



Instruction Manual

Catalog # 171-203001

For use with Bio-Plex Manager Software Version 4.0 and MCV plate III

For technical service, call your local Bio-Rad office, or
in the US, call 1-800-4BIORAD (1-800-424-6723)

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BIO-RAD

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Section 1

Introduction

Qualification of analytical instruments is a formal process of documenting that an instrument is fit for its intended use and that it is kept maintained and calibrated. The Bio-Plex validation kit is used for operational qualification (OQ) of the Bio-Plex suspension array system. The validation kit is designed to validate the operation of all of the primary components of the system and is a valuable tool that allows the user to discriminate between assay and instrumentation problems.

The Bio-Plex validation kit consists of beads to evaluate the following components of the Bio-Plex suspension array system: 1) optics alignment, 2) integrity of fluidics, 3) reporter channel performance, and 4) classify efficiency. A brief definition of the parameter and the principle of each procedure is described, along with complete procedures for evaluating each of the primary components. An explanation of the potential impact of each process on a typical Bio-Plex cytokine assay is included to assist the user in assay troubleshooting and development.

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Section 2

Product Description

The following reagents are included in the Bio-Plex Validation Kit 4.0:

Reagent	Quantity
Optics validation bead set:	
Optics beads 1 and 2 (1×10^5 beads/ml)	2 x 10 ml black bottles
Fluidics validation bead set:	
Fluidics bead 1 (1×10^5 beads/ml)	1 x 10 ml black bottle
Fluidics bead 2	1 x 10 ml black bottle
Reporter validation bead set:	
Reporter blank, 1, 2, 3, 4, 5, and 6 (1×10^5 beads/ml)	6 x 10 ml white bottles
Classify validation bead set:	
Classify bead A(1), B(4), C(40), D(54), E(100) (1×10^5 beads/ml)	5 x 10 ml black bottles

NOTE: Validation should be performed immediately following calibration.

BizCard CD - A CD with the lot specific product specifications.

The following materials are required but not supplied:

Bio-Plex MCV plate

Bio-Rad catalog #171-203032 MCV plate III, use with Bio-Plex Manager 4.0
#171-001010

Bio-Plex Suspension array System

Bio-Rad catalog #171-000001, 171-000002, 171-000003, 171-00004,
171-000005, 171-000006, 171-000007, or 171-000009

Bio-Plex Calibration Kit

Bio-Rad catalog #171-203060

mini vortexer

sterile distilled water

70% isopropanol

10% bleach

bulb pipets

Section 3

Specifications

Specifications for the Bio-Plex validation kit may differ from lot to lot. Please refer to the package insert provided with your validation kit for specifications.

Section for Software Installation

The Bio-Plex Validation Kit 4.0 includes a BizCard-CD containing files with the specifications for the validation beads along with additional information. The file information is automatically loaded into Bio-Plex Manager 4.0 upon installation of the disc.

Section 4

Storage and Handling

The Bio-Plex validation kit beads are stable if stored at 4°C protected from light. When using the Bio-Plex validation kit, remove beads from 4°C storage and dispense into the MCV plate. Return to 4°C storage immediately following use to preserve shelf life. All components are guaranteed for 18 months from the date of manufacture when stored as specified in this manual.

Section 5

Principle of Optics Validation

Principle

The Bio-Plex array reader is a laser-based fluorescence detection system containing sensitive optics components. Alignment of the laser/optics system is critical for optimal instrument performance. A method for the assessment of the optics alignment is included in the validation kit. Acceptable specifications for the alignment procedure are listed in the product insert.

Impact on Assay Performance

The alignment of the optics bench of the Bio-Plex array reader is critical for proper assay performance. Misalignment of the reporter optics path can result in 1) reduced assay sensitivity or 2) poor well-to-well assay precision. Misalignment of the classification optics path can lead to 1) increased read times or 2) misclassification of one assay into another, leading to false positive or negative results. Correlation studies have been performed to determine the direct effect of misalignment on assay performance.

Section 6

Principle of Reporter Validation

Principle

The reporter (RP1) channel is the fluorescence channel used for assay quantitation (See Bio-Plex system hardware manual for more information regarding the principle of Bio-Plex technology). Therefore, validation of this component of the Bio-Plex system is a critical part of operational qualification. R-phycoerythrin (R-PE) is the primary reporter molecule used in Bio-Plex assays. A series of beads dyed with varying intensities of a fluorochrome spectrally matched to R-phycoerythrin are used for this procedure.

The primary reporter channel performance parameters are as follows: dynamic range, linearity, slope, accuracy, and instrument threshold (sensitivity) of the reporter channel response. Each of these parameters is related directly to the performance of the Bio-Plex array reader and has defined acceptable specifications. Definitions for the parameters and the applicability to a typical assay performed on the Bio-Plex array reader are listed below. If any of the parameters are not within the specified range, contact Bio-Rad technical support for assistance.

