Quick Guide

LSM 710 / LSM 710 NLO and ConfoCor 3

Laser Scanning Microscopes

LSM Software ZEN 2008

August 2008

ZEISS

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Contents

Page

Introduction

This LSM 710 / LSM 710 NLO and ConfoCor 3 Quick Guide describes the basic operation of the LSM 710 / LSM 710 NLO and ConfoCor 3 Laser Scanning microscopes with the ZEN 2008 software.

The purpose of this document is to guide the user to get started with the system as quick as possible in order to obtain some first images from his samples.

This Quick Guide does NOT replace the detailed information available in the full user manual or in the manual of the respective microscopes (Axio Imager, Axio Observer, Axio Examiner).

Also, this Quick Guide is written for a user who is familiar with the basics of Laser Scanning Microscopy.

For your safety!
Observe the following instructions:

- The LSM 710 / LSM 710 NLO and ConfoCor 3 Laser Scanning Microscope, including its original accessories and compatible accessories from other manufacturers, may only be used for the purposes and microscopy techniques described in this manual (intended use).
- In the Operating Manual, read the chapter *Safety Instructions* carefully before starting operation.
- Follow the safety instructions described in the operating manual of the microscope and X-Cite 120 lamp / HBO 100 mercury lamp.

Starting the System

Switching on the LSM system

- Switch on the main switch (Fig. 1/**1**) and the safety lock (Fig. 1/**2**).
- When set to **ON** the power remote switch labeled **System/PC** provides power to the computer. This allows use of the computer and ZEN software offline
- To completely switch on the system, now press the **Components** switch to **ON**. This starts the other components and the complete system is ready to be initialized by the ZEN software.

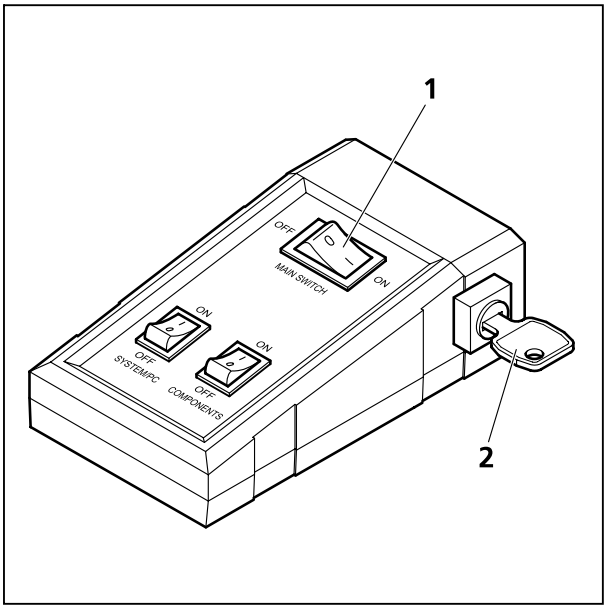


Fig. 1 **Power remote switch**

Switching on the X-Cite 120 or the HBO 100 mercury lamp

- Switch on the main switch of the X-Cite 120 / HBO 100 lamp for reflected light illumination via the power supply as described in the respective operating manual.

Switching on the Ar-ML Laser

- If the Ar-ML laser is required, turn the key (Fig. 2/**1**) and switch it on via the toggle switch (Fig. 2/**2**) on the power supply.
- After about 10 min. switch from standby to run by the switch (Fig. 2/**3**).
- Adjust the required power level with the control knob (Fig. 2/**4**) (default position should be 11 O'clock).

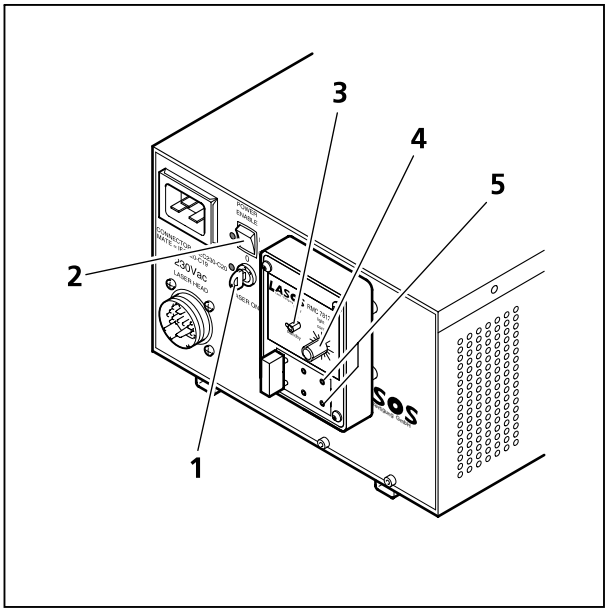
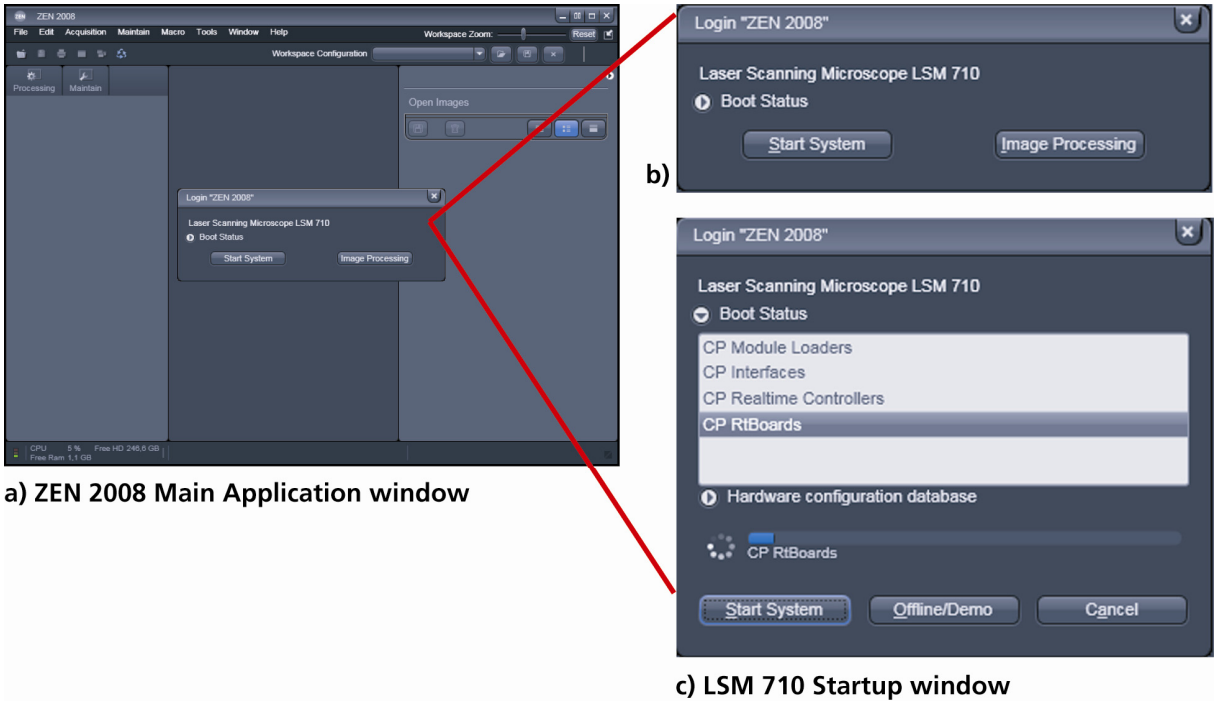


Fig. 2 **Power supply of Ar-ML laser**

Starting the ZEN software

- Double click the **ZEN 2008** icon on the WINDOWS desktop to start the Carl Zeiss LSM software.

The ZEN Main Application Window and the **LSM 710 Startup** window appear on the screen (Fig. 3)



a) ZEN 2008 Main Application window

c) LSM 710 Startup window

Fig. 3 **ZEN Main Application window at Startup (a) and the LSM 710 Startup window (b and c)**

In the small startup window, choose either to start the system **online** ("**Start System**" hardware for acquiring new images) or in **Image Processing** mode to edit already existing images**.** Toggle the little  symbol to view the Boot Status display and get the additional **Offline / Demo** button option:

- Choosing **Start System** initializes the whole microscope system and activates the entire software package for new image acquisition and analysis.
- The **Image Processing** mode ignores all hardware and activates only data handling and image processing functionality for already acquired images.
- The **Offline / Demo** mode reads the current hardware database but does not activate the system hardware for use. Instead, it simulates the system hardware for training purposes.
- Upon clicking the **Start System** button, the **Image Processing** button changes to a **Cancel** button. Click **Cancel** to interrupt/stop the **Startup** of the system.

After Startup, the ZEN Main Application Window (Fig. 4 and Fig. 5) opens. To benefit from all of Zen's features, run the window in its full screen mode.

