieve have to be rinsed twice with distilled water. In case rinsing is not enough the equipment should be replaced (more serious cleaning can be done in the laboratory). The transport vessels containing the throughfall of all throughfall collectors at each macro- and soil water plot are brought to the institute avoiding temperature changes to the sample.

#### 3.5.2 In the laboratory

1. The volume is measured (weighing; 1g = 1mL). Next the transport vessel is marked with its collection date and stored in the refrigerator. Note amount of sample per plot for each week in field log sheet and take time to record usual and unusual events, which may have influenced the sampling (bird droppings etc.). The procedure described above is repeated every week for 4-four- weeks. Thus, after four weeks, 4 transport vessels per plot are stored in the refrigerator.
2. Chemical analyses of throughfall is done using monthly bulk samples per plot. For the monthly bulk samples we prefer to have a 1-one- L volume. Mark this bulk sample with a log number that is retrieved from the EXEL field log spreadsheet.. To obtain this we take a fraction  $x$  from each weekly bottle, where

$$x = 1/(a+b+c+d)$$

In this formula  $a$ ,  $b$ ,  $c$  and  $d$  are the pooled volumes (in L) of the four throughfall collectors for the plot in week 1, week 2, week 3 and week 4, respectively. If  $(a+b+c+d) < 1L$ , use the total volume of the 4 weeks.

3. After the weekly transport vessels have been emptied they are rinsed twice with distilled water, and the sampling date, written on the outside, is removed. Now the transport bottles are ready for re-use.
4. pH, conductivity, absorbency (OD 254<sub>nm</sub>, Colour (OD: 410<sub>nm</sub>), the Al-fractionation and total-P should be performed as soon as possible after the monthly bulk sample aliquot is made
5. Follow the instructions given in "The local lab manual".



### 3.6 Percolation lysimeters

#### *Collection frequency:*

The five-5 L collection containers are left in the field for 1-one week before they are collected. The vessels are replaced with new sample bottles. Every 4-fourth week the samples from each lysimeter are mixed to a bulk sample.

#### 3.6.1 In the field

1. 7 days after start of sampling bring the rinsed 5L graded polyethylene or polypropylene lysimeter bottles out to the respective percolation lysimeters ensuring that the marking on the bottles match the marking on the tubing.
2. Take time to record usual and unusual events, which may have influenced the sampling
3. Exchange the polyethylene or polypropylene lysimeter collection container with the rinsed container for the respective percolation lysimeter ensuring that the marking on the bottles match the marking on the tubing. Write the sampling date on the 5L container.
4. Bring the samples back to the institute avoiding temperature changes to the sample.

#### 3.6.2 In the laboratory

1. Measure (weigh) the amount of sample, write the volume on the container and note the volume in the field log.
2. Store the bulk samples in a cold, dark room (refrigerator at about 4°C)
3. Every 4-fourth week a 1L monthly bulk sample aliquot is taken for chemical processing. Use the same procedure as for the monthly bulk throughfall samples (3.5.2). Mark sample aliquot with log number that is retrieved from the EXEL field log spreadsheet.
4. The sampling containers are rinsed and made ready for re-use
5. pH, conductivity, absorbency (OD 254<sub>nm</sub>, Colour (OD: 410<sub>nm</sub>), the Al-fractionation and total-P should be performed at as soon as possible after the monthly bulk sample aliquot is made
6. Follow the instructions given in "The local lab manual".

### 3.7 Suction lysimeters

#### *Collection frequency:*

Samplers are left for 1-one week before the glass sample vessels are collected. The vessels are replaced with rinsed pre-evacuated sample bottles. Every 4-four weeks the samples from each lysimeter are mixed to a bulk sample.

#### 3.7.1 In the field

1. Pre-evacuate lysimeter glass bottles at Institute to >0.5atm. (= 50 cbar) suction. Avoid touching the inside of the bottle and rubber membrane.
2. 7 days after start of sampling bring the glass bottles out to the respective lysimeters ensuring that the marking on the bottles match the marking on the syringes.