High Versus Low PMT Calibration and Validation

Some assays may require quantitating a range of analyte concentrations greater than what can be achieved running the instrument at a single PMT setting. In these cases, the samples may be run with different detector channel PMT settings, a High RP1 Target setting and a Low RP1 Target setting, which are set during calibration of the instrument. Turning up the PMT voltage (High RP1 Target) will allow for the ability to quantitate lower concentrations of analyte, while turning down the PMT voltage (Low RP1 Target) allows for the ability to quantitate higher concentrations of analyte. A new feature with the Bio-Plex Validation Kit 4.0 is the ability to perform validation at either of these two instrument PMT settings, rather than just at the High RP1 Target, as was previously the case.

Dynamic Range of Reporter Channel

Definition

The theoretical maximum Dynamic Range of the Bio-Plex Array Reader can be represented by the maximum number of channels in the A/D converter. Currently, the available range of channels is from 0 to 32,767, or about 4.5 decades on a log scale. These channels are represented by FI, or Fluorescence Intensity units. The Dynamic Range of the Bio-Plex Array Reader is measured by taking the log of the fluorescence intensity value of the highest reporter bead minus the fluorescence intensity of the blank bead. This value is the calibrated dynamic range of the instrument and is always less than 4.5 log decades because the highest intensity bead in the Validation Kit that is read is less than 32,767 FI units and the blank bead gives a reading that is greater than zero intensity units.

Impact on Assay Performance

It is desirable for the range of the instrument to be greater than the range of the assay. In most experiments, the dynamic range of the assay is far less than that of the instrument. However, in some assays, the same or different analytes may be present in greater than 4.5 log differences in concentration. When this is the case, the sample can be rerun at different PMT settings to extend the overall dynamic range of the measurements.

Linearity of Reporter Channel

Definition

The reporter validation bead set is utilized to construct a plot where the reporter channel median fluorescence intensity values are plotted against the corresponding expected fluorescent intensity values (as determined on an independent calibrated instrument). Instrument linearity is expressed as the coefficient of determination or R-squared (R^2) value.

Impact on Assay Performance

The linearity of the instrument response may directly affect a typical standard or calibration curve in a Bio-Plex assay, thereby impacting the unknown values extrapolated from that curve. If the R^2 value is not within acceptable limits, it may be necessary to realign the optics or check the response of the reporter photomultiplier tube.

Accuracy of Reporter Channel Response

Definition

The accuracy of the reporter channel response is a more stringent measurement of the linearity than the R^2 value. Simply stated, the accuracy of the reporter channel response is the percent difference that the regression line is away from the expected fluorescence intensity data point values.

Impact on Assay Performance

Since accuracy is also a measurement of the linearity of the instrument response, the same principles that apply to linearity also apply to accuracy of the reporter channel response. The accuracy data is evaluated in combination with optics alignment to determine if the Bio-Plex reader will perform according to specifications. It is possible for the accuracy value to fall out of specification before the linearity parameter. This is expected due to the fact that the accuracy parameter is a more sensitive measurement of linearity than the R^2 value. These data are correlated with optics alignment data as well as assay performance to determine when the array reader will not perform according to specifications.

Slope of the Reporter Channel Response

Definition

The slope of the regression line resulting from the plotting of reporter channel mean fluorescent values against expected relative fluorescence is related to the dynamic range of the instrument. The slope of the regression line is a function of the response of the reporter channel photomultiplier tube.

Impact on Assay Performance

The slope of the regression line is directly related to the dynamic range of the instrument. The slope yields direct information about the response of the photomultiplier tube. If the photomultiplier tube signal saturates at low fluorescence values, the dynamic range of the instrument is affected. The slope of the line impacts the dynamic range and the range in turn impacts the quantifiable range of an assay.

Instrument Threshold (Sensitivity)

Definition

Every instrument has an inherent level of background noise. Noise can be attributed to the laser, the PMT, the amplification electronics or the fluidics. The instrument threshold (sensitivity) of the Bio-Plex array reader is one way to demonstrate noise of the system. It is defined as the reporter channel signal of a blank bead which contains no reporter dye.

Impact on Assay Performance

The instrument threshold level demonstrated using the Bio-Plex validation kit is a low fluorescence intensity value. The typical background or zero standard of a Bio-Plex cytokine assay falls at a median fluorescence intensity of ~100. This is a desired result, as the instrument threshold should not limit the assay sensitivity. If the instrument threshold is too close to zero, then a low assay signal may be masked, if too high, then the linearity and dynamic range may be compromised.