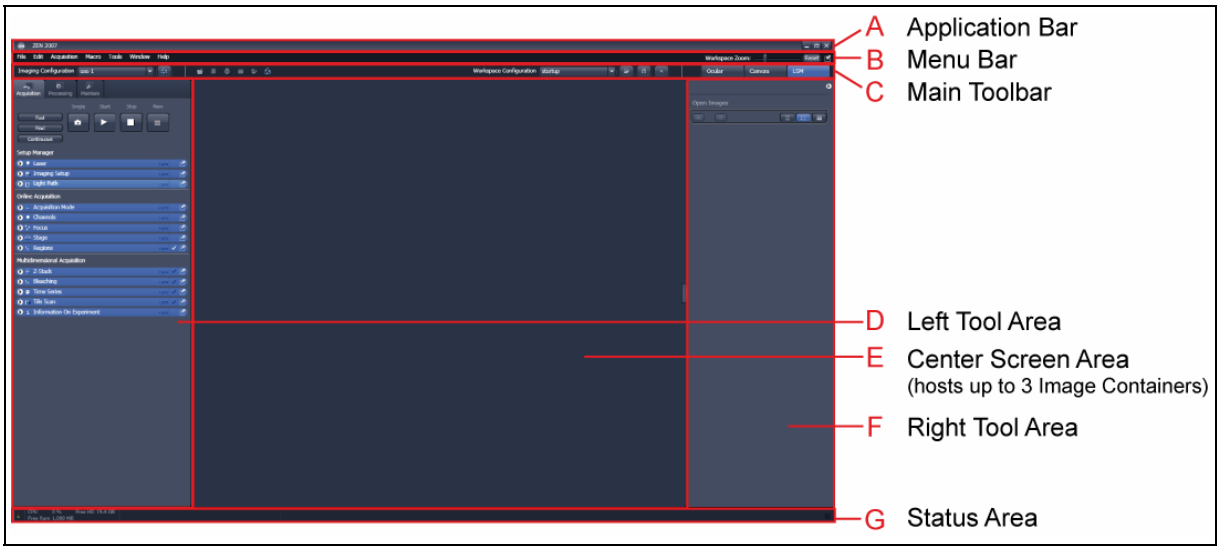


Fig. 4 **ZEN Main Application Window after Startup with empty image container**

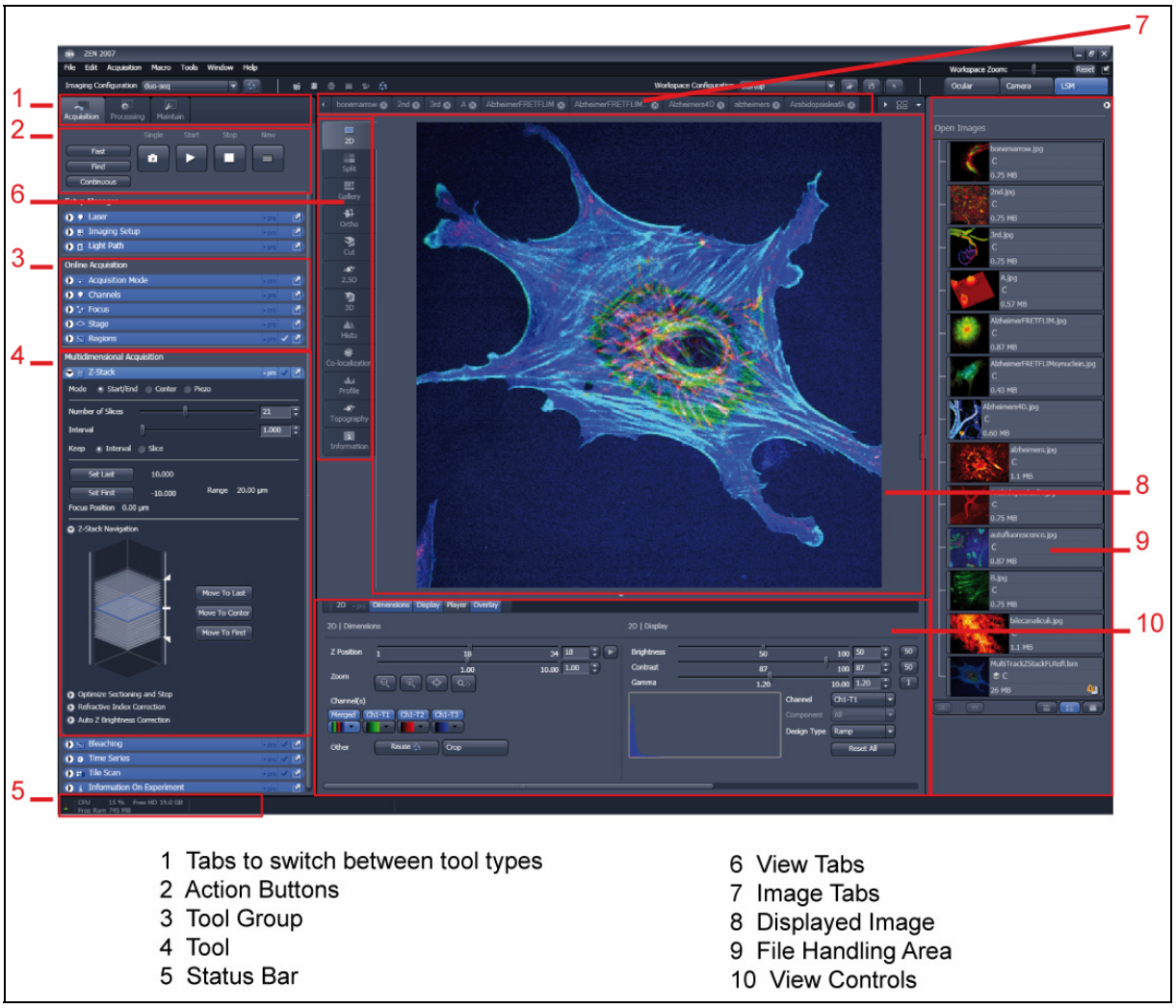


Fig. 5 **ZEN Main Application Window after Startup with several images loaded**

Introduction to ZEN – Efficient Navigation

ZEN - Efficient Navigation - is the new software for the LSM Systems from Carl Zeiss. With the launch of this software in 2007 Carl Zeiss sets new standards in application-friendly software for Laser Scanning Microscopy.

The ZEN 2008 Interface is clearly structured and follows the typical workflow of the experiments performed with confocal microscopy systems:

On the **Left Tool Area** (Fig. 4/**D**) the user finds the tools for image acquisition, image processing and system maintenance, easily accessible via 3 **Main Tabs** (Fig. 5/**1**). All functions needed to control the microscope and to acquire images are bundled in the **Acquisition Tools** (Fig. 5/**3** and **4**). Arranged from top to bottom they follow the logic of the experimental workflow. The area for viewing and interacting with images is centered in the middle of the **Main Application Window**: the **Center Screen Area**. Each displayed image can be displayed and/or analyzed with many view options available through "view tabs" which can be found on the left side of the image. According to the chosen view tab, the required view controls appear in View Control Blocks below each image. File management and data handling tools are found in the **Right Tool Area** (see Fig. 4 and Fig. 5).

Color and brightness of the interface have been carefully adjusted to the typical light conditions of the imaging laboratory, guaranteeing optimal display contrast and minimal stray light for high-sensitivity detection experiments. The **ZEN** software is optimized for a 30" TFT monitor but can also be used with dual-20" TFT setups.

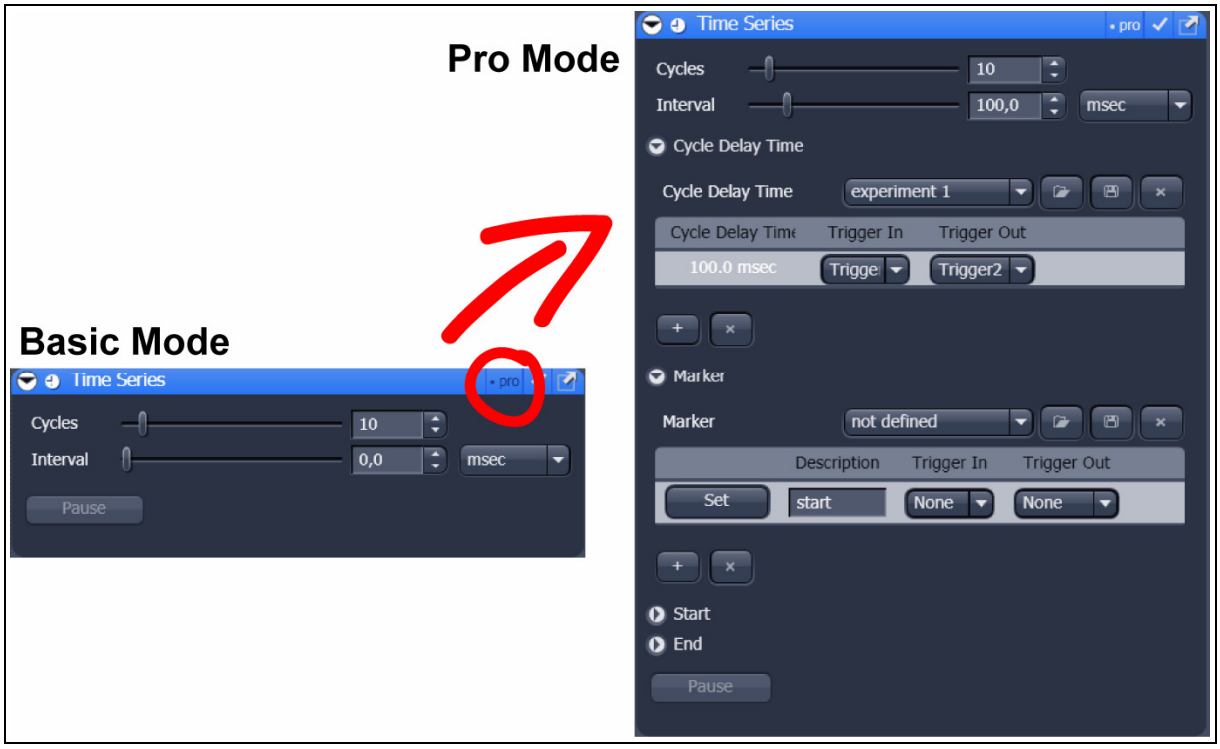


Fig. 6 **Basic and Pro Mode**

A focus in the development of **ZEN 2008** was to fulfill the needs of both basic users and microscopy specialists. Both types of users will appreciate the set of intuitive tools designed to make the use of a confocal microscope from Carl Zeiss easy and fast:

The **Pro-Basic** concept ensures that tool panels are never more complex than needed. In **Basic Mode**, the most commonly used tools are displayed. For each tool, the user can activate **Pro Mode** to display and use additional functionality (Fig. 6).