3. Take time to record usual and unusual events, which may have influenced the sampling in the field log.
4. Exchange the glass lysimeter sampling bottles with the respective new pre-evacuated lysimeter bottles
5. Observe if pressure is lost quickly by e.g. holding the bottle up side down and look for bubbles. If so is the case refer to chap. 3.6 for remedy and make note in field book.
6. Bring the samples back to the institute avoiding temperature changes to the sample.

### 3.7.2 In the laboratory

1. Measure the amount of sample and note the data in the field log.
2. Pour the weekly sample from glass bottle to clean polyethylene or polypropylene 2L lysimeter 4-weekly bulk container ensuring that the marking on the glass bottles match the lysimeter number on the bulk containers.
3. Mark the bulk container with sampling date and sample volume
4. Store the samples in a cold, dark room (refrigerator at about 4°C)
5. Every 4-fourth week a 0.5L monthly bulk sample aliquot is taken for chemical processing. Mark sample aliquot with log number that is retrieved from the EXEL field log spreadsheet
6. The glass lysimeter bottles and (every fourth week) bulk containers are rinsed and stored and made ready for re-use.
7. pH, conductivity, absorbency (OD 254<sub>nm</sub>, Colour (OD: 410<sub>nm</sub>), the Al-fractionation and total-P should be performed at as soon as possible after the monthly bulk sample aliquot is made
8. Follow the instructions given in "The local lab manual".

During frost periods sampling must be stopped and bottles brought back to the institute.

## 3.8 Soil lysimeter problems and remedies

The sampling of soilwater above the groundwater table is not an easy task. The lysimeters are problematic and need ongoing service and maintenance. During very dry conditions only those lysimeters that always collect water may be sampled. A couple of typical problems and possible remedies will here be treated.

### 3.8.1 The lysimeter loses its pressure without sampling water.

- a) During dry conditions the porous wall of the lysimeters are not saturated with water. The lysimeters then become permeable for air and the vacuum will slowly be lost (after > 5 min.). If the problem is not constant the pressure must be reset frequently or the lysimeter temporarily abandoned.
- b) The rubber membranes may begin to leak. Check that the pressure remains stable after evacuating the flasks, if not replace the membrane. Avoid contamination of the inside of membrane. Fit cap tightly.
- c) The syringe from the lysimeter must be allowed to stand freely in the rubber membrane. If not, air will leak between the syringe and membrane

- d) The ceramic cups are fragile and may crack due to frost swelling or if stepped on (organic surface layer). If cracked replace with new lysimeter.
- e) A forest animal may disjoin the Teflon tubing. Rejoin the loose ends using the supplied tubing. If more than one tubing are detached ensure that marking on syringe corresponds to marking at soil surface. Check point c).

3.8.2 The lysimeter does not sample any water but holds its suction.

- a) The teflon tubing or the lysimeter surface may be plugged by organic or other fine material. This may often be solved by gently applying positive pressure through the syringe.
- b) The soil and the lysimeter are moist, but the soil water potential is higher than the applied suction of 50 cbar. Note that the soil water potential is measured weekly (in cbar).

### 3.9 Surface water

*Collection frequency:* weekly samples

#### 3.9.1 In the field

1. Bring an empty clean polyethylene or polypropylene bottle (1 L) with cleaned screw cap on to the sampling site.
2. The bottle should be marked with station name, sampling type (here surface water) and sampling date.
3. Collect the sample close to the discharge (runoff) device. Since a weir is present here, sampling is done at some distance up-stream from the weir because of the risk of chemical contamination from the weir material.
4. Samples are taken in such a way that contamination and sampling of surface film is avoided.
5. When filling the bottle, keep the bottleneck against the current, well below the surface.
6. Rinse the bottle and screw cap 3 times with sampling water prior to sampling full bottle.
7. Avoid touching the inside of the bottle and screw cap
8. Note in the field log that sample is collected and take time to record usual and unusual events, which may have influenced the sampling
9. The sample is transported to the laboratory avoiding temperature changes to the sample.

#### 3.9.2 In the laboratory

1. pH, conductivity, absorbency (OD 254<sub>nm</sub>, Colour (OD: 410<sub>nm</sub>) and the Al-fractionation should be performed at as soon as possible after returning to the laboratory
2. Store the samples in a cold, dark room (refrigerator at about 4°C)
3. Measurement of Alkalinity (only on water samples with pH>5) and total-P should not be stored for more than maximum one day before analysed.
4. Follow the instructions given in "The local lab manual".