Section 7

Principle of Classify Validation

Principle

Bio-Plex technology relies on the ability of the Bio-Plex array reader to discriminate between assay beads impregnated with varying ratios of 2 fluorescent dyes. This is the concept whereby multiplexing within a single well may occur. The periodic evaluation of the classify efficiency is necessary to complete the Bio-Plex array reader qualification process. A series of beads with varying ratios of the classification dyes are analyzed on the Bio-Plex array reader and the efficiency of multiplexing is quantitated. A classify efficiency of >80% is required for optimal results. DD Efficiency is a measure of the percentage of the Classify beads that fall within the DD Gates. The DD gates (Doublet Discriminator) are used to distinguish single beads from bead clusters, bead fragments or other small particles passing in front of the detector. Greater than 75% of the beads should fall within the gates for optimal results.

Impact on Assay Performance

Inefficient classification of beads may have several potential effects on an assay. If a bead region exhibits a classify efficiency of less than 80%, the read time of a 96-well plate may be increased. The Bio-Plex array reader tabulates a specified number of defined events in each region for each well sampled. If the percentage of beads within a specific region is low, the time required to count is increased, therefore the total time to read an entire plate is prolonged. Extremely prolonged assay read times could impact well-to-well precision, since the kinetics of a sandwich assay, for example, are not 100% stable over a period of several hours. Another potential impact of inefficient classification is the misclassification of one assay bead into another bead region. This could yield false positive or negative results for a particular assay. A DD efficiency value less than 75% may increase the read time of the assay and affect results in the same manner as a low classify efficiency.

Section 8

Principle of Fluidics Validation

Principle

The fluidics system of the Bio-Plex suspension array reader requires routine maintenance to prevent clogging and other malfunctions. Strict adherence to the maintenance procedures is mandatory for optimal instrument performance. An assessment of the integrity of the fluidics is automatically performed in the Fluidics Validation procedure. In the fluidics validation test, a sample of beads is analyzed followed by a sample of buffer to assess the carryover of beads from one well to another. This procedure should be performed once per week to ensure that assay results are not adversely affected. The fluidics path, including the sample needle must be completely free of debris and excess beads for optimal array reader performance.

Impact on Assay Performance

If a system is exhibiting a high level of carryover, due to valve malfunction or partially clogged sample needle, a significant percentage of beads may be carried over from one well to another. This phenomenon may adversely affect the median fluorescent intensity values. For example, if a well with a high median fluorescent intensity (FI) is read immediately prior to a well with a low median FI, the signal in the well with the low fluorescent intensity may shift upward. This phenomenon only occurs in extreme cases since the median fluorescent intensity statistic is robust and is not easily shifted by the introduction of a population of beads with a significantly different median FI.

Section 9

Software Utility Installation

Introduction

This section provides instructions for installing the control number information from the BizCard-CD contained in the Validation Kit into the Bio-Plex Manager database. The BizCard-CD contains the control number, expiration date, and specifications for your validation kit.

Procedure

1. Exit out of Bio-Plex Manager prior to proceeding.
2. Insert the BizCard-CD into the CD Rom drive of your computer. The installation program should start automatically. The application can also be launched through Windows Explorer by double clicking on the file "InstallControlNumbers.exe".

3. If the control numbers on the BizCard-CD already exist in the database you will receive the message that the control numbers are already installed and the program will exit. Click on the OK button and remove the BizCard-CD.
4. If the control numbers on the BizCard-CD are not found in the database you will see a dialog asking you to press OK to install the control numbers with their specifications. Click on the OK button.
5. A dialog box will appear informing you that control numbers installation is complete. You may now proceed with validation in Bio-Plex Manager using the current control number.

Section 10

Procedure for Bio-Plex Manager 4.0 and MCV plate III

Introduction

This section provides instructions for use of the Bio-Plex Validation Kit 4.0 with Bio-Plex Manager version 4.0 and MCV plate III. Bio-Plex Manager now provides a fully automated validation routine that sequentially performs all the validation tests without further user intervention. To perform all validations in a single step, follow instructions in Section 10.1. If you wish to perform an individual validation routine, these instructions are provided in Sections 10.2–10.5.

10.1 One-Step Procedure for all Validation Parameters

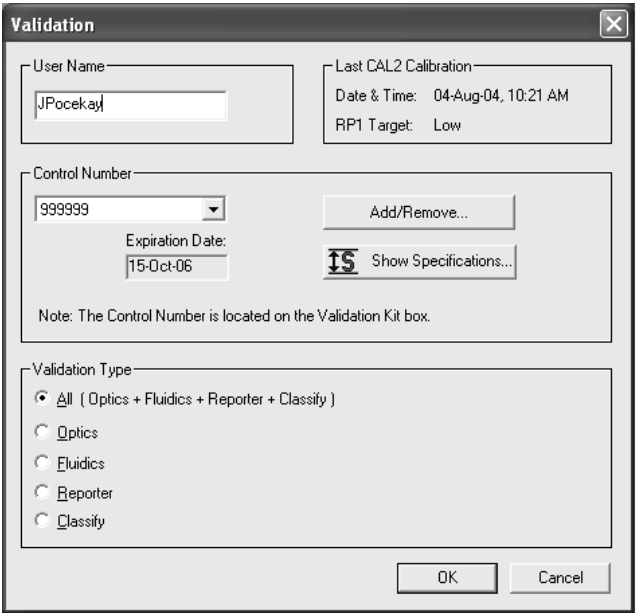
Procedure

1. Turn on the Bio-Plex array reader, microplate platform, and computer as specified in the Bio-Plex hardware and Bio-Plex Manager user manuals.
2. Perform start-up procedure as directed.
3. Calibrate Bio-Plex array reader using CAL1 and CAL2 beads found in the Bio-Plex Calibration Kit according to the Bio-Plex Manager software manual.