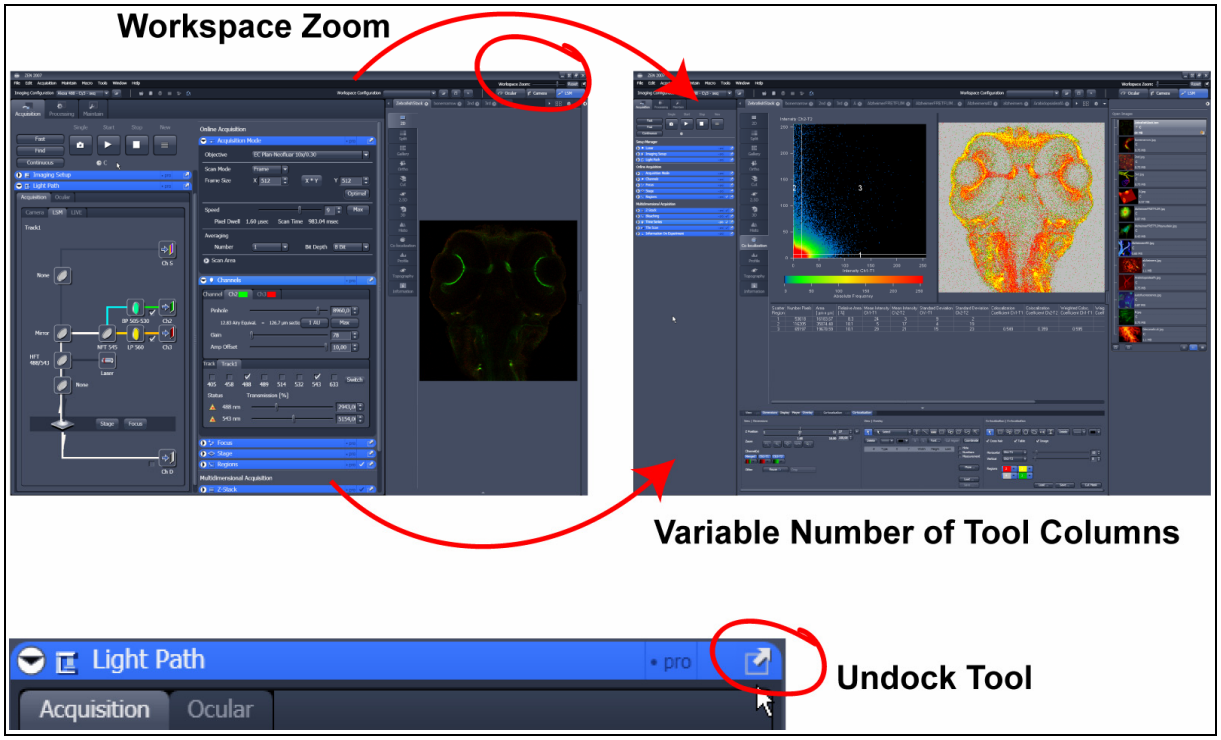


Fig. 7 **ZEN Window Layout configuration**

More features of **ZEN 2008** include:

- The user can add more columns to the **Left Tool Area** or detach individual tools to position them anywhere on the monitor. To add a column, drag a tool group by the title bar (e.g., "Online Acquisition") to the right and a new tool column automatically opens. To detach a tool, click on the little icon on the very right end of the blue tool header bar (Fig. 7).
- Another unique feature in Imaging Software is the scalable ZEN interface. This **Workspace Zoom** allows adjustment of the **ZEN 2008** window size and fonts to the situational needs or your personal preferences (Fig. 7).
- Setting up conventional confocal software for a specific experiment can take a long time and is often tedious to repeat. With ZEN these adjustments have to be done only once – and may be restored with just two clicks of the mouse. For each type of experiment one can now set-up and save the suitable **Workspace Layout**. These configurations can also be shared between users.
- For most controls, buttons and sliders, a **tool tip** is available. When the mouse pointer is kept over the button, a small pop-up window will display which function is covered by this tool/button.

These are just some of the most important features of the ZEN interface. For a more detailed description of the functionality for the **ZEN 2008** software, please refer to the User Manual that is provided with your system.

Setting up a new image document and saving your data

To create a new image document in an empty image container, click the "Single"  or the "Start"  button. The new document is immediately presented in the **Open Images Area.** Remember, an unsaved 2D image in the active image tab will be over-written by a new scan. Multi-dimensional scans or saved images will never be over-written and a new scan will then automatically create a new image document.

Alternatively you can create a new empty image document with the "New"  button in the Action Button area or the "New" function in the File Menu.

Acquired data is **not** automatically saved to disc. Make sure you save your data appropriately and back it up regularly. The ZEN software will ask you if you want to save your unsaved images when you try to close the application.

Note: There is no "image database" any more like in the earlier Zeiss LSM software versions.

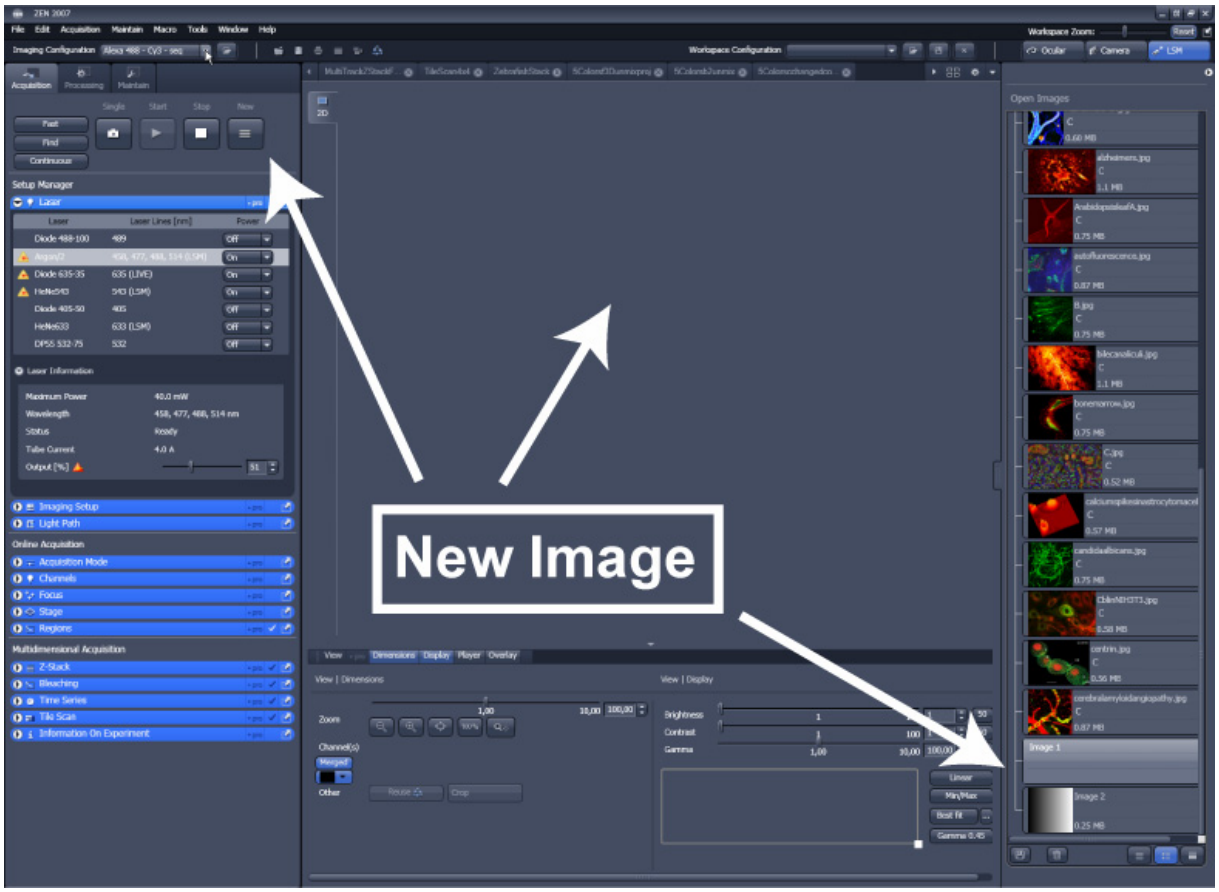


Fig. 8 **New image document in the Open Images Ares**

Advanced data browsing is available through the **File Browser** (Ctrl-F or from the **File Menu**). The File Browser can be used like the WINDOWS program file browser. Images can be opened by a double-click and image acquisition parameters are displayed with the thumbnails (Fig. 9). For more information on data browsing please refer to the detailed operating manual.