### 3.10 Limmigraph

This chapter contains information about maintenance of ISCO data logger, retrieving data, calibrating and editing data

#### 3.10.1 Maintenance required for the Isco 4120

1. Keep all unused connectors capped. This prevents moisture from damaging the pins in the connectors.
2. Change the desiccant every month.
3. Replace the batteries every sixth month. The Isco 4120 Flow Logger requires a 12-volt DC power source. Use two 6-volt lantern batteries. Use only **alkaline** lantern batteries or the flow logger will not work correctly. Lantern batteries are available in local hardware stores. Place an alkaline battery at each end of the battery compartment, so that the springs contact the brass contact plates at the bottom of the compartment. To complete the circuit, the plates extend from one end of the compartment to the other. A short section of nonconductive tape insulates the plates at the center of the compartment, preventing the batteries from touching the wrong contact plate and creating a short circuit.

	Desiccant Cartridge	Desiccant Tube
<b>Location of desiccant</b>	Battery compartment	Exterior of Chase and Quick Disconnect Box
<b>How to remove desiccator</b>	Open compartment door and slide cartridge from slot	Snap the desiccant tube from mounting clip and disconnect it
<b>When to recharge</b>	Desiccant behind inspection window turns PINK, or every month.	Silica Gel: Before all desiccant in the tube turns PINK, or every month.
<b>How to recharge</b>	Heat cartridge in a vented, circulating forced air, convection oven in a well ventilated room.	Pour desiccant particles into shallow pan. Heat particles in a vented, circulating forced air, convection oven in a well-ventilated room.
<b>Oven temperature</b>	150°C	150°C
<b>How long to heat</b>	3 hours	3 hours
<b>Desiccant is recharged when...</b>	Desiccant behind the inspection window turns BLUE	Desiccant particles turn BLUE
<b>When to replace</b>	The desiccant particles no longer turn BLUE when recharged	The desiccant particles no longer turn BLUE when recharged

### 3.10.2 Retrieving data from the Isco 4120

Retrieve data from the Isco 4120 every month. To retrieve data from the 4120 you either can use the “RTD 581” (Rapid Transfer Device) or communication cable connecting directly to your portable PC.

#### *Interrogate Isco 4120 Flow logger with RTD 581*

1. Place the RTD 581 in the interrogator connector.
2. The yellow Power light will start pulsate. After a few seconds the data transfer will start. This is shown by the green light starting to pulsate.
3. When the green light stops pulsate and shows a steady green light, the data transfer is completed.
4. Take the RTD 581 to the laboratory and place it in to the RTD Interface witch is connected to a PC.
5. Start Isco Flowlink 4.1. The program has a Quick Connect button with a picture of the RTD 581 on it. Click on this button and the transfer of data will start.
6. The data will automatically add the site to the workspace in the Flowlink 4.1.

#### *Interrogate Isco 4120 Flow logger with PC*

1. Connect your PC to the instrument. Use an interrogator cable to connect to 4120.
2. Click the Quick Connect button on the toolbar, or press F11 in the Flowlink 4.1 program.
3. Flowlink starts to communicate with the instrument.
4. To retrieve data press the “ Retrieve Data (F8)” button. Flowlink adds the site to the workspace.

### 3.10.3 Calibrate the Isco 4120

Every month when retrieving data, it is necessary to measure the water level in the V-notch. This must be done to insure that the Isco 4120 is measuring the right water level in the V-notch.

1. Place the ruler in the bottom of the V-notch.
2. Aim on the top of the curve of the flowing water in the V-notch, and read the level in mm.
3. Control the level inn the V-notch against the Limnigraph reading. If the digital reading and the manual reading do not collaborate, insert the new value in to the Isco 4120.

#### *Insert new measured liquid level in Isco 4120.*

1. Connect your PC to the instrument. Use an interrogator cable to connect to Isco 4120.
2. Click the Quick Connect button on the toolbar, or press F11 in the Flowlink 4.1 program.
3. Flowlink starts to communicate with the instrument.
4. Go to the Measurements tab in the site picture. The instrument will display the current level in a number-entry box. Set the calibrated liquid level in this box and click the apply button (F9).