Note: Be sure to calibrate immediately before validation. Use either the High or Low RPI Target for CAL2 calibration depending on the type of assay being performed.

4. Remove all validation bead sets from 4°C storage and vortex each and every bottle for 30 sec. This is very important for proper validation.

5. In Bio-Plex Manager Software, select Instrument from the main menu. Select Validation from the pull-down menu. The following dialog box appears: (Figure 1A)

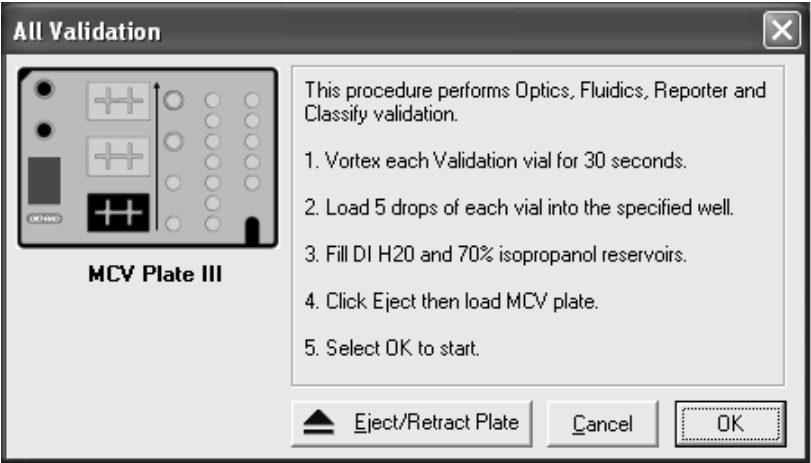


The 'Validation' dialog box contains the following fields and controls:

- User Name:** A text field containing 'JPocekay'.
- Last CAL2 Calibration:** A section containing 'Date & Time: 04-Aug-04, 10:21 AM' and 'RP1 Target: Low'.
- Control Number:** A dropdown menu showing '999999'.
- Expiration Date:** A date field showing '15-Oct-06'.
- Buttons:** 'Add/Remove...', 'Show Specifications...' (with a document icon), 'OK', and 'Cancel'.
- Note:** 'Note: The Control Number is located on the Validation Kit box.'
- Validation Type:** A group box with radio buttons for:
 - ☒ All (Optics + Fluidics + Reporter + Classify)
 - ☐ Optics
 - ☐ Fluidics
 - ☐ Reporter
 - ☐ Classify

Fig. 1A. Main validation dialog.

6. Enter user name and control number. Select All. Select OK. The following dialog appears: (Figure 1B)



The 'All Validation' dialog box contains the following elements:

- MCV Plate III:** An image of the plate with wells marked with '+' signs.
- Instructions:**

This procedure performs Optics, Fluidics, Reporter and Classify validation.

 1. Vortex each Validation vial for 30 seconds.
 2. Load 5 drops of each vial into the specified well.
 3. Fill DI H2O and 70% isopropanol reservoirs.
 4. Click Eject then load MCV plate.
 5. Select OK to start.
- Buttons:** 'Eject/Retract Plate' (with a plate icon), 'Cancel', and 'OK'.

Fig. 1B. All validation dialog.

7. Place 5 drops each of bead into the respective wells on the MCV plate III (see Figure 2 below or Figure 1B). Store beads at 4°C as soon as possible after use. Protect the beads from light.

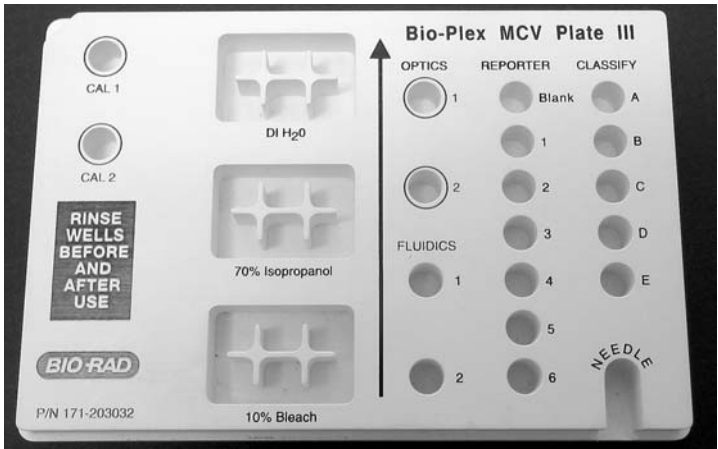


Fig. 2. MCV plate III.