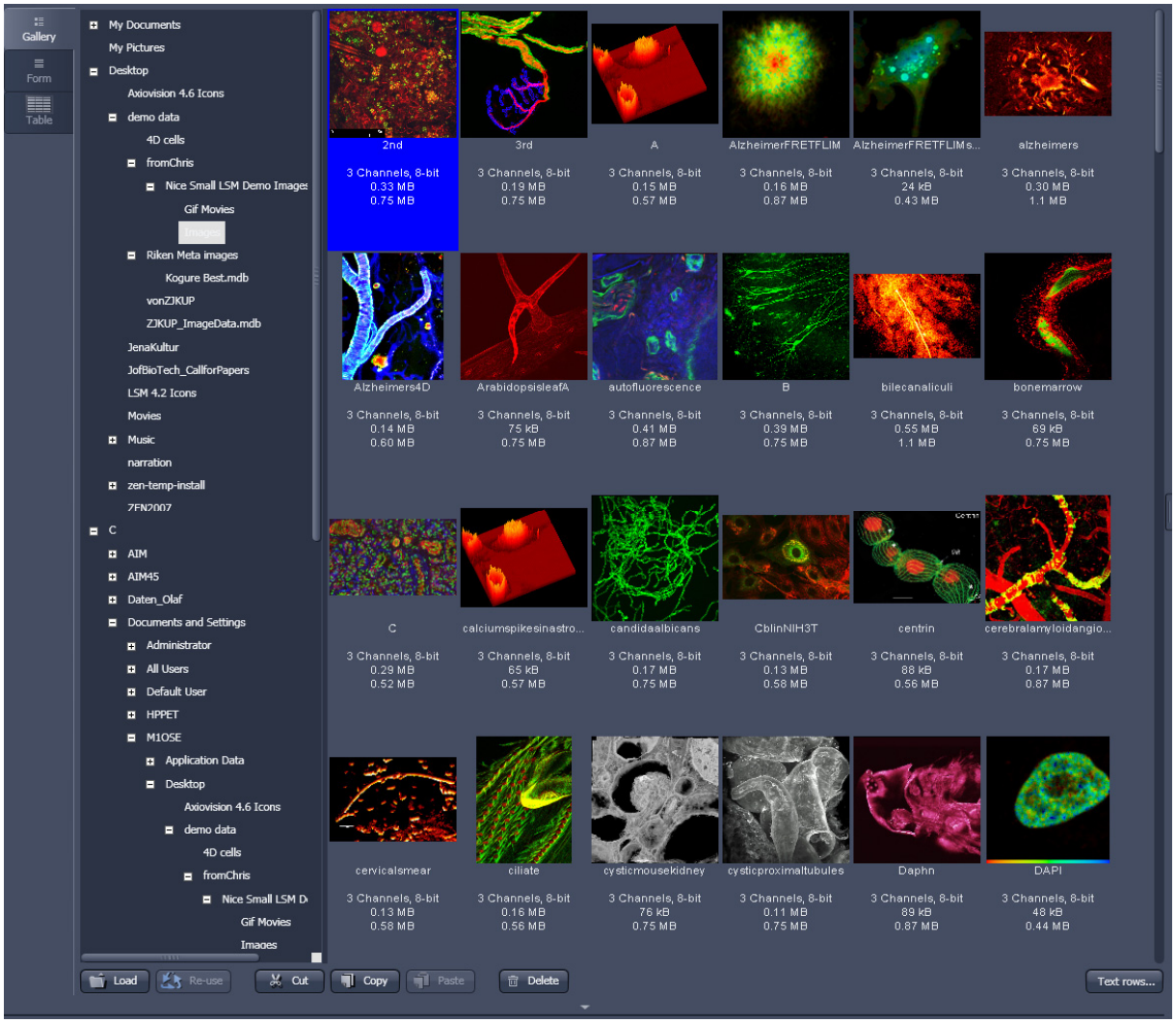


Fig. 9 **File Browser**

Turning on the lasers

- Open the **Laser Tool** in the left tool area (always the first in the list) and activate the lasers you need for your experiment (Fig. 10). Remember, the argon multi-line laser has to first be put to standby for a 5 minute warm-up before it changes to on.
- Zeiss recommends operating the Argon multi-line Laser at a tube current of about 6 A (~50 % output). This is the best compromise between laser stability/power and laser life-time. (tube-current control can be found in **Pro Mode**).

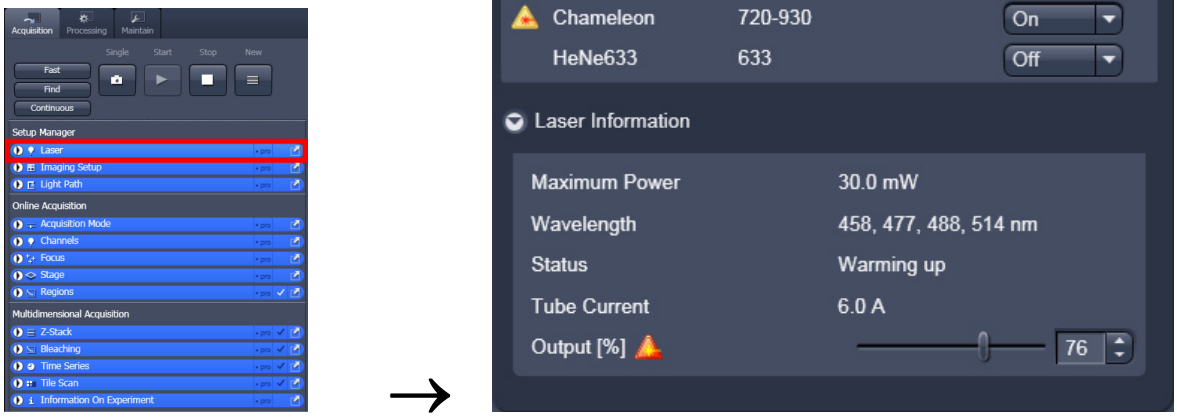
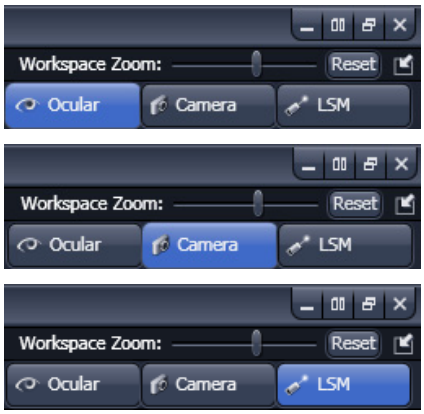


Fig. 10 **Laser Control tool (in Pro Mode)**

Setting up the microscope

Changing between direct observation, camera detection and laser scanning mode

The **Ocular**, **Camera** and **LSM** buttons switch the beam path and indicate which beam path is currently in use for the microscope:



- Click on the **Ocular** button to change the microscope beam path for direct observation via the eyepieces of the binocular tube, lasers are blocked.
- Click on the **Camera** button to change the beam path of the microscope for the port where the camera is attached for camera-based image acquisition.
- Click on the **LSM** button to set the beam path for the LSM 710 system.

Setting up the microscope and storing settings

Click on the **Ocular** button for direct observation. Then choose the **Light Path** tool from the **Left Tool Area**. Since the **Ocular** button has been chosen before, the **Oculars** tab is pre-selected and presented as the front tab in this tool (Fig. 11).

Selecting an objective

- Open the graphical pop-up menu by clicking on the **Objective** symbol and select the objective lens for your experiment (Fig. 11).
- The chosen objective lens will automatically move into the beam path.

Focusing the microscope for transmitted light

- Open the graphical pop-up menu by clicking on the **Transmitted Light** icon (Fig. 12).
- Click on the **On** button. Set the intensity of the Halogen lamp using the slider.
- Clicking outside the pop-up control closes it.
- Place specimen on microscope stage. The cover slip must be facing the objective lens. Remember the immersion medium if the objective chosen requires it!
- Use the focusing drive of the microscope to focus the object plane.
- Select specimen detail by moving the stage in X and Y using the XY stage fine motion control.

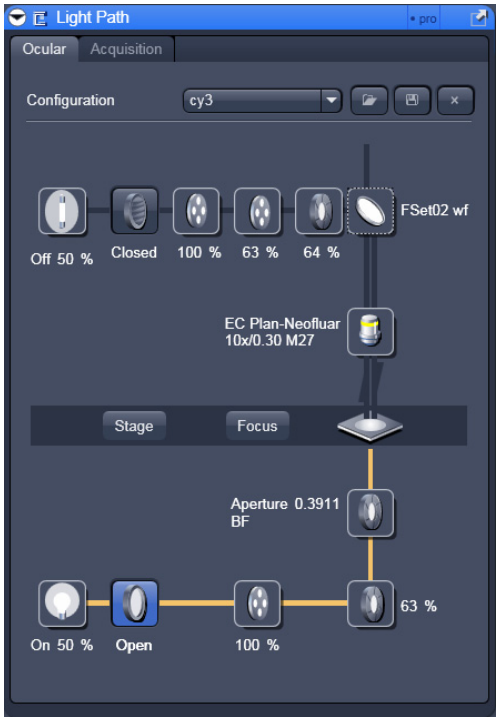


Fig. 11 **Microscope Control window, e.g.: Axio Imager.Z1**

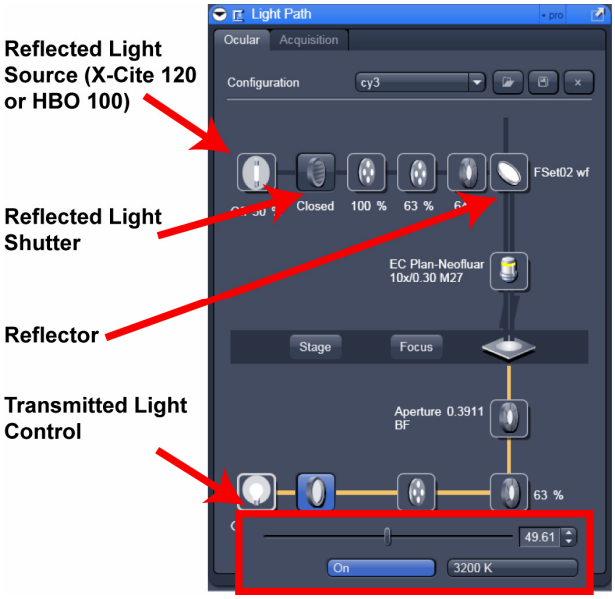


Fig. 12 **Microscope Control window with Transmitted Light pop-up menu**

Setting the microscope for reflected light

- Click on the **Reflected Light** icon to open the X-Cite 120 Controls and turn it on.
- Click on the **Reflected Light** shutter to open the shutter of the X-Cite 120 lamp / HBO100.
- Click on the **Reflector** button and select the desired filter set by clicking on it.

Storing the microscope settings

Microscope settings can be stored as configurations (Fig. 13) by typing a config name in the pull-down selector and pressing the save button. Fast restoration of a saved config is achieved by selecting the config from the pull-down list and pressing the load button. The current config can be deleted by pressing the delete button.