### 3.10.4 Editing Data

1. Start the Fowlink 4.1
2. Click on the Site Folder so the retrieved data will appear.
3. Click on the Site you wish to edit. The “Level” and “Flow rate” will appear.
4. Double click on the “Flow rate”, and a graph of the site will be shown.
5. To edit the time interval, double click on the graph. The graph properties will show up.
6. Go to the “Time Scale” tab in the graph properties. Choose the starting date and time for this plot in either relative or absolute terms. Then enter the desired time span and summary interval.
7. Click “OK” to get the new graph.

When a graph or table is active on the Flowlink desktop, selecting File>Export will export the series data shown on the graph to a CSV file (comma-separated values or comma-delimited).

1. To Export a CSV File from Flowlink menu, select File>Export.
2. Type the path and file name, or browse for the new location by clicking the Select button.
3. Click Export.
4. Flowlink displays its progress on the bare in the status box.

The exported file is now ready to be opened in programs like Microsoft Excel.

### 3.11 Temperature

Temperature is logged daily at same time as bulk precipitation is collected. Minimum and maximum temperature is logged weekly.

#### 3.11.1 In the field

Temperature is read on the electronic thermometers. Record the minimum and maximum temperature value for outdoor (in the top LCD window) by pressing the min. and max. buttons respectively. Blank the memory by pressing both buttons simultaneously. Record the minimum and maximum temperatures on back side of the fieldlog sheet.

### 3.12 Litterfall

#### 3.12.1 In the field

1. Litterfall is collected monthly.
2. Upon sampling, a numbered paper label is dropped into the bag.
3. If the bag is wet, the bag is carefully removed from the frame, and closed with a tie. A new bag is attached to the frame as described. If the bag and the litter are dry, the bag can be opened in the bottom, and the litter (together with the label) is dropped down directly into a paper bag.
4. The four bags from each macro-plot are brought to the laboratory and air-dried.

### 3.12.2 In the laboratory

1. Drying and air dry weighting: The litter is air dried, weighted bag by bag. The weight is logged in the "Sample log weekly" sheet in the Fieldlog spreadsheet. The samples are then stored in plastic bags in a cool and dry place. (In the case of wet litter, it is dried together with the bag, and then removed from the bag.)
2. Sorting: For each macro plot, the monthly samples are pooled into 3-months (13 weeks) samples, and these 16 samples are sorted into 3 fractions; 1) the coarse fraction consisting of branches, twigs and cones, 2) the masson pine needles, and 3) the rest (including leaves, lichens and seeds).
3. Weighting: The three fractions (coarse, masson pine needles, and the rest) are dried for 48 hours at 65°C. The dried fractions are weighed and stored until the end of the year.
4. Preparation for chemical analyses: At the end of the year the samples are pooled into 12 samples, one for each fraction and each macro plot. The samples are sent to Wang Yan Hui at CAF for analysis at CRAES.
5. Chemical analyses (Performed at CRAES in Beijing): The samples are analysed for the macronutrients N, P, S, Ca, K, Mg, as done with the foliar samples. However, it is not necessary to include ringtest material in these analyses.

Calculations: All data is stored in the database as g/m<sup>2</sup>/day.

### 3.13 Measurement of soil temperature and soil water potential

These measurements are done weekly, simultaneously with sampling of throughfall water and lysimeter water (chap. 3.4 - 3.6). At each plot soil temperature is measured first. Make sure that the cable is well connected to the thermometer. In case of strong corrosion the metal tips must be cleaned (e.g. by using a knife). After the temperature has been measured at all depths and the values have been recorded in the field log the measurement of the soil water potential may start.

Before connecting the measuring device to a tensiometer a check of the device should be performed. Switch on the meter, set the temperature to 24°C and press "read" and "test" simultaneously. If the reading is between 95 and 105 the device is functioning well (for details, please consult the manual).

Next the temperature has to be set to the average value that was measured in the soil (press "read" and "temperature" simultaneously to change the setting). Only when the range in soil temperatures between the different depths is more than 2°C the temperature setting has to be changed for the respective tensiometers. When the correct temperature is reached, the surface tensiometer may be connected to the device. Press "read" twice and note the value (this is given in cbar) in the field log. Continue with measuring tensiometers 2, 3, and 4 (calibration, or testing is not necessary now, but possibly the temperature setting has to be changed).