8. Select the Eject button in the dialog box to eject the plate holder.
9. Place the MCV plate III in the microplate platform.
10. Select OK to begin all validation procedures.
11. When the procedure has completed, the results will be displayed in a dialog box as shown in Figure 3.

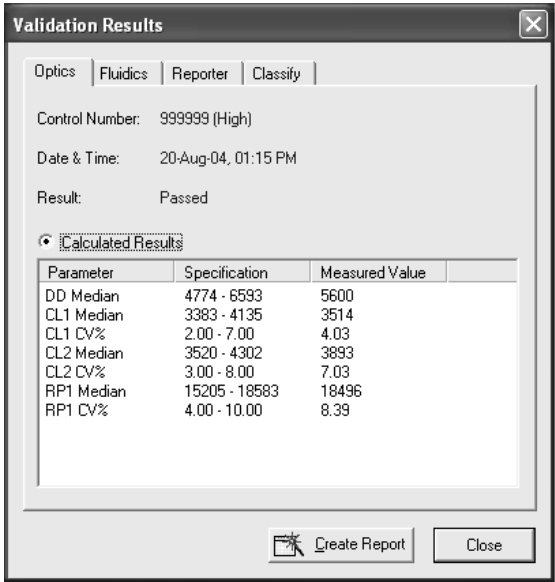


Fig. 3. Validation results.

12. Check the results for each validation. If specifications are not met for any validation, repeat that validation. If the value again is not within specification contact Bio-Rad technical support for assistance.
13. The following sections describe the steps for performing the individual validations (optics, fluidics, reporter, or classify).
14. Validation results are also logged into a validation log in Bio-Plex Manager. To access this log, select View from the main menu then Validation Log. Each type of validation is stored in a separate log for the purpose of tracking data over time. See the Bio-Plex Manager 4.0 user manual for more information on using the validation log.

10.2 Validation of Optics Alignment

Procedure

1. If not already done, follow the procedure for start-up and calibration of the Bio-Plex system.

Note: Be sure to calibrate immediately before validation. Use either the High or Low RP1 Target for CAL2 calibration depending on the type of assay being performed.

4. Remove the optics validation bead set from 4°C storage and vortex each bottle for 30 sec.
5. Place 5 drops each of optics beads 1 and 2 into the respective wells on the MCV plate III.
6. Store optics beads at 4°C as soon as possible after use. Protect the beads from light.
7. In Bio-Plex Manager Software, select Instrument from the main menu. Select Validation from the pull-down menu.
8. Enter user name and control number. Select Optics. Select OK. The following dialog appears: (Figure 4).

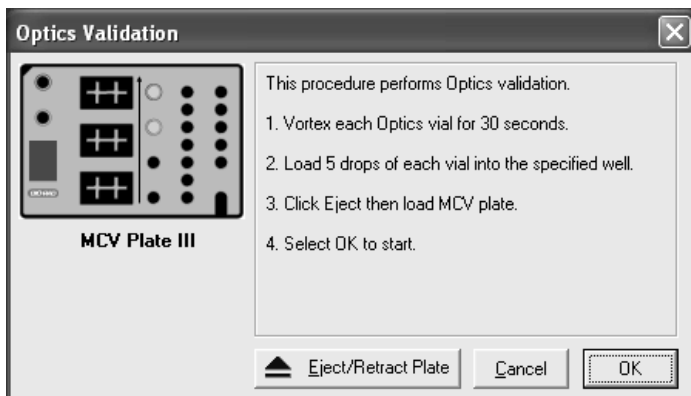


Fig. 4. Optics validation dialog.

9. Select the Eject button in the dialog box to eject the plate holder.
10. Place the MCV plate III in the microplate platform.
11. Select OK to begin the optics validation procedure.
12. When the procedure has been completed, the results will be displayed in a dialog box as shown in Figure 5.

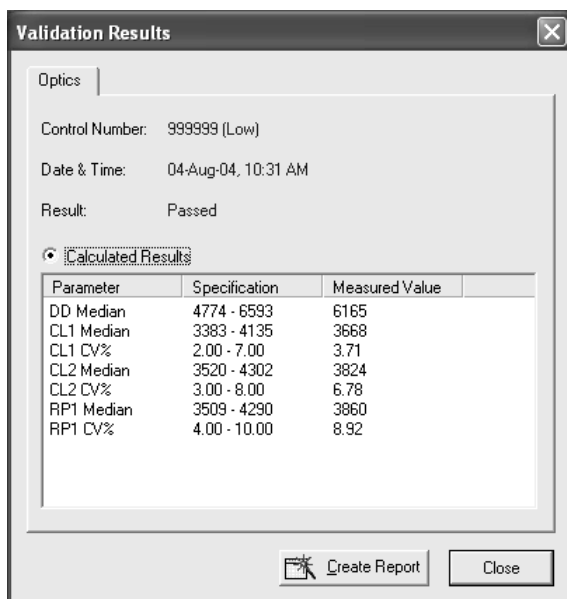


Fig. 5. Optics validation results.