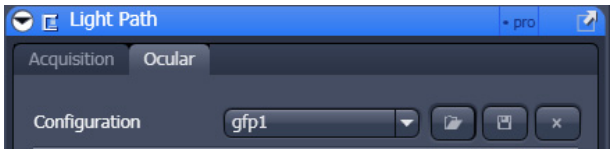
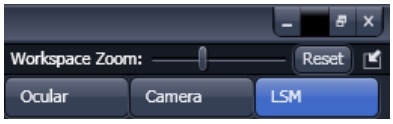


Fig. 13 **Saving Microscope configurations**

In Pro Mode, these configurations can be assigned to buttons that are easier to press.

Note: Depending on the microscope configuration, settings must be done manually if necessary.

Configuring the beam path and lasers



- Click on the **LSM** button.

Choosing a configuration

Simultaneous scanning of single, double and triple labeling:

- Advantage: faster image acquisition
- Disadvantage: Eventual cross-talk between channels

Sequential scanning of double and triple labeling; line-by-line or frame-by-frame:

- Advantage: Only one detector and one laser are switched on at any one time. This reduces cross-talk.
- Disadvantage: slower image acquisition

- Open the **Imaging Setup** and the **Light Path** tool in the **Setup Manager** Tool group to access the hardware control window to set-up the beam path.

The open **Light Path** is shown in Fig. 14.

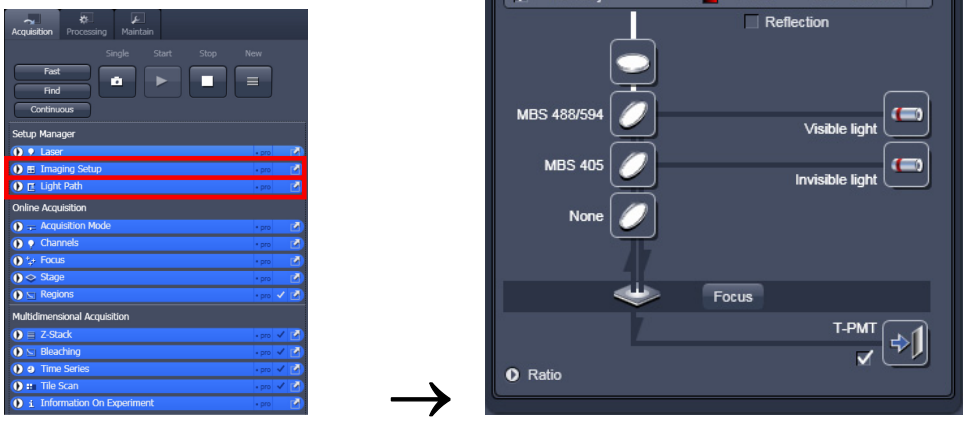


Fig. 14 **Light Path tool for a single track (LSM)**

Settings for track configuration in Channel Mode

Fig. 15 **Imaging Setup tool for a single track (LSM)**

- Select **Channel Mode** if necessary (Fig. 15).
- Click on the **LSM** tab (Fig. 14).

The **Light Path** tool displays the selected track configuration which is used for the scan procedure.

- You can change the settings of this panel using the following function elements:

Activation / deactivation of the excitation wavelengths (check box) and setting of excitation intensities (slider). If necessary open the **Laser Control** tool (see above).

Selection of the main dichroic beam splitter (MBS) from the relevant list box.

Selection of an emission filter through selection from the relevant list box.

Activation / deactivation (via check box) of the selected channel (Ch 1-4, monitor diode ChM, QUASAR detectors ChS1-8, transmission ChD) for the scanning procedure and assigning a color to the channel.

Fig. 16 **Detection Bands & Laser Lines display**

- Select the appropriate filters and activate the channels.
- Click the **Laser** icon to select the laser lines and set the attenuation values (transmission in %) in the displayed window.
- For the configuration of the beam path, please refer to the application-specific configurations depending on the used dyes and markers and the existing instrument configuration.
- In the Imaging Setup tool the **Detection Bands & Laser Lines** are displayed in a spectral panel (Fig. 16) to visualize the activated laser lines for excitation (vertical lines) and activated detection channels (colored horizontal bars).

- For **storing** a new **configuration** enter a desired name in the first line of the **Configurations** list box (Fig. 17) and click  **Store**.
- For loading an existing configuration select it from the list box and click on the  **Load** button.
- For deleting an existing configuration select it in the list box and click on  **Delete**.

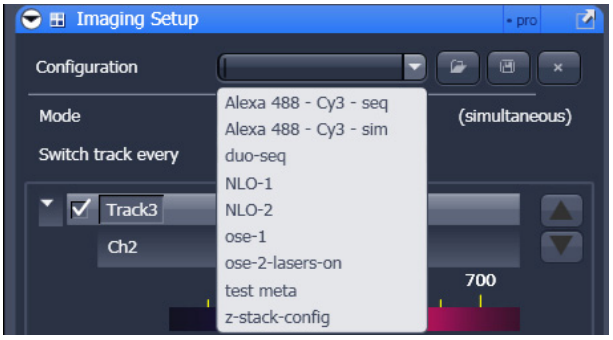


Fig. 17 **Track Configurations window**

Settings for multiple track configurations in Channel Mode

Multiple track set-ups for sequential scanning can be defined as one configuration (**Channel Mode Configuration**), to be stored under any name, reloaded or deleted.

The maximum of four tracks with up to 8 channels can be defined simultaneously and then scanned one after the other. Each track is a separate unit and can be configured independently from the other tracks with regard to channels, Acousto-Optical Tunable Filters (AOTF), emission filters and dichroic beam splitters.

The following functions are available in the **List of Tracks** panel in the **Imaging Setup Tool** (Fig. 15, Fig. 16 and Fig. 17).

Switch track every

Line Tracks are switched during scanning line-by-line. The following settings can be changed between tracks: Laser line, laser intensity and channels.

Frame Tracks are switched during scanning frame-by-frame. The following settings can be changed between tracks: Laser line and intensity, all filters and beam splitters, the channels incl. settings for gain and offset and the pinhole position and diameter.

Frame Fast The scanning procedure can be made faster. Only the laser line intensity and the **Amplifier Offset** are switched, but no other hardware components. The tracks are all matched to the current track with regard to emission filter, dichroic beam splitter, setting of Detector Gain, pinhole position and diameter. When the **Line** button is selected, the same rules apply as for **Frame Fast**.

Add Track button

An additional track is added to the configuration list in the **Imaging Setup Tool**. The maximum of four tracks can be used. One track each with basic configuration is added, i.e.: Ch 1 channel is activated, all laser lines are switched off, emission filters and dichroic beam splitters are set in accordance with the last configuration used.

 **Remove** button

The track marked in the **List of Tracks** panel is deleted.

A click on this arrow button will move the selected track (highlighted in light grey) one position upwards in the list box.

A click on this arrow button will move the selected track (highlighted in light grey) one position downwards in the list box.

Scanning an image

Setting the parameters for scanning

- Select the **Acquisition Mode** tool from the **Left Tool Area** (Fig. 18).
- Select the **Frame Size** as predefined number of pixels or enter your own values (e.g. 300 x 600) in the **Acquisition Mode** tool. Click on the **Optimal** button for calculation of appropriate number of pixels depending on objective N.A. and λ.

The number of pixels influences the image resolution!

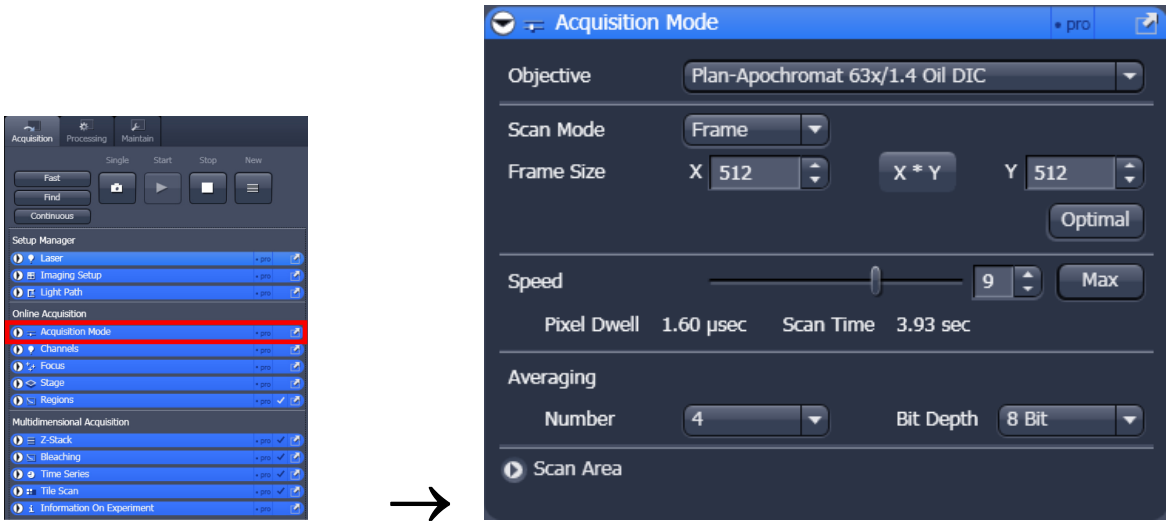


Fig. 18 **Acquisition Mode tool**

Note: When using an Axioskop 2 FS MOT, make sure you set the correct objective manually in the **Acquisition Mode** tool. This ensures correct calibration, calculation of pinhole and Z stack optimization, etc. The Axioskop 2 FS MOT does not automatically detect the objective lens.

Adjusting scan speed

- Use the **Scan Speed** slider in the **Acquisition Mode** tool (Fig. 18) to adjust the scan speed.

A higher speed with averaging results in the best signal-to-noise ratio. Scan speed 8 usually produces good results. Use speed 6 or 7 for superior images.