### 3.14 Measurement of nitrogen mineralization

#### *Collection frequency:*

Measurements are done during only one complete year. Sampling tubes are left in the field for 2 months before being collected. So this means there will be 6 sampling occasions. At time zero 12 tubes are installed per intensive macro-plot, 6 of these are immediately excavated, and brought back to the laboratory. After 2 months the remaining 6 tubes are excavated, brought back to the laboratory. At the same time 12 new tubes are installed again, and 6 of these are excavated and brought back to the

lab. The other 6 are left in the field and are excavated 2 months later. This procedure is repeated for one full year.

#### 3.14.1 In the field

1. Twelve tubes (individually marked) are hammered into the top soil after the L layer is carefully removed (note the F and H layer are not removed). The tubes can be placed along two 1 meter long lines (about 10 cm apart) with a distance of about 10 cm between the tubes. The tubes should be covered with a plastic cap, which has small holes to allow gas exchange.
2. The 6 tubes situated along one of the lines are excavated and brought back to the lab, while the remaining 6 stay in the soil for two months. Important: Make sure that the soil cores in the tubes remain intact during transport.
3. Note date and time of installation.

#### 3.14.2 In the laboratory

1. Upon arrival in the lab the soil cores have to be removed from the tubes with as little disturbance of the structure as possible. Use a piston to push the core out and collect it on clean laboratory tissue. With a sharp knife the soil core is divided into F+H, A+AB and B1 horizon (if applicable) and the length of each section is measured and recorded.
2. The soil sections are stored at  $-18^{\circ}\text{C}$  until extraction with KCl.
3. Samples of 10 gram from the organic horizons and 40 g of the mineral layers are weighed into 100 ml Erlenmeyer flasks and 50 ml 1M KCl is added. The flasks are gently shaken (100 strokes per minute) for 30 minutes. Suspensions are filtered (Blauband, 125 mm, Schleicher & Schull, Germany). Filtrates are stored at  $4^{\circ}\text{C}$  until analysis, which should be done as soon as possible. Concentrations of  $\text{NO}_3$  and  $\text{NH}_4$  are determined using colorimetric techniques (e.g. flow injection analysis). For details see Torstveit, 2000.

#### 3.15 Macro-, core- and meso plot and tree marking

Plot and tree markings have to be maintained. Missing sticks must be replaced. The  $1\text{ m}^2$  plots must be kept undisturbed, i.e. trampling, collections of plants, digging etc. must be avoided. All marking of corners must be kept intact. The plots must be looked after regularly to ensure that the marking is kept intact, but in a way that keep the plots undisturbed; i.e. use the sketch maps and avoid trampling also inside  $10\times 10\text{m}$  plots as far as possible. If necessary supply with new markers, but do not mark a plot if the position is uncertain, just mark on the sketch map. Always report to the Chinese and Norwegian WP-leaders when markers have been removed from a plot or any other disturbance of plots. Always mark on sketch maps and make notes on any disturbances and removal of markers.

Tree numbers and breast height points must be regularly checked and refreshed. This is important as they are quickly vanishing due to heavy showers, high temperatures and fast tree growth.



#### 4. LOGGING BOOK - HANDLING OF DATA

Registration and handling of field and laboratory data is conducted by means of an EXCEL spreadsheet Workbook ("The log-book"). The EXCEL Workbook consist of 10 excel spreadsheets.

- a) Each EXEL Workbook contain data for a 4-week period.
- b) The first two sheets are for daily fieldwork. The next 4 sheets are weekly field log forms. These are to be printed out and used in the field. All white fields should filled inn
- c) After fieldwork is completed a copy is made which replaces the form in the fieldlog. The data in the form is punched into the EXEL file and the original copy is stored in safe place.
- d) The 7<sup>th</sup> and 8<sup>th</sup> sheet holds bookkeeping with sample -date, -volume and log numbers
- e) The 9<sup>th</sup> through 11<sup>th</sup> sheets are the main sheets containing all the data for the month
- f) In the 9<sup>th</sup> sheet there are quality control columns that should be controlled