10.3 Validation of Fluidics Integrity

Procedure

1. If not already done, follow the procedure for start-up and calibration of the Bio-Plex system.

Note: Be sure to calibrate immediately before validation. Use either the High or Low RP1 Target value for the CAL2 calibration depending on the type of assay being performed.

2. Select instrument, then validation from the main menu and select fluidics in the dialog box.
3. Enter user name and control number. Select OK, the following dialog box appears (Figure 10).

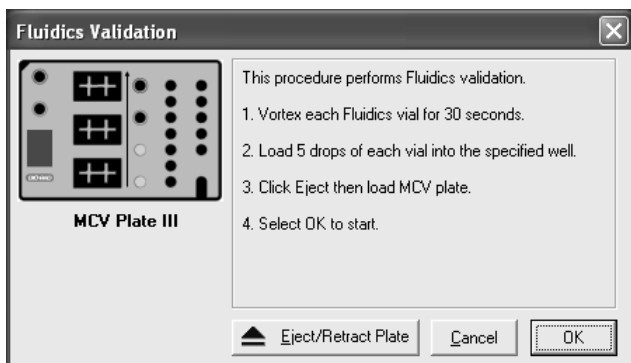


Fig. 6. Fluidics validation dialog.

4. Add 5 drops each of fluidics beads 1 and 2 to the designated wells on the MCV plate III.
5. Select the Eject button in the dialog box to eject the plate holder.
6. Place the MCV plate III in the microplate platform.
7. Select OK to begin the fluidics validation procedure.
8. When the procedure has been completed, results will appear in a dialog box as shown in Figure 7:

Validation Results

Fluidics

Control Number: 999999 (Low)

Date & Time: 04-Aug-04, 10:37 AM

Result: Passed

☒ Calculated Results

Parameter	Specification	Measured Value
% Carryover	< or = 4.0%	1.35%

Create Report Close

Fig. 7. Fluidics validation results.

- If value is not within range, repeat the procedure. If value again is not within the specification, contact Bio-Rad technical support for assistance.

10.4 Validation of Reporter Channel Performance

Procedure

- If not already done, follow the procedure for start-up and calibration of the Bio-Plex system.

Note: Be sure to calibrate immediately before validation. Use either the High or Low RP1 Target value for the CAL2 calibration depending on the type of assay being performed.

- Remove the reporter validation bead set from 4°C storage and vortex each bottle for 30 sec.
- Place 5 drops of each reporter bead into the corresponding reporter well labeled as Blank, 1, 2, 3, 4, 5, and 6 in the MCV plate III (see Figure 8).
- Store reporter beads at 4°C as soon as possible after use. Protect the beads from light.
- Fill the DI H₂O and 70% isopropanol reservoirs.
- Select Instrument from the main menu. Select Validation from the pull-down menu. A dialog will appear. Enter user name and control number. Select Reporter Validation. Select OK. The following dialog appears: (Figure 8)

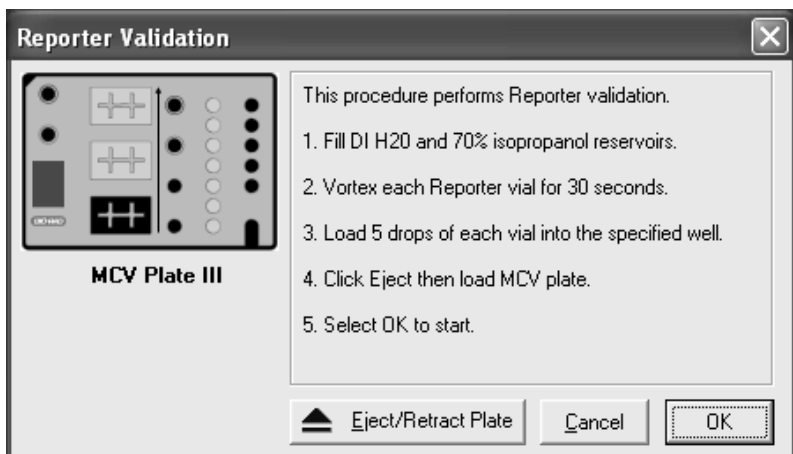


Fig. 8. Reporter validation dialog.