Choosing the dynamic range

- Select the dynamic range 8 or 12 Bit (per pixel) in the **Bit Depth** pull-down in the **Acquisition Mode** tool (Fig. 18).

8 Bit will give 256 gray levels; 12 Bit will give 4096 gray levels. Publication quality images should be acquired using 12 Bit data depth. 12 Bit is also recommended when doing quantitative measurements or when imaging low fluorescence intensities.

Setting scan averaging

Averaging improves the image by increasing the signal-to-noise ratio. Averaging scans can be carried out line-by-line or frame-by-frame. Frame averaging helps to reduce photo-bleaching, but does not give quite as smooth of an image.

- For averaging, select the **Line** or **Frame** mode in the **Acquisition Mode** tool.
- Select the number of lines or frames to average.

Adjusting pinhole size

- Select the **Channels** tool in the **Left Tool Area**.
- Set the **Pinhole** size to **1 AU** (Airy unit) for best compromise between depth discrimination and detection efficiency.

Pinhole adjustment changes the **Optical Slice thickness**. When collecting multi-channel images, adjust the pinholes so that each channel has the same **Optical Slice thickness**. This is important for colocalization studies.

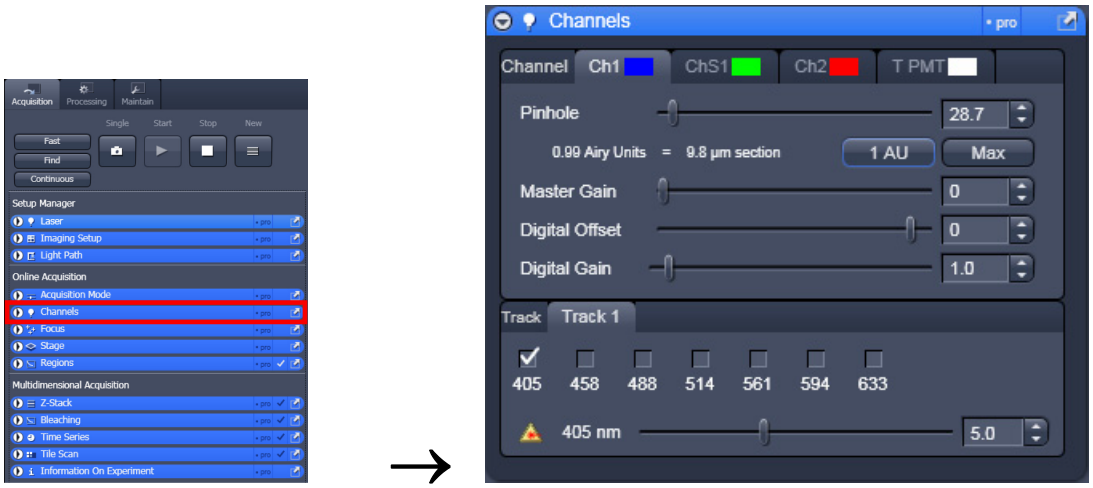


Fig. 19 **Channels tool**

Image acquisition

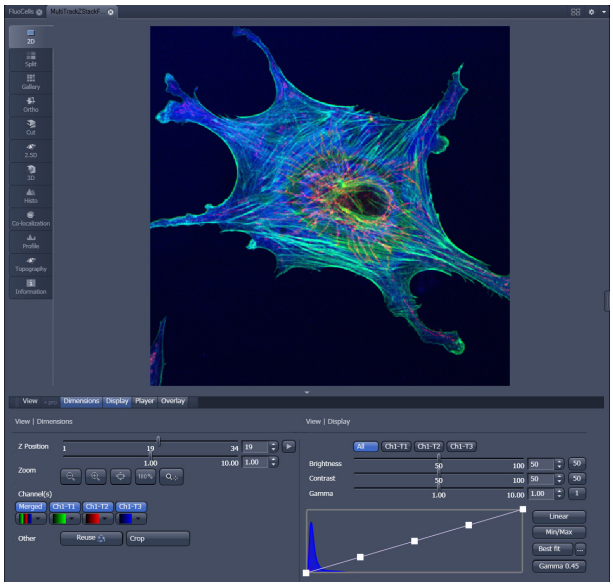


Fig. 20 **Image Display**

Once you have set up your parameter as defined in the above section, you can acquire a frame image of your specimen.

- Use one of the **Find**, **Fast**, **Continuous,** or **Single** buttons to start the scanning procedure to acquire an image.
- Scanned images are shown in separate windows.
- Click on the **Stop** button to stop the current scan procedure if necessary.

Select **Find** for automatic pre-adjustment of detector gain and offset.

Select **Fast** for continuous fast scanning – useful for finding and changing the focus.

Select **Continuous** for continuous scanning with the selected scan speed.

Select **Single** for recording a single image.

Select **Stop** for stopping the current scan procedure.

Image optimization

Fig. 21 **View Dimensions Control Block**

Choosing Range Indicator

- In the **View – Dimensions** View Option Control Block, click inside the color field in the 

button under the channel button (Fig. 21).
Note: Clicking on the right hand side of the 

button leads to a list of colors.

The scanned image appears in a false-color presentation (Fig. 22).

If the image is too bright, it appears red on the screen. Red = saturation (maximum).
If the image is not bright enough, it appears blue on the screen. Blue = zero (minimum).

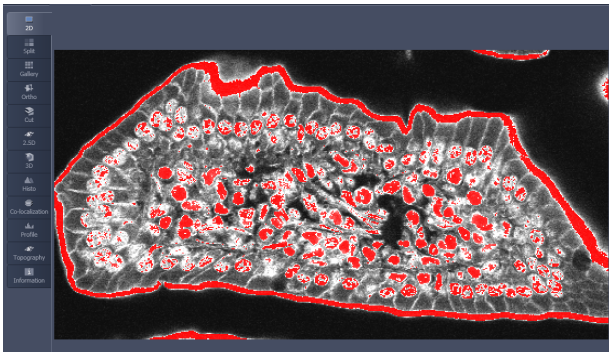


Fig. 22 Image Display

Adjusting the laser intensity

- Set the **Pinhole** to **1** Airy Unit (Fig. 23).
- Set the **Detector Gain** high.
- When the image is saturated, reduce AOTF transmission in the **Laser** control section of the **Channels Tool** (Fig. 23) using the slider to reduce the intensity of the laser light to the specimen.

Adjusting gain and offset

- Increase the **Digital Offset** until all blue pixels disappear, and then make it slightly positive (Fig. 23).
- Reduce the **Master Gain** until the red pixels only just disappear.

Fig. 23 Channels tool

Scanning a Z stack

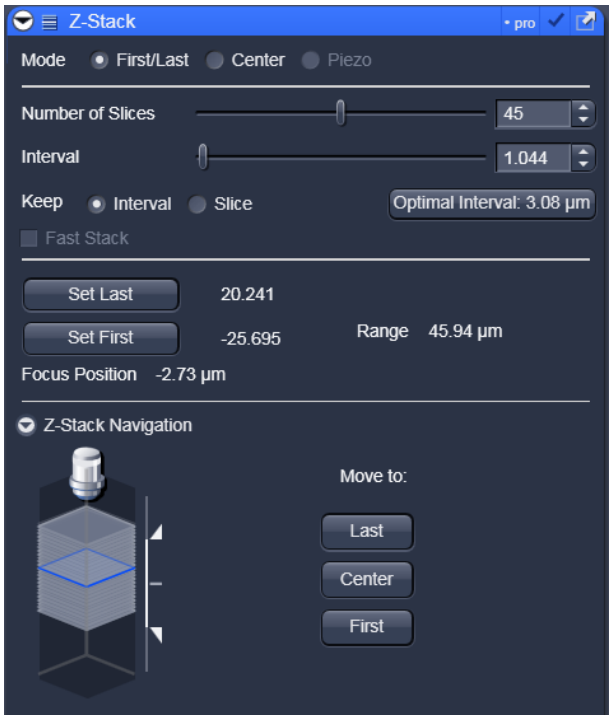


Fig. 24 **Z Stack tool**

- Open the **Z Stack** tool in the **Left Tool Area**.
- Select **Mode First/Last** on the top of the **Z Stack** tool.
- Click on the 

button in the **Action Button** area.

A continuous XY-scan of the set focus position will be performed.

- Use the focus drive of the microscope to focus on the upper position of the specimen area where the Z Stack is to start.
- Click on the **Set First** button to set the upper position of the Z Stack.
- Then focus on the lower specimen area where the recording of the Z Stack is to end.
- Click on the **Set Last** button to set this lower position.
- Click on the 

button to set number of slices to match the optimal Z-interval for the given stack size, objective lens, and the pinhole diameter.
- Make sure that the activation tickbox is set  .
- Click on the  **Start** button to start the recording of the Z Stack.

Note: When  is not activated, the respective tool and its set parameters are not included in the multidimensional image acquisition. If no multidimensional tool is activated, the  **Start** button is grayed out and only single images can be scanned.

Storing and exporting image data

- To save your acquired or processed images, click on the **Save** or **Save As** button in **File Menu** (Fig. 25/**1**), click on the  button at the bottom of the **File Handling Area** (Fig. 25/**2**) or click the  button in the **Main Toolbar** (Fig. 25/**3**).

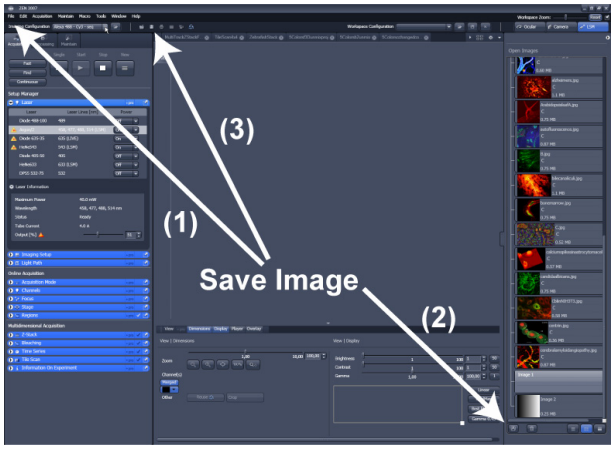


Fig. 25 **Save Image buttons in ZEN**

- The WINDOWS **Save As** window appears.
- Enter a file name and choose the appropriate image format. Note: the LSM 5 format is the native Carl Zeiss LSM image data format and contains all available extra information and hardware settings of your experiment.
- Click on the **SAVE** button.

If you close an image which has not been saved, a pop-up window will ask you if you want to save it. Choosing "yes" will lead you to the WINDOWS Save As window.

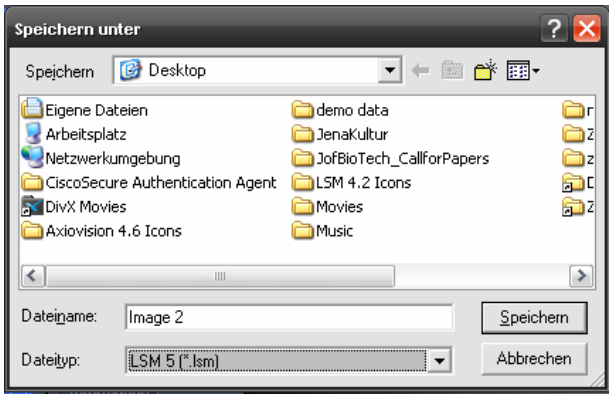


Fig. 26 **Save as window**

To export image display data, a single optical section in raw data format or the contents of the image display window including analysis and overlays, choose **Export** from the **File Menu**. In the **Export** window you can select from a number of options and proceed to the WINDOWS Save As window to save the exported data to disk.

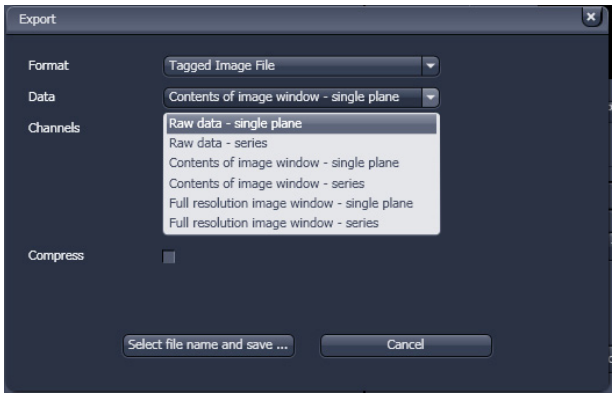
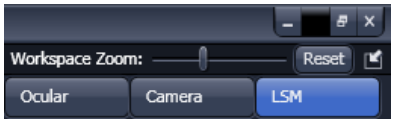


Fig. 27 **Export window**

Using the ConfoCor 3 module



- Click on the **LSM** button.
- Use the **ConfoCor** 3 Tool Group in the **Left Tool Area** to acquire and analyze **FCS** data.

Fig. 28 **ConfoCor 3 Tool Group**

Setting a configuration

- Open the **Measure** toolbar to access experiment parameter controls.

- Select the **System Configuration** controls in the **Measure** tool.

The **Light Path** and **Pinhole** panels of the **Measure** window display the selected track configuration which is used for the FCS procedure and the pinhole size (see Fig. 29).

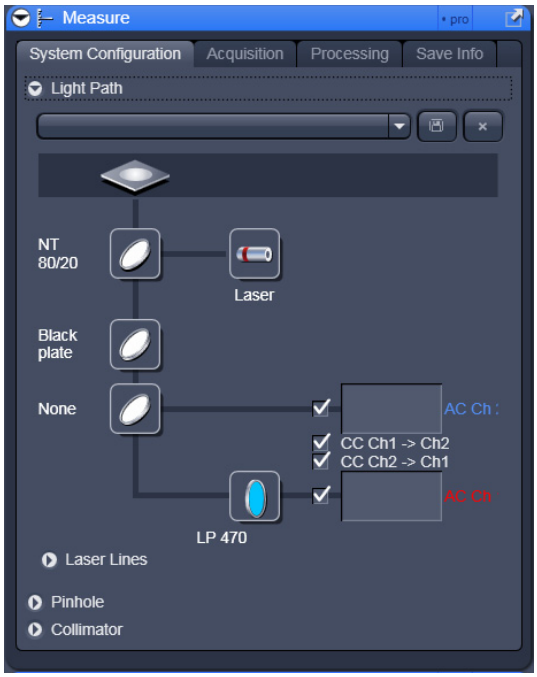


Fig. 29 **The ConfoCor 3 Measure Tool: System Configuration**

You can change the settings of this panel using the following function elements:

Activation / deactivation of the excitation wavelengths (check box) and setting of excitation intensities (slider). Open the **Laser Control** tool via the **Laser** icon.

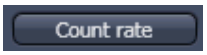
Selection of the main dichroic beam splitter (HFT) or secondary dichroic beam splitter (NFT) position through selection from the relevant list box.

Selection of a block filter through selection from the relevant list box.

Selection of an emission filter through selection from the relevant list box.

Activation / deactivation (via check box) of the selected channel.

Setting of the Pinhole diameter (via slider or input box).



Press the **Count rate** button to open the Real Time display window for the detector **Count rate** in all active channels. Adjust the Laser power In the **Laser Control** panel to obtain a satisfactory count rate.

Press **Adjust Pinhole** to align the pinhole for each newly defined beam path. After adjusting the sample carrier, align the pinhole in x and y by first conducting a coarse and then a fine alignment.

Starting a measurement

- Open the **Measure** toolbar to access experiment parameter controls**.**

- Select the **Acquisition** controls in the **Measure** tool.

The **Light Path** and **Pinhole** panels of the **Measure** window display the selected track configuration which is used for the FCS procedure and the pinhole size (see Fig. 30).

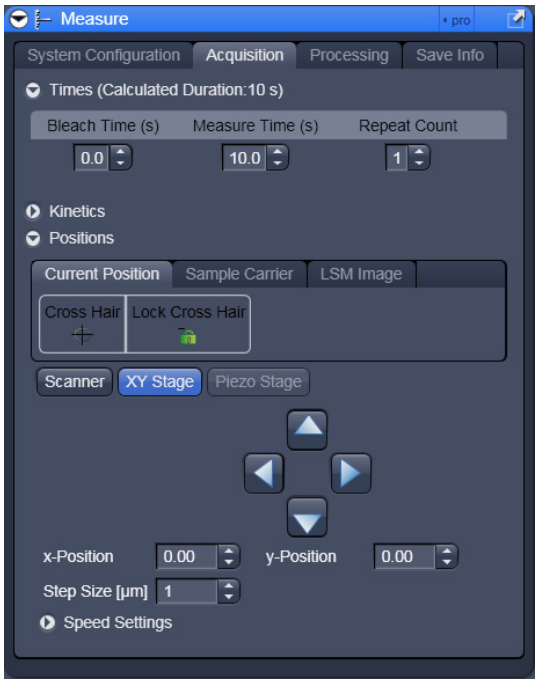


Fig. 30 **The ConfoCor 3 Measure Tool: Acquisition**

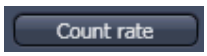
The **Times, Kinetics, and Position** panels of the **Acquisition** window display the selected measurement conditions and the positions which are used for the FCS experiment.

You can change the settings of this panel using the following function elements:

Enter the bleach time, measurement time, and repetition number in the corresponding input boxes.

Activate / deactivate a kinetic procedure by ticking the **Kinetics** check box. Enter the time distance between measurements and the cycle number in the corresponding input boxes.

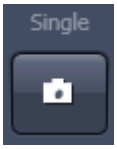
Select a sample carrier position and be sure that the focus is within the sample volume solution or location in an LSM image.



Press the **Count rate** button to open the Real Time display window for the detector **Count rate** in all active channels. This allows you to optimize your experiment by changing the laser power and the pinhole size while monitoring the count rate.

Press the **New** button to open a new **FCS diagram** into an image container. If a measurement is triggered, all data are displayed in that window if highlighted.

Press the **Start** button to trigger a measurement. All defined positions will be approached consecutively.

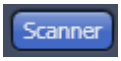


Press the **Single** button to trigger one measurement at the highlighted or first defined position.

Press the **Stop** button to end a measurement. All data accumulated so far will be available and can be stored.

Press the **xyz-Scan** button to display the current coordinates. You can define boundaries where a scan is performed with simultaneous acquisition of the count rate. This allows you, for example, to identify labeled molecules accumulated in the membrane.

Press **XY Stage** if positioning should be accomplished by the motorized stage.