7. Select the Eject icon in the dialog box to eject the plate holder.
8. Place the MCV plate III in the microplate platform.
9. Select OK to start the reporter validation procedure.
10. When the procedure is completed, values will be displayed in a dialog box as shown below: (Figure 9)

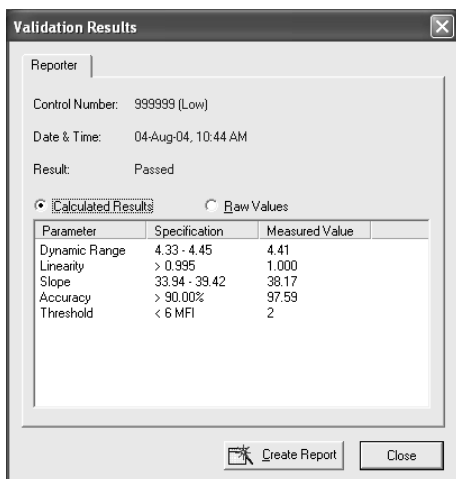


Fig. 9. Reporter validation results.

11. The results will also be logged into a validation log in Bio-Plex Manager 4.0 software. See Bio-Plex Manager 4.0 user manual for more information on using the validation log.

12. Repeat procedure if values are not within specifications. If any values are again not within acceptable ranges, contact Bio-Rad technical service for assistance.

10.5 Validation of Classify Efficiency

Procedure

1. If not already done, follow the procedure for start-up and calibration of the Bio-Plex system.

Note: Be sure to calibrate immediately before validation. Use either the High or Low RP1 Target value for the CAL2 calibration depending on the type of assay being performed.

2. Remove the classify validation bead set from 4°C storage and vortex each bottle for 30 sec.
3. Place 5 drops of each classify bead into the corresponding classify well labeled as A, B, C, D, and E in the MCV plate III (see Figure 10).
4. Store stock vials at 4°C as soon as possible after use. Protect beads from light.
5. Select Instrument from the main menu. Select Validation from the pull-down menu.
6. Enter the User name and select control number. Select classify validation. Select OK. The following dialog box appears. (Figure 10)
7. Select the Eject button to eject the plate holder.

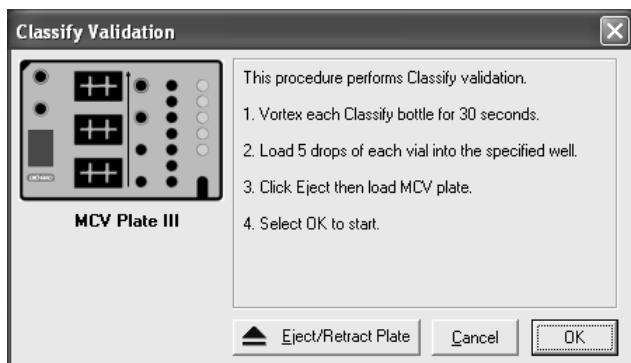


Fig. 10. Classify validation dialog.

8. Place the MCV plate III on the microplate platform.
9. Select OK to start the classify validation procedure.
10. When the procedure is completed, values will be displayed in a dialog box as shown below (Figure 11). The classify efficiency and DD efficiency results may be accessed in this view.

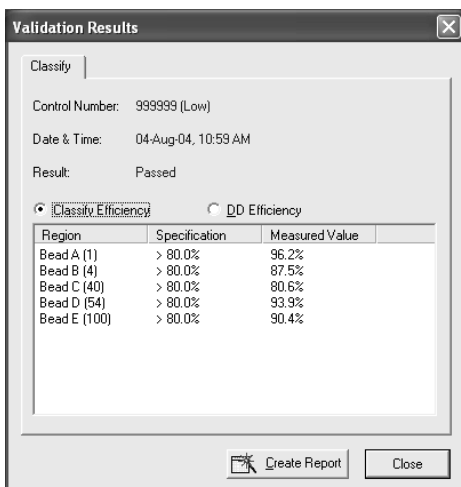


Fig. 11. Classify validation results.

11. The results will also be logged into a Validation Log in Bio-Plex Manager 4.0 software. To access this log, select view from the main menu then Validation Log. Each type of validation is stored in a separate log for the purpose of tracking data over time. See Bio-Plex Manager 4.0 software user guide for more information on using the validation log.
12. If any values do not meet specifications, repeat the procedure. If values are again not within specifications, contact Bio-Rad technical service for assistance.

10.6 Generating a Validation Report

Procedure

The results from each validation procedure are sent to a validation log in Bio-Plex Manager 4.0 software. This log may be used to create individual reports as well as track multiple validation results over time. Each type of validation is logged into a separate view: optics validation, fluidics validation, reporter validation and classify validation. You may maneuver through each of the views using either the main menu items or the toolbar icons. The specifications for each control number of the validation kit are also shown in a separate window below the results. If All validation was selected, an entry matching the specific date and time will appear in each of the validation logs. All of the validation results for a specific date and time will be included in a created report. The entire log may be printed by selecting Print then Results from the main menu.

A general procedure for creating a report from the Validation log is shown below. For more detailed instructions on the use of the validation log, consult the Bio-Plex Manager 4.0 user guide.

1. Open Bio-Plex Manager 4.0 software by clicking on the application icon on the desktop.



2. Select View from the main menu then select Validation Log from the pulldown menu. The validation log will open.
3. Choose the desired validation log by using the main menu or the toolbar icons for optics, fluidics, reporter and classify validation.



4. Click on the desired entry in the validation log. The selected row will be highlighted in black.
5. Select the create report icon. A report will automatically be generated in Microsoft Excel.



6. Print the report in Excel by selecting File then Print from the main menu. Alternatively, you may use the print button in Excel.

10.7 Validation Kit Worksheet and Report Form Examples

The following is a sample validation report from Bio-Plex Manager 4.0 software. Note that the values included here are for demonstration purposes only. Consult your product insert for values specific to your product control number.



Suspension Array System Validation Report

Test Performed by: JPocekay	Date/Time: 04-Aug-04 11:55 AM
Validation Kit Control #: 999999 (Low)	Expiration Date: 15-Oct-06
RP1 Target Value: 3885	RP1 PMT Voltage: 571.67
Reader Serial #: LX10000266002	Access Level: Unrestricted

I. Optics Validation

Result: Passed

Parameter	Specification	Measured Value	Pass/Fail
DD Median	4774 - 6593	6133	Pass
CL1 Median	3383 - 4135	3615	Pass
CL1 CV%	2.00 - 7.00	3.52%	Pass
CL2 Median	3520 - 4302	3883	Pass
CL2 CV%	3.00 - 8.00	6.93%	Pass
RP1 Median	3509 - 4290	3878	Pass
RP1 CV%	4.00 - 10.00	8.89%	Pass

II. Fluidics Validation

Result: Passed

Parameter	Specification	Measured Value	Pass/Fail
% Carryover	< or = 4.0%	1.3%	Pass

III. Reporter Validation

Result: Passed

A. Calculated Values

Parameter	Specification	Measured Value	Pass/Fail
Dynamic Range	4.33 - 4.45	4.40	Pass
Linearity	> 0.995	1.000	Pass
Slope	33.94 - 39.42	37.57	Pass
Accuracy	> 90.00%	96.94%	Pass
Threshold	< 6 MFI	2	Pass

B. Raw Values

Bead	Median FI
Blank	2
1	8
2	54
3	656
4	1576
5	4453
6	25059

IV. Classify Validation

Result: Passed

A. Classify Efficiency

Classify Bead	Specification	Measured Value	Pass/Fail
Bead A (1)	> 80.0%	98.1%	Pass
Bead B (4)	> 80.0%	93.5%	Pass
Bead C (40)	> 80.0%	90.5%	Pass
Bead D (54)	> 80.0%	95.3%	Pass
Bead E (100)	> 80.0%	93.0%	Pass

B. DD Efficiency

Parameter	Specification	Measured Value	Pass/Fail
DD Efficiency	> or = 75.0%	96.7%	Pass

Comments: _____

Reviewed by: _____ Date: _____

Section 11

Troubleshooting Guide

Problem

Optics validation procedure shows value outside of acceptable range

Reporter validation procedure shows value outside of acceptable range

Classify validation procedure shows value outside of acceptable range

Fluidics validation procedure shows value outside of acceptable range

Cause

Problem with the optical component of the array reader

Problem with the optical component of the array reader

Problem with the calibration or optical component of the array reader

Problem with fluidics lines, valves or sample needle of array reader.

Solution

Repeat the procedure. If value is still out of range, contact Bio-Rad technical support.

Repeat the procedure. If value is still out of range, contact Bio-Rad technical support.

Repeat the procedure. If values are still out of range, contact Bio-Rad technical support.

Repeat procedure. If value is still out of range, contact Bio-Rad technical support.

Section 12

Ordering Information

Catalog #	Description
171-203001	Bio-Plex Validation Kit 4.0, includes optics validation, fluidics validation, reporter validation, and classify validation bead sets for approximately 50 validation routines
171-203060	Bio-Plex Calibration Kit, includes CAL1 and CAL2 calibration beads for approximately 50 daily calibration routines
171-203032	Bio-Plex MCV Plate III for use with Bio-Plex Manager 4.0 and Validation kit 4.0

Section 13

Reference

Alder, Henry: Introduction to Probability and Statistics. Alder HL and Roessler EB (eds) W.H. Freeman, San Francisco, p118 (1968).

By purchasing this kit, which contains fluorescent labeled microsphere beads authorized by Luminex, you, the customer, acquire the rights under Luminex's patent rights* to use certain portions of this kit, including without limitation the microsphere beads contained herein, only with Luminex's laser-based fluorescent analytical test instrumentation known under the name of Luminex 100, for example as marketed by Bio-Rad Laboratories, Inc., in the Bio-Plex system.

* Including, but not limited to US patent 5,981,180; 6,046,807; 6,057,107

Certain Bio-Plex validation kit components are licensed under US patent 5,723,218

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