Press **Scanner** if the positioning should be accomplished by the scanning mirrors

After measurement completion, the data is displayed in the FCS Correlation diagram within an Image Container (see Fig. 31)

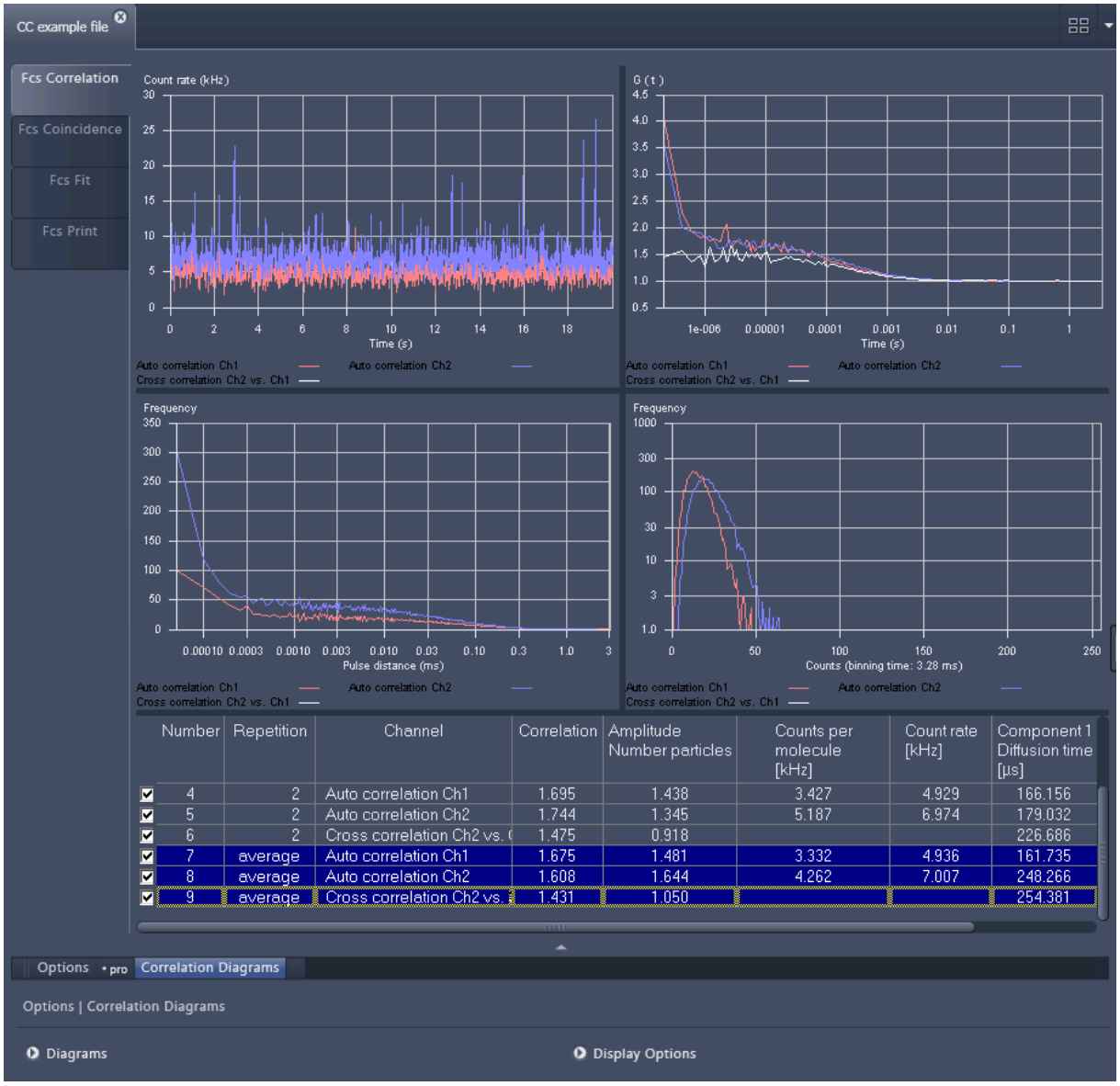
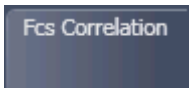
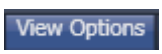


Fig. 31 **FCS Correlation diagram**

You have the following function elements:



Activate the **FCS Correlation** panel to display measured data (Fig. 31).

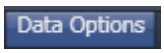


Press the **View Options** button to define the graph you want to display.

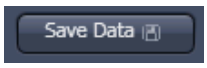
Press the **Count rate** button to display the count rate trace.

Press the **Correlation** button to display the correlation function.

Press the **Photon counting histogram** button to display the photon distribution per time unit.



Press the **Data Options** to handle your data.



Press the **Save Data** button to open the **Save** window. You can save the whole data set in an ANSI text format. Optionally you can save the raw data trace if that option was set in the FCS **Options**.

Pressing the **Reuse** button will set the system configuration to exactly the same values, as used in the experiment.

Pressing the **Reload** button will open the current measurement, if stored raw data are available. This allows you to alter the parameters of your mathematical calculations.

Analyzing the data

The acquired FCS data is analyzed in the Fit display of the FCS diagram (see figure Fig. 32).

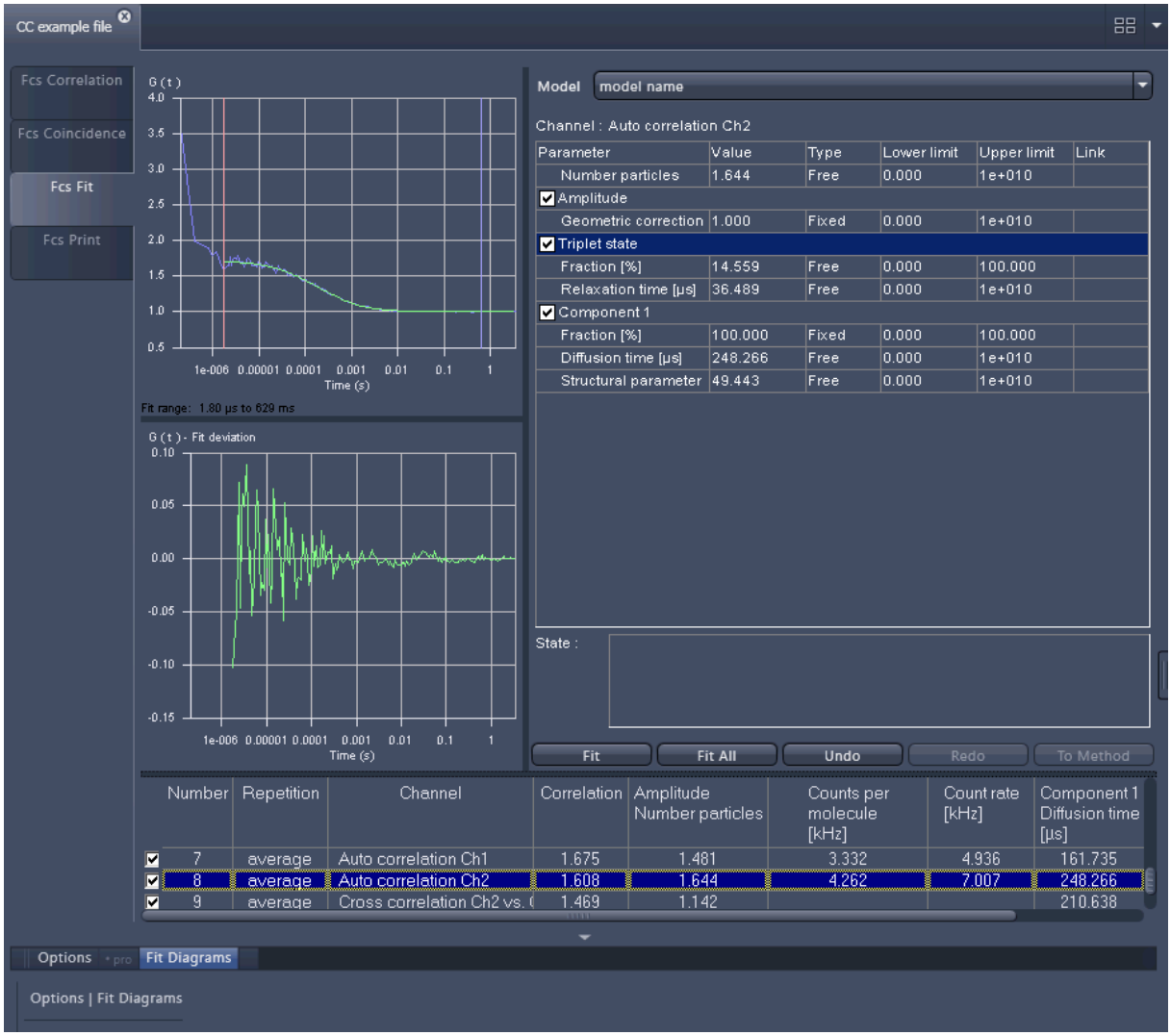
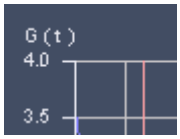


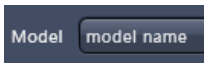
Fig. 32 **FCS Fit diagram**

You have the following options:

Activate the **FCS Fit** panel to display fitted data (Fig. 32).

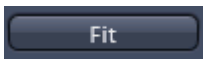


Set the red and blue bars to define the start and end points of the curve fit window.



Load a predefined model from the Model drop-down menu. You can assemble a model by pressing the **Model** tool in the **ConfoCor** tool group.

Define the conditions of the fit by activating / deactivating terms, setting the type of a parameter (fixed, free, or start value), defining limits and globally link parameters in the **Model** table.

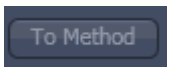


Pressing the **Fit** button will fit the current loaded correlation functions to the defined model. The fitted data will be displayed in the **Model** and **Result** tables.

Pressing the **Fit all** button will take all ticked channels and fit them according to the chosen model.

Pressing the **Undo** button will cancel the last operation, or previous ones as well, if the button is pressed repeatedly.

Pressing **Redo** will redo the last cancelled operation, or previous ones, if the button is pressed repeatedly.



Pressing the **Write to Method** button will write back the settings to the method. If the method is stored, the settings will be active when the method is selected the next time.

Switching off the system

- Click on the **File** button in the **Main Menu** bar and then click on the **Exit** button to leave the **ZEN 2008** software.
- If any lasers are still running you should shut them off now in the pop-up window indicating the lasers still in use.
- Shut down the computer.
- **Switch off the Ar-ML laser with 1st the standby switch (Fig. 2/3) and 2nd the main switch (Fig. 2/2), and wait until the fan of the Argon laser has switched off. Don't turn the key switch yet.**
- On the power remote switch turn off the **Components** switch and the **System/PC** switch (Fig. 1).
- Switch off the X-Cite 120 lamp or the HBO 100 mercury burner.
- Switch off the Ar-ML laser of by the main key switch on the power supply (Fig. 2/**1**).